International Journal of Chemical Studies

P-ISSN: 2349–8528 E-ISSN: 2321–4902 www.chemijournal.com IJCS 2021; SP-9(1): 346-349 © 2021 IJCS Received: 24-10-2020 Accepted: 30-12-2020

P Mayengbam

Associate Professor, Department of Veterinary Physiology and Biochemistry, College of Veterinary Sciences and Animal Husbandry, Central Agricultural University, Selesih, Aizawl, Mizoram, India

RC Upadhyay

Principal Scientist and Head Retired, Dairy Cattle Physiology Division, National Dairy Research Institute, Karnal, Haryana, India

TC Tolenkhomba

Assistant Professor (SG), Department of Animal Genetics and Breeding, College of Veterinary Sciences and Animal Husbandry, Central Agricultural University, Selesih, Aizawl, Mizoram, India

Corresponding Author: P Mayengbam

Associate Professor, Department of Veterinary Physiology and Biochemistry, College of Veterinary Sciences and Animal Husbandry, Central Agricultural University, Selesih, Aizawl, Mizoram, India

Neutrophil lymphocyte ratio of Karan Fries heifers in natural and artificial climatic condition

P Mayengbam, RC Upadhyay and TC Tolenkhomba

DOI: <u>https://doi.org/10.22271/chemi.2021.v9.i1f.11837</u>

Abstract

A study was carried out to find out the effect of temperature humidity index on neutrophil lymphocyte ratio of Karan-Fries heifers in five different periods based on maximum ambient temperature/minimum ambient temperature (Tmax/Tmin) *viz.* P1: <20 °C/<10 °C; P2: >20 °C/<10 °C, P3: >25 °C/<15 °C; P4: >35 °C/<20 °C and P5: >35 °C/>20 °C. The study investigated the effect of acute thermal exposure on neutrophil lymphocyte ratio of Karan-Fries heifers. Exposures were done at 40 °C and 50% RH and 45 °C and 50% RH in climatic chamber for 4 consecutive hours. Neutrophil leukocyte ratio was calculated from differential leukocyte count of blood smears stained with Leishman's stain. Neutrophil lymphocyte ratio of Karan-Fries. The study revealed good thermotolerance of Karan-Fries cattle both in natural climatic condition and thermal stress in climatic chamber.

Keywords: Cold, heat, Karan-Fries, neutrophil lymphocyte ratio, stress

Introduction

The haematological parameters have been a measure for detection and diagnosis of various diseases in animals. Factors contributing to variation in haematological parameters include age, sex, stress, diet, body condition, reproductive status, recent activity, hydration, ambient temperature, and altitude (Roland *et al.* 2014) ^[1]. Animals continuously experience some degree of physiological stress due to their exposure to inevitable stressors like extremes of temperature that may be in the form of heat during summer or cold during winter. A study of 2010 on comparison of reference intervals of healthy North American cows from 1957 to 2006 indicated significant increase in neutrophil count whereas reference intervals for lymphocyte, monocyte, and eosinophil counts as well as hemoglobin concentration had decreased (George *et al.*, 2006)^[2].

The neutrophil lymphocyte ratio in adult cattle of different breeds and origin is approximately 1:2 (Kinkaid 1999; Tornquist and Rigas, 2010; Wood *et al.*, 2010) ^[3-5]. In presence of stress, the leukogram of cattle indicated neutrophilia (Tornquist and Rigas, 2010) ^[3] and lymphopenia (Jones and Allison, 2007) ^[6]. Neutrophil lymphocyte ratio had been indicated to be a marker of acute (Swan and Hickman, 2014) ^[7] and chronic (Hickman, 2017) ^[8] stress in laboratory animals, several other types of stress in cattle (Lynch *et al.*, 2010) ^[9] and long-term stress in pigs (Sanchez *et al.*, 2019) ^[10]. However, there is no report so far on neutrophil lymphocyte ratio of Karan-Fries cattle both in natural climatic condition and in response to thermal exposures.

