



P-ISSN: 2349-8528

E-ISSN: 2321-4902

www.chemijournal.com

IJCS 2020; 8(3): 2909-2914

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Received: 24-03-2020

Accepted: 28-04-2020

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Study of microbial load for storage and optimization conditions of Malta (*Citrus Sinensis*) using response surface methodology

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DOI: <https://doi.org/10.22271/chemi.2020.v8.i3ap.9655>

Abstract

Response surface methodology (RSM) was used to determine the optimum storage conditions of malta fruits that gives minimum microbial load or growth and maximum shelf life retained value. Scavenger (3–5 g), polythene thickness (75–125 gauge) and fungicide concentration (75–200 ppm) were the factors investigated. Experiments were designed according to Box–Behnken design with these three factors, including central points. For each response, a second-order polynomial model was developed using multiple linear regression analysis. Applying desirability function method, optimum storage conditions were found to be 5 g scavenger, 125-gauge polythene thickness and 200 ppm fungicide concentration. All the parameters significantly affect the response. It is observed that all factors are suitable to reduce the microbial load which is helpful for extending the shelf life of malta fruits up to four months at ambient condition. Box Benkhen can be successfully applied for modeling and optimizing the storage parameters. Applying desirability function method, optimum storage conditions were found to be 5 g scavenger, 125-gauge polythene thickness and 200 ppm fungicide concentration.

Keywords: Box–Behnken, RSM, storage of malta, microbial load and shelf life

Introduction

Malta or Sweet orange (*Citrus sinensis*) is a member of the citrus family (Snart 2006) [12]. Citrus spp. consists of group of fruits belonging to the Rutaceae family. Malta is having high acceptability due to its attractive color, distinctive flavor and taste. The major medicinal properties of orange include anti-bacterial, anti-fungal, anti-diabetic, cardio-protective, anti-cancer, anti-arthritis, anti-inflammatory, anti-oxidant, anti-Tubercular, anti-asthmatic and anti-hypertensive (Parle and Chatuevedi; 2012) [8]. However, malta is available for only one or two months, therefore it cannot be stored for longer period under ambient conditions and cannot be transported to distant places due to its short shelf life. Citrus fruits are non-climacteric in nature and their eating quality cannot be improved after harvest, so care should be taken at the optimum harvesting time period. Enhancement of shelf-life of malta become necessary for the purpose of increasing the local raw materials, decreasing the post-harvest losses and for using the processed product out of season (Dasmohapatra *et al.*, 2011) [1] and all these factors affect the national economy as well as state which will be beneficial for farmers also. In order to store malta fruits active packaging (consists of scavenger such as KMnO₄ and activated charcoal) was used prior to fungicide treatment. In view of this, active packaging for storage of malta was investigated. During the storage the formation of CO₂ is enhanced due to respiration reactions, and more and more the ethylene is produced from packaged fruits and vegetables. Accumulated gases produced during the storage time, have to be removed from the package to avoid the food deterioration and the packaging destruction so there should be provision of a system which can remove these gases and for this purpose scavenger is a good option (Floros, 1997) [2]. Moreover, the development of these activities will make it possible to increase the value of local raw materials; decrease post-harvest losses and to use processed products out of season. It is also helpful in increasing farmers' income; and integrates family-type rural farming into the market economy as it is a method to store the commodity at ambient

condition as no cold storage, extra energy and electricity required for this method (Richa *et al.* 2016) [11].

Three effective factors; scavenger amount, polythene thickness, and fungicide concentration were used for enhancement of shelf life of malta. The statistical method using response surface methodology (RSM) has been proposed to determine the influences of individual factors and the influence of their interactions. RSM has been reported to be an effective tool for optimization of a process when the independent variables have a combined effect on the desired response. RSM is a collection of statistical and mathematical system that has been successfully used for developing, improving and optimizing such processes (Koocheki *et al.* 2009) [5]. It is a technique to design experiments, build models, evaluate the effects of several factors, and to attain the optimum conditions for desirable responses with a limited number of planned experiments. RSM helps to demonstrate how input variables over some specified region of interest affects particular response and what independent variables will yield a maximum (or minimum) for a specific response (Khuri and Cornell, 1996) [4]. The response surface design was used in this study: 1) to find how microbial count comprising of several levels of shelf life factors can be simplified, 2) to determine how microbial count and shelf life (as responses) are affected by changes in the level of storage parameters, 3) to determine the optimum combination of storage parameters that yields the best storage life. These objectives were quantitatively measured by RSM for optimization of microbial count, shelf life and color

