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Bio-efficacy of tricyclazole 22.5% W/V + azoxystrobin 7.5% W/V against sheath blight disease of paddy

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Abstract

A new combination fungicide Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) was tested against rice sheath blight diseases under field condition during *Kharif* 2015 and *Rabi* 2015-16. The combination fungicide Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha was found effective against sheath blight diseases by recording least Percent Disease Index (PDI) of 14.44 during *Kharif* 2015 and 13.35 during *Rabi* 2015-16. Significant increase in grain yield (61.53 q/ha) was also observed in the plots treated with Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha in *Kharif* 2015 (61.53 q/ha) and *Rabi* 2015-16 (64.62 q/ha), whereas, other fungicide treatments recorded the yield in a range of 48.88 – 59.53 q/ha (*Kharif* 2015) and 46.26 – 61.28 q/ha (*Rabi* 2015-16).

Keywords: Combination fungicides, rice, sheath blight, tricyclazole 22.5% W/V, azoxystrobin 7.5% W/V (6.79% W/W)

Introduction

Rice (*Oryza sativa* L.) is one of the most important food crops worldwide. In Asia, more than 90% of the rice is grown and consumed by nearly 60% of the world's population [1, 2]. It is affected by many biotic and abiotic stresses, among the biotic stresses; sheath blight is very important disease. The pathogen, *Rhizoctonia solani* has a wide host range infecting different crops and weeds [3]. Sheath disease of rice is a complex biotic stress, caused by *R. solani* (sheath blight), *R. oryzae* (sheath spot) and *R. oryzae-sativae* (aggregate sheath spot) [2]. Among these three, sheath blight of rice caused by *R. solani* (teleomorph: *Thanatephorus cucumeris*) is a destructive disease in many rice growing areas of the world, and this could reduce the grain yield by 58.60% [1, 4, 5]. In India, an estimation of losses due to the sheath blight disease alone has been up to 54.3% [6-8]. The disease has got more importance in intensive rice production systems due to excess use of nitrogenous fertilizers [8,9].

Previously, in India either solo or combination fungicides have been reported against different fungal diseases of rice such as blast [10-12], sheath blight [8, 10, 11, 13, 14], sheath rot [14], stem rot [15] and false smut [16-18]. Sheath blight disease is one of the most important diseases of rice but there is no genetic resistance available in rice against this disease [19]. Management of sheath blight of rice through fungicides is successful in majority of the cases [8, 11, 14, 20, 21]. For sheath blight, most of the fungicides like carbendazim, captafol, mancozeb, thiophanate, carboxin, zineb, benomyl, chloroneb, edifenphos, iprobenphos, etc. have been found effective under field conditions [8, 11, 22-24]. Along with these fungicides some of the new molecules have arrived which have combination fungicides like Trifloxystrobin 25% + Tebuconazole 50% 75WG, Azoxystrobin 18.2% + Difenoconazole 11.4% SC, Kresoxim methyl 40% + Hexaconazole 8% WG and Azoxystrobin 11% + Tebuconazole 18.3% w/w SC have been shown to control the sheath blight disease under field condition [8, 11, 20, 21, 25].

Although, fungicides are very effective in managing the fungal diseases, continuous use of same fungicide can lead to development of fungicide tolerance or even resurgence in fungal population; therefore, it is inevitable to search for new group of molecules with different mode

of action. New information generated on diverse fungicides with different modes of action can be offered to farmers for effective control of fungal diseases. In this view, the present study was undertaken to appraise the field efficacy of Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) against sheath blight disease of paddy under field conditions.

Materials and methods

A field experiment was carried out to evaluate bioefficacy of a fungicide Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) during *Kharif* -2015 and *Rabi* 2015-16 on sheath blight disease of rice at the AICRP-Rice, ARS, Gangavathi (UAS Raichur), Karnataka. The test variety BPT5204 was used. The experimental plots were laid out in randomized block design with three replications of eight treatments with a plot size of 40 sq. m and seedlings of 30 days old were planted in trail plots at 20×15 cm spacing.

