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## *In vitro* evaluation of the chemical fungicides against the fungus *Botrytis ricini*

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**Abstract**

Castor (*Ricinus communis* L.) is an important non-edible oilseed crop which experienced serious declines in area under cultivation in Telangana state due to yield losses caused by gray mold pathogen *Botryotinia ricini*. *B. ricini* fungus were isolated from infected castor plants in ICAR-IIOR, Hyderabad and maintained in oat meal gar medium. Use of chemical fungicides is an important tool the management of gray mold disease in castor. Five systemic fungicides, Bavistin (Carbendazim), Score (Difenoconazole), Amistar (Azoxystrobin), Ergon (Kresoxim methy) and three combination formulation fungicides such as Merivon (Fluxapyroxad + Pyraclostrobin), Cabrio Top (Pyraclostrobin + Metiram) and Sectin (Fenamidone + Mancozeb) were evaluated for *in vitro* efficacy at concentrations of 100, 250, 500, 750 and 1000 ppm against gray mold pathogen. The poisoned food technique was used and radial mycelial growths were measured seven day after inoculation. Bavistin was found to be the most effective fungicide followed by Merivon have shown high fungitoxicity against *B. ricini*. Amistar were the least effective. The results of this present investigation will be helpful for future research on botrytis fungicide management and for castor growers in controlling gray mold disease of castor.

**Keywords:** Castor, gray mold, fungicides

**Introduction**

Castor, *Ricinus communis* L., is important non-edible oilseed crop belong to the family Euphorbiaceae. It is widely cultivated in the tropics and sub tropics regions for its seed from which 50-55% castor oil is extracted (Weiss, 1973; Purseglove, 1968; Gobin et al., 2001; Adefris and Nigussie, 1993) [21, 17, 7, 1]. Castor oil is used in industries (Cosmetic base, lubricant, paints) and pharmaceuticals (Gobin et al., 2001 and Purseglove, 1968) [7, 17]. Among the various diseases in castor, gray mold disease caused by *Botryotinia ricini* Godfrey, emerged as a serious destructive disease especially in Telangana and Andhra Pradesh state when maturity of the racemes coincides with the cyclonic rains. Successful control could not be achieved causing great damage under favourable weather conditions leads to huge yield loss. Anjani et al., (2004) [3] reported that botrytis fungus can cause yield loss of 80-100% and this affects the income of castor farmers in terms of foreign exchange.

Very little information is available regarding its management of gray mold disease of castor. Some workers with varied success have used different chemicals and fungicides. *In vitro* studies conducted reveals that some fungicides restrict or prevent the growth of *B. ricini* pathogens (Yamuna, 2015; Gurvinder Kaur, 2015; Suyl et al., 2011) [22, 9, 20]. Fungicides like tebuconazole and iprodione are being recommended (Chagas et al., 2014) [5] and there is still a need to identify a better fungicide to control *Amphobotrys ricini* infection on castor. In view of the economic importance of the crop and the effect of gray mold diseases on the seed yield, there is a need to recommend specific fungicidal management for disease under field conditions. The aim of the study was to determine *in vitro* effects of chemical fungicides on the mycelial growth of *B. ricini* pathogen isolated from castor raceme.

**Materials and Methods****Isolation and maintenance of the pathogen**

Isolation of the test fungal pathogen was made from gray mold infected capsules collected from the ICAR-IIOR, Rajendranagar, Hyderabad. The infected capsules were cut into small pieces and were surface sterilized by immersing in 0.1% mercuric chloride for 30 seconds and

placed on oat meal enriched medium containing petri plates after thorough washings in sterile distilled water. The plates were incubated at 23±1°C temperature for 3 to 4 days. The fungal growth emerging from diseased capsule pieces were picked up and the culture was further purified by single spore isolation method. The pure culture of the pathogen was maintained on oat meal enriched medium by periodical transfers (Prasad and Bhuvaneswari *et al.*, 2014) [16].

### Identification of the pathogen

The fungus associated with the disease was brought into pure culture on Oat meal enriched medium and identified based on the descriptions given by Stevens (1918) [19], Godfrey (1923) [8] and Moses and Reddy (1989) [15].