2. Materials and methods

Fresh malta fruits were used for storage study. The fresh malta was procured from the market of Almora district, Uttarakhand; the storage study was conducted for 4 months in the lab. Fruits were dipped in azoxystrobin fungicide solution for 1 min to resist fungus attack and after that fruits were kept under ambient conditions at least 5-6 h so that the moisture of fungicide solution get evaporated from malta surface and fruit become safe for storage i.e. microorganism could not attack on malta due to absence of moisture. Digital balance was used for weighing the treated malta fruits and then two malta fruits were kept in one box. On the base of box, decided amount of scavenger (KMnO₄ and activated charcoal) was kept in petri dish. Boxes were packed with polythene bag of different thickness (permeability) and stored at room temperature for further analysis for microbial load and shelf life as per method prescribed (Ranganna, 2005) [9]. All the treated samples were kept for four months storage. For storage of malta, mill board boxes (18.5×15×10cm) were used. Each box has base in which six perforations were made and each side face has two perforations for the exchange of gases so that accumulation of gases does not take place and fresh air was maintained inside the box to prevent the an aerobic condition inside the box.

2.1. Experimental design

Independent variables should satisfy important assumptions which are as follows

- Variables should be measurable,
- Variables should be continuous and
- Variables should be controllable by experiments, with negligible errors,

The RSM course of action was carried out as follows (Trinh and Kang, 2010) [13]:

1. For sufficient and dependable measurement of the response of interest a series of experiments were performed
2. The second-order response surface mathematical model with the best fit was developed.
3. The optimum response value was determined.

RSM was used as it helps to reduce the number of experiments without affecting the accuracy of results and to decide interactive effects of variables on the response. Box-Behnken designs are response surface designs especially made to require only 3 levels, coded as -1, 0, and 1. Box-Behnken designs may consist of 3 to 21 factors. They are formed by combining two levels factorial design with incomplete block design (Khuri and Cornell, 1996) [4].

Box-Behnken design, which is a very efficient design tool for fitting second-order models, was selected for use in this study. Based on literatures and preliminary experiments conducted the level chosen for scavenger (X₁), polythene thickness (X₂) and fungicide (X₃) for enhancing the shelf life of malta under ambient condition. After defining the desired value ranges of the independent variables, they were coded as ±1 as the factorial points, 0 as the center points. The ranges of variables as in coded and actual form for the design of experiments are shown in Table 1.

Table 1: Levels of independent variables in coded and actual form for storage study of malta fruits

Independent variables		Coded Levels		
Name	Code	-1	0	+1
		Actual Levels		
Scavenger (g)	X ₁	3	4	5
Polythene thickness (gauge)	X ₂	75	100	125
Fungicide concentration (ppm)	X ₃	100	150	200

*coded and actual values used in response surface methodology for experiment design

2.2. Measurements of quality parameters

2.2.1 Microbial count of Malta during storage

After the preparation of samples, microbial load in fermenting liquor evaluated by pour plating method.

2.2.1(a) Preparation of serial dilution series

One ml of malta juice sample was withdrawn from each flask and diluted into nine ml of distilled water, which was 10⁻¹ dilution. In the same way, dilution series up to 10⁻⁹ dilution levels were prepared.

One ml sample from 10⁻³ to 10⁻⁹ dilution level was inoculated into three different petri plates and PDA media was poured over the samples and mixed thoroughly by rotating clockwise and anticlockwise several times. The plates were set in duplicates. All plates were incubated at 30 °C. After three days, the number of colonies on each plate were observed, counted and expressed as CFU/ml of Malta juice

2.2.1(b) Total viable counts in Malta.

Each petri plate was divided into four equal parts and then the colonies were counted in a single part and then multiplied by 4, this gives the total number of colonies in a single Petri plate. (Ranganna, 2005) [9].

$$\text{Colony forming unit (CFU)} = \frac{\text{No of colonies} \times \text{Dilution Factor}}{\text{Aliquot sample}} \quad (1)$$

2.3 Second order polynomial model

A complete second mathematical model (Equation 2) model was fitted to the data and the adequacy of the model was tested considering the coefficient of multiple determinations (R^2), Fisher's F-test and lack of fit (LOF). The models were then used to interpret the effect of various parameters on the response. Optimization of process parameters was carried out at the end of the analysis. The optimized values of process parameters were used in storage and contours were developed for selected parameters.