The experiment comprises of eight treatments with three replications. A new fungicide Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) was tested in four doses (@ 800, 900, 1000 and 2000 ml/ha) along with Tebuconazole + Trifloxystrobin 75 WG (@200 g/ha), Tricyclazole 75% WP (@300 g/ha), Azoxystrobin 25% SC (@300 ml/ha). Two sprays of the fungicide were made at 15 days interval starting from the initiation of the disease. Artificial inoculation of sheath blight disease was done at 45 days after transplanting by following the 'mycelium with typha grass' method described previously [26].

Observations were recorded on disease severity in each treatment before and after two sprays as per the standard method. The observations of severity of sheath blight disease were recorded using 0-9 scale (SES, IRRI, 1996) at before and after each spray. In each replicated plot of the treatments, twenty randomly selected hills were selected and scored as per scale. The percent disease index (PDI) of plants was calculated by the following formula as presented below.

$$PDI = \frac{\text{Sum of numerical rating}}{\text{Total no. of hills observed} \times \text{Maximum grade}} \times 100$$

Results and discussion

In the recent years, combination fungicides are being widely used for the management of fungal diseases under field

conditions due to their broad spectrum, curative action and low dosage compared to their solo formulation. In paddy, the efficacy of such combination products in managing fungal disease has been reported previously [8, 11, 14, 20, 21, 25].

In the present study, field experiment revealed that the treatment of Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha after two applications at 15 days interval recorded least PDI of sheath blight disease in *Kharif* 2015 (14.44) and *Rabi* 2015-16 (13.35). The same dose was statistically on par with that of same treatment at 2000 ml/ha. The data also suggest that the standard check treatment Tebuconazole + Trifloxystrobin 75 WG at 200 g/ha recorded significantly more PDI over that Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha at final observation. Maximum PDI was recorded in untreated control (Table 1). The higher field bio-efficacy of combination fungicides against sheath blight disease of paddy has been reported previously where, combination fungicides Azoxystrobin 11% + Tebuconazole 18.3% w/w SC at 750-1000 ml/ha and Trifloxystrobin 25% + Tebuconazole 50% at 0.4 g/l were reported as effective against sheath blight disease of paddy [8, 11].

Optimum use of fungicides has been reported to enhance the crop yield due to reduction in disease load [8, 11, 12, 27, 28]. In the present study different level of severity of sheath blight in different treatment was observed (Table 1) and that was reflected in the final grain yield (Table 2). Significant increase in the grain yield in *Kharif* 2015 (61.53 q/ha) and *Rabi* 2015-16 (64.62 q/ha) was observed in the treatment treated with Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha followed by the same chemical at 2000 ml/ha (Table 2). Whereas, the other fungicidal treatments recorded the yield in a range of 48.88 – 59.53 q/ha in *Kharif* 2015 and 46.26 – 61.28 q/ha in *Rabi* 2015-16. The fair raise in the yield was mainly due to the reduced severity in the sheath blight disease of paddy. Our results are in conformity with those of previous reports [8, 11, 12, 20, 27] reported that fungicides application increases the yield of rice.

In conclusion, present investigation provides the field efficacy of a fungicide Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) at 1000 ml/ha for management of sheath blight disease of paddy.

Table 1: Effect of Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) application of against Sheath blight disease of rice

S. No.	Treatments	Dose	Sheath Blight (PDI)						Pooled	Percent Disease Reduction over Control	
			<i>Kharif</i> -2015			<i>Rabi</i> -2015-16				<i>Kharif</i> - 2015	<i>Rabi</i> - 2015-16
			Initial score	15 days after 1 st spraying	15 days after 2 nd spraying	Initial score	15 days after 1 st spraying	15 days after 2 nd spraying			
1	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	800	25.33	23.33	21.53	29.99	25.55	20.03	20.78	52.7	63.94
2	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	900	33.33	25.55	20.55	31.33	22.22	21.55	21.05	54.8	61.2
3	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	1000	31.55	18.88	14.44	31.11	17.77	13.35	13.89	68.3	75.96
4	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	2000	25.55	16.45	13.55	35.55	17.56	11.23	12.39	70.25	79.78
5	Nativo (Tebuconazole +	200	31.11	28.58	22.53	29.99	22.55	19.55	21.04	50.5	64.8