The present study was carried out to find out the effect of temperature humidity index on neutrophil lymphocyte ratio of Karan-Fries heifers. The study further investigated the effect of acute thermal exposure on neutrophil lymphocyte ratio of Karan-Fries heifers.

Materials and Methods

Rearing and maintenance of animals

A total of 30 Karan-Fries heifers were selected from the herd maintained at National Dairy Research Institute, Karnal (NDRI), Karnal. The animals were in the age group of 2-2.5 years and average body weight was 301.3±6.91kg. The animals were fed an ad lib roughage and water as per Kearl standard (Kearl, 1982)^[11].

Maintenance ration of concentrate mixture (12% CP and 60% TDN) consisting of mustard cake, maize, wheat bran, rice bran, mineral mixture and salt was fed @1kg/animal.

Study in natural climatic condition

Five different combinations of maximum ambient temperature (Tmax) and minimum ambient temperature (Tmin) were selected based on the climatograph prepared from the records of past 10 years. The conditions for five different temperature conditions (Tmax/Tmin) were P1: <20 °C/<10 °C; P2: >20 °C/<10 °C; P3: >25 °C/<15 °C; P4: >35 °C/<20 °C and P5: >35 °C/>20 °C.

Record of temperature and temperature humidity index

Prevailing temperature in Karnal as recorded at Central Soil Salinity Research Institute (CSSRI), Karnal were obtained. Everyday records of minimum ambient temperature, maximum ambient temperature, dry bulb and wet bulb temperature recorded at I: 0722\0830 and II: 1422 h IST were obtained for a period of 1 year. The temperature humidity index (THI) as proposed by National Research Council (National Research Council, 1971)^[12] was calculated by using dry bulb (db) and wet bulb (wb) temperatures in the following formula.

THI = 0.72(Tdb + Twb) + 40.6

Thermal exposure in climatic chamber

Karan-Fries heifers were kept in a climatic chamber (22'6''x10'10''x8') which was insulated and thermostatically fitted with heat convector for thermal exposure. Prior to initiation of thermal exposure animals were kept in the climatic chamber at the prevailing temperature for 4h every day for 10 days to accustom with the chamber environment. Thermal exposure was done at two intervals. Exposure I was done at $40\pm1^{\circ}$ C, 50% relative humidity (RH) during P3. Exposure II was done at $45\pm1^{\circ}$ C, 50% RH during P4. Thermal exposure was carried out for 4h continuously.

The THI during the thermal exposures were calculated (Johnson *et al.*, 1962) ^[13] by using ambient temperature (T) and relative humidity (H) in the the following formula.

$$\text{THI} = 0.08\text{T} + \text{RH}\frac{\text{T}-14.4}{100} + 46.4$$

Blood sampling

Blood samples were collected from each of animals in natural climatic condition in 5 different periods. Blood samples were also collected from the animals in the thermal exposure at 0h, 1h, 2h, 3h and 4h of thermal exposure. A few drops of blood samples were collected from jugular vein by using Di-sodium salt of EDTA coated vacutainer tubes with needles.

Calculation of neutrophil lymphocyte ratio

A small drop of freshly collected blood was placed in the central line of a slide about 1-2 cm, from one end. A spreader slide with a smooth edge was placed at an angle of 45° onto the slide and then moved back to the blood drop. The drop was spread out along the line of contact of the spreader slide. The blood film was dried by waving the slide once in the air. Leishman stain (0.15% in methyl alcohol) was poured enough on the smear to cover it fully and allowed to act for 2 min. Distilled water was added twice the amount of stain and allowed to mix thoroughly by blowing air onto the mixture. Staining was done for 10 min. The smear was washed in the running tap water until colour stops draining. It was allowed

to dry in air by keeping the slide in inverted position with the broad end of the film up.

Differential leukocyte count (DLC) was done by using oil emersion objective at 100x of an inverted binocular light microscope in a strip running the whole length of the film. The lateral edges of the film were avoided. The film was inspected from head to the tail. Approximately more than 200 cells were counted from one slide. DLC were expressed in %. From the result of DLC neutrophil lymphocyte ratio was calculated.