The second order mathematical response function for three independent variables has the following form:

$$Y = \beta_0 + \sum_{i=1}^3 \beta_i X_i + \sum_{i=1}^2 \sum_{j=i+1}^3 \beta_{ij} X_i X_j + \sum_{i=1}^3 \beta_{ii} X_i^2 \quad (2)$$

Or

$$Y = \beta_0 + \beta_1 X_1 + \beta_2 X_2 + \beta_3 X_3 + \beta_{12} X_1 X_2 + \beta_{13} X_1 X_3 + \beta_{23} X_2 X_3 + \beta_{11} X_1^2 + \beta_{22} X_2^2 + \beta_{33} X_3^2 \quad (3)$$

where,

β_0 , β_i , β_{ii} , β_{ij} are coefficient of regression (coefficients of regression of linear, interaction and quadratic terms, respectively)

X_i , X_j are independent variables (where, $i=1, 2$, and $j=1, 2, \dots$)

Y is the predicted response (microbial count or load and shelf life) used as a dependent variable

x_i ($i = 1, 2$) the input predictors or controlling variables (factors)

Multiple linear regression analysis was used for determining the coefficient parameters by employing the software Design-Expert (version 8). Design-Expert was also used to find the 2-D contour plots of the response models. The value of p in the model represented the probability of significance. Model analysis, lack-of-fit test and R^2 (coefficient of determination) analysis is used to check the adequacies of the models (Lee *et al.* 2000 and Weng *et al.* 2001) [6, 14]. The lack-of fit evaluates the failure of a model to represent data in the experimental domain at which points were not included in the regression and variations in the models cannot be accounted by random error (Montgomery 1984) [7]. Coefficient of variation (CV) indicates the relative dispersion of the experimental points from the model prediction.

3. Results and discussion

3.1. Model fitting

Seventeen observed response values observed were used to compute the mathematical model using the least square method. The two dependent variables as microbial count and shelf life were correlated with the three factors (scavenger amount, polythene thickness and fungicide concentration), using the second-order polynomial, as represented by Eqn 2. After fitting experimental data, quadratic regression models were developed, as shown in Eqn (4) and (5).

$$\text{CFU or microbial count (Y)} = 6.6 \times 10^5 - 2.37 \times 10^4 X_1 - 3.37 \times 10^4 X_2 - 1.45 \times 10^5 X_3 - 1.75 \times 10^4 X_1 X_2 - 2.0 \times 10^4 X_1 X_3 + 1.0 \times 10^4 X_2 X_3 + 4.82 \times 10^4 X_1^2 - 2.67 \times 10^4 X_2^2 + 4.57 \times 10^4 X_3^2 \quad (4)$$

Where,

Y = Colony forming unit (CFU/ml)/ microbial count

$$\text{Shelf life (Y)} = 102.00 + 8.75 X_1 + 13.75 X_2 + 0.000 X_3 + 7.50 X_1 X_2 + 0.000 X_1 X_3 + 0.000 X_2 X_3 - 2.25 X_1^2 - 7.25 X_2^2 + 0.25 X_3^2 \quad (5)$$

Y = shelf life (days)

Where, X_1 , X_2 and X_3 are the amount of scavenger, thickness of polythene and concentration of fungicide. The coefficients with one factor (the ones in front of X_1 or X_2 or X_3) represent the effects of the certain factor, while the two factors coefficients (the ones in front of $X_1 X_2$ or $X_1 X_3$ or $X_2 X_3$) and quadratic terms (the ones in front of X_1^2 or X_2^2 or X_3^2) represent the interaction between the two factors and the effect of quadratic term respectively. The synergistic effect was shown by positive sign effect, while the negative sign indicates an antagonistic effect.