	Trifloxystrobin 75 WG)										
6	Tricyclazole 75% WP	300	33.33	35.55	39.99	28.88	39.99	45.55	42.77	12.2	18.01
7	Azoxystrobin 25 SC	300	29.99	27.77	25.55	30.33	25.55	24.44	24.99	43.9	56.00
8	Control	--	33.33	39.99	45.55	26.66	41.11	55.55	50.55	--	--
CV @ 5%			NS	11.5	10.8	NS	10.5	12.85			
CD			NS	3.95	4.2	NS	3.42	4.6			

Table 2: Effect of application of Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W) on grain yield *Kharif*-2015 and *Rabi* -2015-16

S. No.	Treatments	Product Dose (ml or gm/ ha)	Grain Yield (q/ha)	
			<i>Kharif</i> – 2015	<i>Rabi</i> – 2015-16
1	Tricyclazole 22.5%W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	800	52.55	58.65
2	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	900	53.55	54.82
3	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	1000	61.53	64.62
4	Tricyclazole 22.5% W/V (20.36% W/W) + Azoxystrobin 7.5% W/V (6.79% W/W)	2000	59.53	61.28
5	Nativo (Tebuconazole + Trifloxystrobin 75 WG)	200	54.26	55.23
6	Tricyclazole 75% WP	300	49.53	48.85
7	Azoxystrobin 25 SC	300	48.88	46.26
8	Control	--	41.53	39.25
CV @5%			13.56	10.22
CD			3.8	4.11