Statistical Analysis

The data were analyzed by using SPSS 16 version. One way analysis of variance was carried to find out the difference between different periods and between different hours of thermal exposure (Snedecor and Cochran, 1989)^[14].

Ethical Approval for Animal Experimentation

Ethical approval was obtained from Institutional Animal Ethical Committee for thermal exposure and blood sampling.

Results ad Discussion

Average ambient temperatures and THI during different temperature conditions and thermal exposures in climatic chamber have been presented in Table 1 and 2 respectively. The neutrophil lymphocyte ratio of Karan-Fries heifers in natural climatic condition and thermal exposure have been presented in Table 3 and 4 respectively.

Ambient temperature and THI

The lowest Tmin and lowest average ambient temperature (Tav) were recorded in P1. The highest Tmax and the highest Tav were recorded in P5. Heat stress in dairy cattle starts at THI of 72 which corresponds to 22°C at 100% humidity, 25°C at 50% humidity, or 28°C at 20% humidity (Johnson et al., 1963) ^[15]. In the present study, average THI recorded in P4 and P5 were higher than 72 indicating presence of heat stress. THI >72 was found to cause thermal stress in Sahiwal and Karan-Fries in the form of changes in physiological indices (Mayengbamm et al., 2015; Mayengbam et al., 2016) [16-17]. Experience of thermal stress was also evident in Sahiwal and Karan -Fries cattle during THI >72 by presence of increase in expression of heat stress markers (HSP70) in Sahiwal and Karan-Fries cattle (Mayengbam et al., 2016) [17] and antioxidant enzyme genes (Cu-SOD and Mn-SOD) in Sahiwal and Karan-Fries cattle (Mayengbamm et al., 2015)^[16].

 Table 1: Ambient temperature and temperature humidity index in different periods

Parameters		Periods						
		P1	P2	P3	P4	P5		
Т	Max	16.70	20.40	30.80	36.00	37.79		
	Min	2.50	9.00	13.60	17.80	23.16		
	Average	9.60	14.70	22.20	26.90	30.48		
T _{db} (°C)	Max	18.40	19.80	30.50	36.20	35.73		
	Min	3.60	10.40	14.90	20.50	27.00		
	Average	11.00	15.10	22.70	28.35	31.37		
T _{wb} (°C)	Max	11.40	14.80	20.10	20.70	24.45		
	Min	2.80	9.50	14.30	17.40	22.98		
	Average	7.10	12.15	17.20	19.05	23.72		
THI	Max	62.05	65.51	77.01	81.51	83.93		
	Min	45.21	54.98	61.62	67.86	76.58		
	Average	53.63	60.25	69.32	74.69	80.26		

The THI recorded inside the climatic chamber during thermal Exposure I and II were below the stress level as compared to

previous records (Johnson *et al.*, 1963) ^[15]. However, acute thermal exposure at such THI levels were found to cause thermal stress in Sahiwal and Karan-Fries cattle and the same

was detected by presence of higher expression of heat stress marker HSP70 mRNA (Mayengbam and Upadhyay, 2014)^[18] and HSP70 protein (Mayengbam *et al.*, 2016)^[17].

Table 2: Ambient temperature and temperature humidity index during thermal exposures

E	Climatic chamber		Environment						
Exposure	T (°C)	RH (%)	THI	Tmax	Tmin	Tav (°C)	Tdb (°C)	Twb (°C)	THI
Exposure I	40±1	50	62.4	30.80	13.60	22.20	22.70	17.20	69.32
Exposure II	45±1	50	65.3	36.00	17.80	26.90	28.35	19.05	74.69