It is noted that coefficients with p value < 0.01 are those with significant effects toward the microbial load and shelf life, thus those variables with $p > 0.01$ can be omitted from the regression. Moreover, under the assumption of the insignificant effect of interaction, the variables X_2^2 , X_1 and, X_3 , respectively are neglected for microbial count and $X_2 X_3$, $X_1 X_3$ and quadratic term of X_1 and X_3 are neglected for shelf life. Furthermore, it is observed that variable X_3 fungicide addition play important role to stop the growth off fungus. Therefore, equation with significant term can be written as follows-

$$\text{CFU or Microbial count (Y)} = 6.6 \times 10^5 - 3.37 \times 10^4 X_2 - 1.45 \times 10^5 X_3 + 4.82 \times 10^4 X_1^2 + 4.57 \times 10^4 X_3^2 \quad (6)$$

$$R^2 = 97.99\% \text{ and Adj R-Squared} = 95.40\%$$

Equation (6): Microbial count will increase by decreasing the value of scavenger; polythene thickness and fungicide concentration. The effect of polythene thickness (3.37) is the highest and as compare to amount of scavenger (2.37).

$$\text{Shelf life (Y)} = 102.00 + 8.75 X_1 + 13.75 X_2 + 7.50 X_1 X_2 - 7.25 X_2^2 \quad (7)$$

$$R^2 = 96.12\% \text{ and Adj R-Squared} = 91.13\%$$

Equation (7): There will be an increase in shelf life with increase of all three parameters. The polythene thickness has got more effect on shelf life will the value of 13.75 while a scavenger has less effect as compare to polythene thickness 8.75.

3.2. Validation of the Models

The developed quadratic model is usually used to examine for ensuring it provides an adequate approximation to the actual system. When model shows an adequate fit, then proceeding with an investigation and optimization of the fitted response surface is likely to give good or real results. Graphical techniques, graphical and numerical methods were used as a primary tool and confirmation to validate the models (Trinh and Kang, 2010) [13].

The developed models were then checked using a numerical method employing the coefficient of determination (R^2), adjusted R^2 (R^2_{adj}), and then calculated as shown in Equation (8) and (9) (Haber and Runyun, 1977) [3], the coefficient of determination R^2 indicates how much of the observed variability in the data was accounted for by the model, while by taking into account the number of covariates or predictors in the model R^2_{adj} modifies R^2 .

$$R^2 = 1 - \frac{SS_{Residual}}{SS_{Model} + SS_{Residual}} \quad (8)$$

$$R^2_{Adj} = 1 - \frac{n-1}{n-p} (1 - R^2) \quad (9)$$

Where, SS is the sum of the squares, n experiments number, and p the number of predictors (term) in the model, not counting the constant term. Experimental data were used to develop the quadratic model in this study with the values of R^2 higher than 90%, say 97.99% and 96.12% of microbial load and shelf life of stored Malta, respectively. Furthermore, an R^2_{adj} very close to the R^2 values for the response microbial load than the shelf life ensures a satisfactory adjustment of the quadratic models to the experimental data and having least residual error in the model. Therefore, the regression models explained to check the effect of each response on independent variables well.

3.3 Microbial count (Colony Forming Unit (CFU)) and shelf life

Colony Forming Unit (CFU) increases as storage time is increased and as the substrate is being utilized it becomes constant. During storage CFU ranged from $5.0E+4$ to $9.0E+4$ during the course of storage. Maximum cell count was observed for experiment no. 5 (3 g scavenger level, 100 gauge polythene packaging and 100 ppm azoxystrobin fungicide) of storage while minimum was at experiment no. 12 at (4 g scavenger, 100 gauge polythene and 200 ppm azoxystrobin fungicide concentration). The microbial counts or load did not exceed the acceptable limits in the treatments; however, higher counts were observed at the end of storage.

ANOVA on microbial count model, as shown in Table 2, demonstrates that the models were highly significant ($p < 0.01$). A model F value 4.12 implies that the model is significant. The Fisher F-test with a very low probability value ($P_{model} \geq F$ at 0.01) demonstrates a very high significance for the regression model. The determination coefficient (R^2) checks the goodness of fit of the model. The coefficient of determination (R^2) was 97.99% for microbial count. The lack of fit F-value of 1.34 indicates the lack of fit is not significant relative to the pure error. There is a 38.05% chance that a lack of fit F-value this large could occur due to noise. Non-significant lack of fit indicates good fit model. The Pred R^2 of 82.31% is in reasonable agreement with the Adj R^2 of 95.40%. The signal-to-noise ratio is measured by Adequate precision (Adeq precision) whose desirable value is 4. Noise ratio of 20.781 indicates an adequate signal.