References

- Guyer D, Tuttle A, Rouse S, Volrath S, Johnson M, Potter S *et al.* Activation of latent 171 transgenes in arabidopsis using a hybrid transcription factor. *Genetics*. 1998; 149:633-639.
- Nagaraj BT, Gururaj Sunkad, Pramesh D, Naik MK, Patil MB. Characterization of *Rhizoctonia* Species Complex Associated with Rice Sheath Disease in Karnataka. *Agricultural Research*. 2019; 8:191-196.
- Nagaraj BT, Gururaj Sunkad, Pramesh D, Naik MK, Patil MB. Host Range Studies of Rice Sheath Blight Fungus *Rhizoctonia solani* (Kuhn). *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(11):3856-3864.
- Groth DE. Effects of cultivar resistance and single fungicide application on rice sheath blight, yield, and quality. *Crop Protection*. 2008; 27:1125-1130.
- Bernardes J, Michelangelo S, Marcello Z, Wenxiang W, Daohong J, Li S *et al.* Genetic structure of populations of the rice-infecting pathogen *Rhizoctonia solani* AG-1 IA from China. *Phytopathology*. 2009; 99:1090-1099.
- Rajan CPD. Estimation of yield losses due to sheath blight of rice. *Indian Phytopathology*. 1987; 40:174-177.
- Roy AK. Sheath blight of rice in India. *Indian Phytopathology*. 1993; 46:97-205.
- Pramesh D, Maruti, Saddamhusen A, Muniraju KM, Guruprasad GS. A New Combination Fungicide Active Ingredients for Management of Sheath Blight Disease of Paddy. *Advances in Research*. 2016; 8(5):1-7.
- Savary S, Mew TW. Analyzing crop losses due to *Rhizoctonia solani*: Rice sheath blight, a case study. In: Sneh B, Javaji Hare S, Neate S, Dijst G, (eds). *Rhizoctonia* species: Taxonomy, Molecular Biology, Ecology, Pathology and Disease Control, Kluwer, Dordrecht, 1996, 237-244.
- Kumar PMK, Sidde Gowda DK, Rishikant M, Kiran Kumar N, Pandurange Gowda KT, Vishwanath K. Impact of fungicides on rice production in India In: *Fungicides showcases of integrated plant disease management from around the world (open access chapter)*, 2013, 77-98.
- Pramesh D, Maruti, Muniraju KM, Mallikarjun K, Guruprasad GS, Mahantashivayogayya K *et al.* Bio-efficacy of a Combination Fungicide against Blast and Sheath Blight Disease of Paddy. *Journal of Experimental Agriculture International*. 2016; 14(4):1-8.
- Pramesh D, Nataraj K, Guruprasad GS, Mahantashivayogayya, Reddy BGM. Evaluation of a new strobilurin group of fungicide for the management of blast disease of paddy. *American Journal of Experimental Agriculture*. 2016; 13(5):1-6.
- Bag MK. Performance of a new generation fungicide Metominostrobin 20SC against sheath blight disease of rice. *Journal of Mycopathological Research*. 2011; 49(1):167-169.
- Pramesh D, Alase S, Muniraju KM, Kirana Kumara M. A Combination Fungicide for the Management of Sheath Blight, Sheath Rot and Stem Rot Diseases of Paddy. *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(9):3500-3509.
- Pramesh D, Saddamhusen A, Muniraju KM, Kirana Kumara M. Management of Stem Rot Disease of Paddy Using Fungicides. *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(10):3046-3051.
- Muniraju KM, Pramesh D, Mallesh SB, Mallikarjun K, Guruprasad GS. Novel Fungicides for the Management of False Smut Disease of Rice Caused by *Ustilagoidea virens*. *International Journal of Current Microbiology and Applied Sciences*. 2017; 6(11):2664-2669.
- Savitha AS, Nagaraja A, Pramesh D, Chethana BS. Bioefficacy of novel fungicide molecules in the management of false smut of rice caused by *Ustilagoidea virens*. *International Journal of Chemical Studies*. 2019; 7(4):3208-3212.
- Sharanabasav H, Pramesh D, Chidanandappa E, Saddamhusen A, Amoghvarsha C, Raghunandana A Prasanna Kumar MK, Raghavendra BT, Naik RH, *et al.* Field evaluation of fungicides against false smut disease of rice. *Journal of Pharmacognosy and Phytochemistry*. 2020; 9(3):1453-1456.
- Bonman J, Khush G, Nelson R. Breeding rice for resistance to pests. *Annual Review of Phytopathology*. 1992; 30:507-528.
- Bhuvaneshwari V, Raju KS. Efficacy of New Combination Fungicide against Rice Sheath Blight Caused by *Rhizoctonia solani* (Kuhn). *Journal of Rice Research*. 2012; 5(1-2).

21. Kumar PMK, Veerabhadraswamy AL. Appraise a combination of fungicides against blast and sheath blight diseases of paddy (*Oryza sativa* L.). Journal of Experimental Biology and Agricultural Sciences. 2014; 2:49-57.
22. Dash SC, Panda S. Chemical control of rice sheath blight disease. Indian Phytopathology. 1984; 37:79-82.
23. Kannaiyan S, Prasad NN. Effect of foliar spray of certain fungicides on the control of sheath blight of rice. Madras Agricultural Journal. 1984; 71:111-114.
24. Singh R, Sihna AP. Comparative efficacy of local bioagents, commercial bioformulation and fungicide for the management of sheath blight of rice under glass house conditions. Indian Phytopathology. 2004; 57:494-496.
25. Bag MK, Saha S. Fungitoxic effect of Nativo 75 wg (trifloxystrobin 25%+tebuconazole 50%) on grain discoloration (GD) disease of rice in West Bengal. Pestology. 2009; 33:47-49.
26. Bhaktavatsalam G, Satyanarayana K, Reddy APK, John VT. Evaluation of sheath blight resistance in rice. International Rice Research Newsletter. 1978; 3:9-10.
27. Bag MK, Yadav M, Mukherjee AK. Bioefficacy of strobilurin based fungicides against rice sheath blight disease. Trancryptomics. 2016; 4:128.
28. Biswas A, Bag MK. Strobilurins in management of sheath blight disease of rice: A review. Pestology. 2010; 34:23-26.