Neutrophil lymphocyte ratio

During different periods (P1-P5) neutrophil lymphocyte ratio of Karan-Fries ranged from 0.41±0.02 to 0.48±0.02 which was well within the normal range (Kinkaid 1999; Tornquist and Rigas, 2010; Wood et al., 2010) ^[3-5]. There was no significant change in neutrophil lymphocyte ratio of Karan-Fries during different periods. In Holando Argentino cows, neutrophil lymphocyte ratio was found to increase in summer (Tolini et al., 1917)^[19]. During P4 and P5 of present study, THI as high as 74.69 and 80.26 respectively were recorded in Karnal. The present study found no effects of high THI on neutrophil lymphocyte ratio. There was however increase in expression of heat stress marker HSP70 (Mayengbam et al., 2016) [20] and HSP70 mRNA in response to THI>72 in Karan-Fries (Mayengbam et al., 2016) ^[17]. Karan-Fries maintained stable neutrophil lymphocyte ratio throughout the year even during the periods of extremes of temperature in winter and summer. The present finding indicated better adaptability of Karan-Fries to extremes of climatic conditions.

 Table 3: Neutrophil lymphocyte ratio (mean± SEM) of Karan-Fries

 heifer during different periods

P1	P2	P3	P4	P5
0.44	0.43	0.42	0.41	0.48
±0.02	±0.03	±0.03	±0.02	±0.02

When Karan-Fries heifers were subjected to acute thermal stress in Exposure I with 40°C and 50% RH and Exposure II with 45°C and 50% RH for 4 consecutive hours and neutrophil lymphocyte ratio was estimated there was no significant change in neutrophil lymphocyte ratio. In cattle, several types of stress viz. post parturient stress (Kulberg et al., 2002)^[21], exercise stress (Garcia-Belenquer et al., 1996) ^[22] abrupt weaning (Lynch et al., 2010) ^[9] and sole ulcer (O'Driscoll et al., 2015)^[23] were found to increase neutrophil lymphocyte ratio. The present finding indicated that Karan-Fries maintained stable neutrophil lymphocyte ratio in presence of acute thermal stress. There were however reports that Karan-Fries cattle exhibited higher expression of heat stress marker (HSP70) in response to acute thermal stress (Mayengbam and Upadhyay, 2014 16) and antioxidant stress marker viz. Mn-SOD and Cu-SOD (Mayengbam et al., 2015 15). Presence of stable neutrophil lymphocyte ratio in response to acute thermal stress indicated high adaptability of Karan-Fries to acute thermal stress.

 Table 4: Neutrophil lymphocyte ratio (mean± SEM) of Karan-Fries

 heifer during thermal exposure

Exposure	0h	1h	2h	3h	4h
Ι	0.41 ± 0.05	0.41 ± 0.00	0.45 ± 0.02	0.45 ± 0.01	0.47 ± 0.01
II	0.43 ± 0.03	0.47 ± 0.01	0.40 ± 0.02	0.42 ± 0.02	0.46 ± 0.01

Conclusion

The study found good thermotolerance of Karan-Fries cattle in winter as well as summer. Karan-Fries heifers maintained a stable neutrophil lymphocyte ratio in natural climatic condition even in the extremes of cold and hot in winter and summer. Acute thermal exposure caused no significant change in neutrophil lymphocyte ratio of Karan-Fries heifers.

Acknowledgement

Authors are grateful to ICAR Network Project on 'Impact, Adaptation of Vulnerability of Indian Agriculture to Climate Change' for financial support.