### Screening of fungicides against *B. ricini* under *in vitro* conditions

The laboratory experiment was conducted at the ICAR-IIOR, Hyderabad during 2018 on gray mold disease of castor. The study was conducted in a Complete Randomized Design (CRD) with seven treatments and four replications. Seven different fungicides belonging to different groups/ mode of action (Table 1) such as Bavistin (Carbendazim), Score (Difenoconazole), Amistar (Azoxystrobin), Ergon (Kresoxim methy) and three combination formulation fungicides *viz.*, Merivon (Fluxapyroxad + Pyraclostrobin), Cabrio Top (Pyraclostrobin +Metiram) and Sectin (Fenamidone + Mancozeb) were screened against the *B. ricini* under laboratory conditions to find out their relative efficacy in inhibiting the mycelial growth of the pathogen in culture media by the "Poisoned food technique" (Schmitz, 1930) [18] at 100, 250, 500, 750, 1000 ppm concentrations respectively. Oat meal agar medium was prepared and amended with different concentrations of the fungicides. About 20 ml of sterilized culture medium was poured in each 9 cm petri dish. After solidification, the plates were inoculated with a 5 mm disk of seven days-old *B. ricini* culture. Four replications were maintained for each concentration and radial growth was recorded. The Petri plates were incubated at 23±1°C. One set of control was maintained without adding any fungicide to the medium. The observations were recorded until the control plate was full of growth of the pathogen. The data of radial growth of fungal colony was measured in millimeters. The per cent mycelial inhibition over control was calculated by the following formula given by Bliss (1934) [4].

$$\text{Per cent inhibition over control} = [(C-T)/T] \times 100$$

Where, C = Growth of fungus in control; T = Growth of fungus in treatment

### Results and Discussion

The efficacy of seven fungicides were tested under *in vitro* by poisoned food technique and the results are presented in Table 2 and 3. Most of the fungicides were effective in inhibiting the mycelial growth of the *B. ricini* pathogen to varying degrees. Significant difference in inhibiting the mycelial growth of the pathogen among the fungicides was observed.

Out of four systemic fungicides tested, complete mycelial inhibition (100%) of growth of the pathogen over control was observed with Bavistin (Carbendazim) followed by Score (Difenoconazole) at 100 ppm concentration when compared to control. Amistar (Azoxystrobin) was least effective with 16.6 per cent mycelial growth inhibition at 100 ppm concentration (Table 2). Similarly, results of combination fungicide reveals that Merivon (Pyraclostrobin + Metiram) shown complete inhibition (100%) of growth of the pathogen over control followed by sectin (Fenamidone + Mancozeb) with mean per cent inhibition of 87.5 per cent whereas CabrioTop (Pyraclostrobin+ Metiram) was least effective with 78.3 per cent in inhibiting the growth of the pathogen at 100 ppm concentration (Table 3).

The benzimidazole fungicides were the first systemic fungicides that exhibited broad spectrum of activity against *Botrytis* spp. (Delp, 1995) [6]. These fungicides do not prevent conidial germination, but at low concentrations, they inhibit hyphal growth and cause distortion of germ tubes (Leroux *et al.*, 1999) [13]. Carbendazim fungicide can inhibits the development of fungi by interacting with  $\beta$ -tubulin and stopping hyphal growth and cell division (Hollomon *et al.*, 1998) [10]. Similar *in vitro* results were obtained by Yamuna (2015) [22] that fungicides such as Bavistin (carbendazim), Folicur (tebuconazole), Tilt (propiconazole) and Nativo (trifloxystrobin + tebuconazole) showed 100% mycelial inhibition of the *B. ricini* pathogen at 250 and 500 ppm concentrations by poison food technique. The laboratory data revealed that Quintal and Contaf at 50, 100, 150 and 200 ppm showed complete mycelial growth inhibition of *Botrytis cinerea* under *in vitro* condition followed by Avtar (91.16%) and Score (63.99%), respectively whereas Hilate was least effective (11.68%) (Gurvinder Kaur, 2015) [9]. Hosen *et al.* (2010) [12] and Hosen (2011) [11] concluded that the iprodione and carbendazim was effective in inhibition of spore germination *B. cinerea*. The reduction in disease severity by iprodione + carbendazim, difenaconazole and hexaconazole are in consonance with the findings of Madhu Meeta *et al.* (1986) [14] and Agarwal and Tripathi (1999) [2]. Suyl *et al.* (2011) [20] tested seven fungicides under *in vitro* for the control of grey mold of chickpea. Fungicides Bavistin (0.1%), Quintal (0.1%) and Derosal Plus (0.3%) showed complete mycelial inhibition of *B. cinerea* in chickpea.