The shelf life ranged for the storage from 80 to 120 days. Maximally all the treated Malta fruit showing 80 days storage life. The maximum shelf life was observed for Expt. 4 (5g of scavenger (5 g $KMnO_4$ + 5 g of activated charcoal), 125 gauge thick polythene wrap and 150 ppm of fungicide whose shelf life was 120 days while minimum range of shelf life was observed at Expt. (3 g of scavenger (3 g $KMnO_4$ + 3 g of activated charcoal), 150 ppm of azoxystrobin fungicide ($X_3 = 0$) and 75 gauge thickness polythene wrap. Ethylene scavenger reduce the Ethylene content which is a main ripening inducing agent of fruits may cause the premature ripening of some products, even ruin others. Beside this packaging material maintain accumulation of CO_2 , restricted intake of O_2 from atmosphere, low level of relative humidity of the air in the storage space, and the lower rate of

respiration of fruits. Therefore, these combinations could help to enhance the shelf life. This finding is similar to Reddy *et al.*, 2008^[10].

ANOVA on shelf life model, as shown in Table 2, demonstrates that the models were highly significant ($p < 0.01$). A model F value 19.26 implies that the model is significant. The Fisher F-test with a very low probability value ($P_{model} \geq F$ at 0.01) demonstrates a very high significance for the regression model. The determination coefficient (R^2) checks the goodness of fit of the model. The coefficient of determination (R^2) was 96.12% for shelf life. The lack of fit F-value of 0.42 indicates the lack of fit is not significant relative to the pure error. There is a 75.10% chance that a "Lack of Fit F-value" this large could occur due to noise. The Pred R^2 of 80.60% is in reasonable agreement with the Adj R^2 of 91.13%. The signal-to-noise ratio is measured by Adequate precision (Adeq precision) whose desirable value is 4. Noise ratio of 15.149 indicates an adequate signal.

The model was found to be adequate and the best fit equation were developed in order to draw contour plots for showing the effect of independent variables on those responses and to select the range of variables for an acceptable Malta storage condition. Lines or curves of constant response values are drawn on a plane or graph whose coordinate axes represent the levels of independent variables and the response is visualized perpendicular to the plane of paper. Series of contour lines of equal response value were generated which provided useful information for understanding the effect of two independent parameters on the dependent variable.

Table 2: Total effect of individual parameter on microbial count for stored Malta

Source	DF	Microbial count		Shelf life	
		MS	F-Value	MS	F-Value
Model	9	2.55E+9	4.12**	288.98	19.26***
Scavenger (X_1)	4	4.0E+10	65.96***	214.705	14.31***
Polythene thickness (X_2)	4	3.25E+9	5.35**	489.705	32.647***
Fungicide conc ⁿ (X_3)	4	4.47E+10	73.71***	0.065	0.00433
Linear	3	6.03E+10	99.43***	708.33	47.22***
Quadratic	3	7.2E+9	11.87***	80.96	5.3**
Interactive	3	1.06E+9	1.748	75	5**
Error	7	6.05E+8		15	
Total	16				

***, ** Significant at 1 and 5% level of significance respectively

3.4 Coding of the variables was done as per the following

3.4.1 Storage

The independent variables were coded as X_1 , X_2 and X_3 for scavenger, polythene and fungicide as per given below, respectively.

$$X = \frac{\text{Independent variable} - \text{central value}}{\text{Interval gap}}$$

$$X_1 = \frac{\text{Scavenger} - 4}{1}$$

$$X_2 = \frac{\text{Polythene} - 100}{25}$$

$$X_3 = \frac{\text{Fungicide} - 150}{50}$$

3.5 Numerical optimization of process parameters for of malta

Optimization is a process of making compromises between responses, to achieve a common target. Numerical optimization was carried out using Design-Expert 8.0.6 statistical software and carried out to predict the optimum storage condition of malta fruits within selected ranges which generated the desired response goal. The desired goals for each factor and response were chosen and different weights were assigned to each goal. The responses namely microbial load and shelf life, were taken into consideration for optimization. The goal seeking begins at random starting points and proceeds up and down the steepest slope on the response surface for a maximum and minimum value of the response respectively. Importance to the responses and independent variables were given on the basis of objectives of the study. Maximum importance (++++) was given to the minimum to microbial count and maximize the shelf life.

Optimization of processing conditions was carried out to keep the scavenger and fungicide in range and shelf life retained. Based on mentioned criteria, the optimization was carried out. During optimization, 17 solutions were obtained in case of storage of malta, out of which the one that suited the criteria most was selected. The goal setup for optimization of shelf life of malta and the constraints are given in Table 3.