References

- Roland L, Drillich M, Iwersen M. Hematology as a diagnostic tool in bovine medicine. Journal of Veterinary Diagnostic Investigation 2014;26(5):592-598.
- 2. George JW, Snipes J, Lane VM. Comparison of bovine hematology reference intervals from 1957 to 2006. Veterinary Clinical Pathology 2010;39:138-148.
- Kincaid RL. Assessment of trace mineral status of ruminants: A review. Journal of Animal Science 1999;77(E Suppl):1-10.
- 4. Tornquist SJ, Rigas J. Interpretation of ruminant leukocyte responses. In: Schalm's veterinary hematology, ed. Weiss DJ and Wardrop KJ. 6th ed., Wiley, Ames, IA 2010, 307-313.
- Wood D, Quiroz-Rocha GF. Normal hematology of cattle. In: Schalm's veterinary hematology, ed. Weiss DJ and Wardrop KJ. 6th ed., Wiley, Ames, IA 2010, 829-835.
- 6. Jones ML, Allison RW. Evaluation of the ruminant complete blood cell count. Veterinary Clinics of North America: Food Animal Practice 2007;23:377-402.
- Swan MP, Hickman. Evaluation of the neutrophillymphocyte ratio as a measure of distress in rats. Lab Animal (NY) 2014;43(8):276-282. Doi: 10.1038/laban.529.
- Hickman DL. Evaluation of the neutrophil: lymphocyte ratio as an indicator of chronic distress in the laboratory mouse. Lab Animal (NY) 2017;46:303-307. doi:10.1038/laban.1298.
- Lynch EM, Earley B, McGee M, Doyle S. Characterisation of physiological and immunological responses in beef cows to abrupt weaning and subsequent housing. BMC Veterinary Research 2010;6:37 http://www.biomedcentral.com/1746-6148/6/37
- Sanchez NCB, Carroll JA, Corley JR, Broadway PR, Callaway TR. Changes in the hematological variables in pigs supplemented with yeast cell wall in response to a Salmonella challenge in weaned pigs. Frontier in Veterinary Science 2019. https://doi.org/10.3389/fvets.2019.00246

- 11. Kearl LC. Nutrient Requirements of Ruminants in Developing Countries. International Feedstuffs Institute, Utah Agricultural Experiment Station, Utah State University Logan, Utah USA 1982.
- 12. National Research Council. A Guide to Environmental Research on Animals. National Academy of Science, Washington DC 1971.
- 13. Johnson HD, Rangsdale AC, Berry IL, Sanklin MD. Effect of various temperature humidity combinations on milk production of Holstein cattle. Missouri Agricultural Experiment Station Research Bulletin 1962, 791.
- Snedecor GW, Cochran WG. Statistical Methods. 8th Edn. Affiliated East-West Press Pvt. Ltd., New Delhi 1989.
- 15. Johnson HD, Ragsdale AC, Berry IL, Shanklin MD. Temperature-humidity effects including influence of acclimation in feed and water consumption of Holstein cattle. Missouri Agricultural Experiment Station Research Bulletin 1963, 846.
- 16. Mayengbam P, Tolenkhomba TC, Upadhyay RC. Mn-SOD and Cu, Zn-SOD mRNA expression in relation to physiological indices of Sahiwal and Karan-Fries heifers during different temperature humidity indices. Journal of Agrometeorology 2015;17(2):172-178.
- 17. Mayengbam P, Tolenkhomba TC, Upadhyay RC. Expression of heat-shock protein 72 mRNA in relation to heart rate variability of Sahiwal and Karan-Fries in different temperature-humidity indices Veterinary World 2016;9(10):1051-1055.
- Mayengbam P, Upadhyay RC. Heat Shock Protein 72 expression of Sahiwal and Karan-Fries during thermal stress. Indian Journal of Dairy Science 2014;67(2):147-153.
- 19. Tolini F, Fernandes G, Mayer N, Maiztegui L, Munoz Griselda, Pagni C *et al.* Influence of heat stress on white blood cells in Holando Argentino cows. Revista Agronomica del Noroeste Argentino 2017;37(1):67-75.
- 20. Mayengbam P, Upadhyay RC, Tolenkhomba TC. Seasonal variation in HSP72 expression of Sahiwal and Karan-Fries cattle. Indian Veterinary Journal 2015;92(4):34-36.
- 21. Kulberg S, Storset AK, Heringstad B, Larsen HJS. Reduced levels of total leukocytes and neutrophils in Norwegian cattle selected for decreased mastitis incidence. Journal of Dairy Science 2002;85:3470-3475.
- 22. Garcia-Belenquer S, Gascon JP, Acena C, Revilla R, Mormede P. Differences in the biological stress responses of two cattle breeds to walking up to mountain pastures in the Pyrenees Veterinary Research 1996;27(4-5):515-526.
- 23. O'Driscoll K, McCabe M, Earley B. Differences in leukocyte profile, gene expression, and metabolite status of dairy cows with or without sole ulcers. Journal of Dairy Science 2015;98:1685-1695.