**Table 1:** Details of fungicides employed in the present investigation

| Trade Name | Fungicide name                | IUPAC name  | Formulation * | Source of Supply     |
|------------|-------------------------------|---|---------------|----------------------|
| Bavistin   | Carbendazim                   | Methyl (1H-benzimidazol-2-yl)carbamate  | 50 WP         | BASF                 |
| Score      | Difenoconazole                | 1-((2-(2-Chloro-4-(4-chlorophenoxy)phenyl)-4-methyl-1,3-dioxolan-2-yl)methyl)-1H-1,2,4-triazole   | 25 EC         | Syngenta             |
| Amistar    | Azoxystrobin                  | Methyl (2E)-2-{2-[6-(2-cyanophenoxy) pyrimidin-4-yloxy]phenyl}-3-methoxyacrylate  | 25 SC         | Syngenta             |
| Ergon      | Kresoxim methy                | (2E)-(methoxyimino){2-[(2-methylphenoxy) methyl] phenyl}acetic acid   | 44.3 SC       | Rallis India Limited |
| Merivon    | Fluxapyroxad + Pyraclostrobin | 1H-Pyrazole-4-carboxamide, 3-(difluoromethyl)- 1-methyl-N-(3',4',5'-trifluoro[1,1'-biphenyl]-2-yl) + [2-[[[1-(4-chlorophenyl)- 1H-pyrazol-3-yl]oxy] methyl] phenyl] methoxy-, methyl ester            | 25 SC         | BASF                 |
| Cabrio Top | Pyraclostrobin + Metiram      | Methyl[2-({[1-(4-chlorophenyl)-1H-pyrazol-3yl] oxy} methyl) phenyl] methoxycarbamate Tris [ammine [ethylenebis (dithiocarbamato)]zinc(2+)] [tetrahydro-1,2,4,7-dithiadiazocine-3,8 dithione], polymer | 20 WG         | BASF                 |

|        |                       |  |       |                    |
|--------|-----------------------|--|-------|--------------------|
| Sectin | Fenamidone + mancozeb | 5S)-5-Methyl-2-(methylsulfanyl)-5-phenyl-3-(phenylamino)-3,5-dihydro-4H-imidazole-4-one+ Manganese(2+) zinc 1,2-ethanediyldicarbamodithioate | 60 WG | Bayer Crop Science |
|--------|-----------------------|--|-------|--------------------|

\*WP - Wettable Powder; EC- Emulsifiable Concentrate; SC- Suspension Concentrate; WG- Wettable Granules

**Table 2:** Radial colony growth along with %inhibition of *B. ricini* with systemic fungicides

| Concentration (ppm) | Fungicides*        |                |                    |                |                    |                |                    |                |
|---------------------|--------------------|----------------|--------------------|----------------|--------------------|----------------|--------------------|----------------|
|                     | Bavistin           |                | Score              |                | Amistar            |                | Ergon              |                |
|                     | Radial growth (mm) | Inhibition (%) |
| 100                 | 0 (0.0)            | 100            | 6.5 (14.5)         | 92.7           | 75 (60.3)          | 16.6           | 51 (45.5)          | 43.3           |
| 250                 | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 57 (49.0)          | 38.1           | 0 (0.0)            | 100            |
| 500                 | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 19.5 (26.1)        | 82.7           | 0 (0.0)            | 100            |
| 750                 | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 7.5 (15.7)         | 98.3           | 0 (0.0)            | 100            |
| 1000                | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 0 (0.0)            | 100            |
| Control (Untreated) | 90 (100)           | 0.0            | 90 (100)           | 0.0            | 90 (100)           | 0.0            | 90 (100)           | 0.0            |
| Factors             | SE(d)              |                |                    |                | C.D.               |                |                    |                |
| Fungicides (A)      | 0.56               |                |                    |                | 1.11               |                |                    |                |
| Concentrations (B)  | 0.62               |                |                    |                | 1.25               |                |                    |                |
| Factor(A X B)       | 1.25               |                |                    |                | 2.49               |                |                    |                |

\*Mean of four replications, () Data in parentheses are arcsine transformed values

**Table 3:** Radial colony growth along with %inhibition of *B. ricini* with combination fungicides

| Concentration (ppm) | Fungicides*        |                |                    |                |                    |                |
|---------------------|--------------------|----------------|--------------------|----------------|--------------------|----------------|
|                     | Merivon            |                | Cabrio Top         |                | Sectin             |                |
|                     | Radial growth (mm) | Inhibition (%) | Radial growth (mm) | Inhibition (%) | Radial growth (mm) | Inhibition (%) |
| 100                 | 0 (0.0)            | 100            | 19.5 (26.1)        | 78.3           | 11 (19.2)          | 87.7           |
| 250                 | 0 (0.0)            | 100            | 4.0 (11.4)         | 95.5           | 0 (0.0)            | 100            |
| 500                 | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 0 (0.0)            | 100            |
| 750                 | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 0 (0.0)            | 100            |
| 1000                | 0 (0.0)            | 100            | 0 (0.0)            | 100            | 0 (0.0)            | 100            |
| Control (Untreated) | 90 (100)           | 0.0            | 90 (100)           | 0.0            | 90 (100)           | 0.0            |
| Factors             | SE(d)              |                |                    | C.D.           |                    |                |
| Fungicides (A)      | 0.25               |                |                    | 0.51           |                    |                |
| Concentrations (B)  | 0.33               |                |                    | 0.66           |                    |                |
| Factor(A X B)       | 0.57               |                |                    | 1.14           |                    |                |

\*Mean of four replications, () Data in parentheses are arcsine transformed values

## Conclusion

Bavistin (50 WP) systemic fungicide showed the complete mycelial inhibition of *B. ricini* pathogen at 100 ppm using poison food technique. So, Bavistin 50 WP may be suggested to use by the castor grower for the management of castor gray mold disease.