Table 3: Constraints and optimum values of process parameters and responses for optimization of stored malta

Names	Goal	Lower limit	Upper limit
Scavenger amount	Is in range	-1	+1
Polythene thickness	Maximize	-1	+1
Fungicide Concentration	Is in range	-1	+1
Microbial count	Minimize	5000	9000
Shelf life	Maximize	80	120

The optimum solution was emerged out as storage condition is 5 g scavenger (5 g KMnO_4 and 5 g activated charcoal), 125 gauge polythene thickness and 200 ppm fungicide which is optimized condition for enhancing the shelf life of malta fruit to retain its quality, in order to obtain optimized storage as microbial load and shelf life. Fig 1 shows contour plot for microbial load as a function of scavenger and fungicide at optimum point which depicts microbial count increases by decreasing the value of polythene thickness and fungicide concentration while Fig 2 shows contour plot for shelf life as a function of scavenger with polythene thickness at optimum point. Shelf- life of malta fruit increases with increase in scavenger amount and polythene thickness.

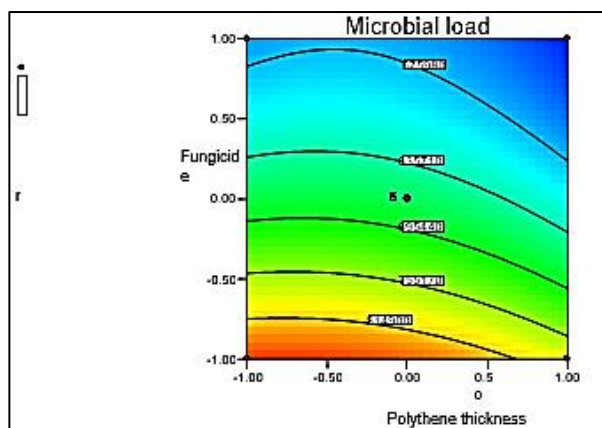


Fig 1: Contour plot for microbial load as a function of scavenger and fungicide at optimum point

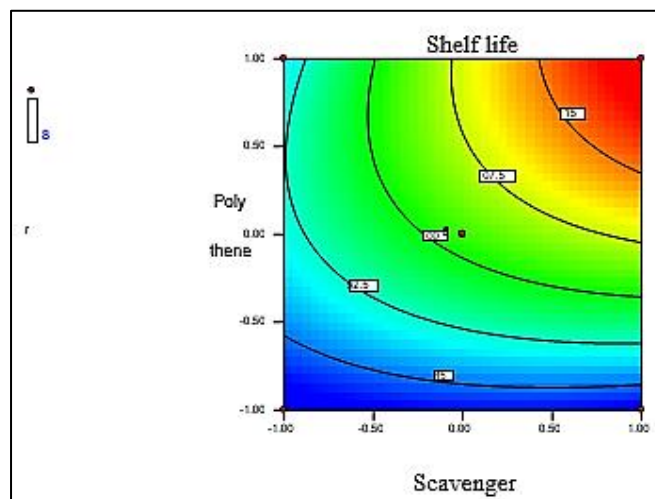


Fig 2: Contour plot for shelf life as a function of scavenger with polythene thickness at optimum point

4. Conclusion

RSM was effectively used to determine the effect of scavenger, polythene thickness and fungicide during storage of malta fruits. A total of 17 combinations of these independent variables were formed to see the effect on different dependent variables microbial count and shelf life. On the basis of experiment studies and data analysis it can be concluded that microbial count and shelf life could be maintained by using scavenger, polythene and fungicide. The optimum condition is reached by using 5 g scavenger (5 g KMnO_4 and 5 g activated charcoal), 125 gauge polythene thickness and 200 ppm of fungicide concentration to get minimum microbial load while maintaining maximum shelf life by experimental and analytical data. The results of a confirmation experiments were found to be in good agreements with the values predicted by model. It demonstrates that to get maximum amount of information, in a short period with the least number of experiments, Box Benken can be successfully applied for modeling and optimizing the storage parameters.

Acknowledgements

This research was carried out in the department of Post Harvest Process & Food Engineering and funded by the Indian Council of Agriculture Research under All India coordinated research project on Post Harvest Technology. We sincerely thanks to Head, PHPFE, Director Research and Dean, College of Technology, G.B.P.U.A. & T., Pantnagar for providing necessary requirements for smooth conducting of the research work.

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