## References

- Adefris T, Nigussie A. Castor in Ethiopia: Production, Utilization and research. Oilcrops Newsletter. 1993; 10:40-43.
- Agarwal A, Tripathi HS. Biological and chemical control of botrytis grey mould of chickpea. Journal of Mycology and Plant Pathology. 1999; 29:52-56.
- Anjani K, Raoof MA, Ashoka P, Reddy V, Hanumanta Rao C. Sources of resistance to major castor (*Ricinus communis*) diseases. Plant Genetic Resources Newsletter. 2004; 137:46-48.
- Bliss C. The methods of probits. Science. 1934; 79:38.
- Chagas HA, Basseto MA, Rosa DD, Toppa EVB, Furtado EL, Zanotto MD. Evaluation of fungicides, essential oils and biological agents on *Amphobotrys ricini* control in castor bean (*Ricinus communis* L.). Summa Phytopathologica. 2014; 40(1):42-48.
- Delp CJ. Benzimidazole and related fungicides. In: Lyr H (ed.) Modern Selective Fungicides. Gustav Fisher Verlag, Jena, Germany, 1995, 291-303.
- Gobin AML, Uguru MI, Deckers J, Castor. In: Crop Production in Tropical Africa. Romain H. Reamaekers (ed), 2001, 725-733.
- Godfrey GH. Gray mold of Castor bean. Journal of Agricultural Research. 1923; 23(9):679-715.
- Gurvinder Kaur. Variability and management studies of *Botrytis cinerea* causing grey mould in gladiolus. Dissertation. Dr Yashwant Singh Parmar University of Horticulture and Forestry, Nauni, Solan, 2015, 1-200.
- Hollomon DW, Butters JA, Barker H, Hall L. Fungal beta-tubulin, expressed as a fusion protein, binds benzimidazole and phenylcarbamate fungicides. Antimicrobial Agents and Chemotherapy. 1998; 42:2171-2173.
- Hosen MI. Cultural, physiological comparison and fungicidal sensitivity between two isolates of *Botrytis cinerea* and *Stemphylium botryosum*. Emirates Journal of Food and Agriculture. 2011; 23:120-29.
- Hosen MI. Physiological variability and *in vitro* antifungal activity against *Botrytis cinerea* causing botrytis gray mold of chickpea (*Cicer arietinum* L.). Spanish Journal of Agricultural Research. 2010; 8:750-856.
- Leroux P, Chapeland F, Desbrosses D, Gredt M. Patterns of cross resistance to fungicides in *Botryotinia fuckeliana* (*Botrytis cinerea*) isolates from French vineyards. Crop Protection. 1999; 18:687-697.

14. Madhu Meeta, Bedi PS, Jindal KK. Host range of *Botrytis cinerea* the incitant of grey mould of gram. Plant Disease Research. 1986; 3:77-78.
15. Moses GJ, Reddy RR. Grey rot of castor in Andhra Pradesh. Journal of Research. Andhra Pradesh Agricultural University. 1989; 17:74-75.
16. Prasad RD, Bhuvaneswari R. A modified medium for improved sporulation of gray mold pathogen, *Botryotinia ricini* (Godfrey) Whetzel in castor (*Ricinus communis* L.). Journal of oilseeds research. 2014; 31(1):79-81.
17. Purseglove JW. Tropical crops: Dicotyledons: London Longman. Green and Co. Ltd. London, 1968, 346.
18. Schmitz H. Poisoned food technique. Industrial and Engineering Chemistry Analyst Ed. 1930; 2:361.
19. Stevens HE. Grey mold of castor beans. Florida Grower. 1918; 18(8):6.
20. Suyal Usha, Tripathi HS. Management of grey mould (*B. cinerea*) of chickpea through fungicides. Pantnagar Journal of Research. 2011; 9(2):210-213.
21. Weiss EA. Castor, Sesame and Safflower. Leonard Hill, London, 1973, 901.
22. Yamuna C. Studies on gray mold of castor incited by *Botryotinia ricini* (Godfrey) Whetzel with special reference to disease Dissertation. Telangana State Agricultural University. Hyderabad, 2015, 1-121.