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# Screening of colocasia genotypes for natural occurrence and reactions against leaf blight disease

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### **Abstract**

Twenty four genotype of colocasia were collected from throughout the Chhattisgarh state and screened against leaf blight disease in natural occurrence and infection at field condition for reactions during *Kharif* season of 2018 and 2019. Results revealed that among screened genotypes, three genotypes (IGCOL-PITH-17-1, IGCOL-PKJ-17-1 and IGCOL-CHLN-17-1) showed least terminal per cent disease infection (0.00%) in both years (during 2018 & 2019). Likewise the average terminal per cent disease infection also 0.00% and were significantly superior over rest of the genotypes. In reaction levels, three genotypes (IGCOL-PITH-17-1, IGCOL-PKJ-17-1 and IGCOL-CHLN-17-1) were found having no reactions *i.e.* resistant, six genotypes (IGCOL-BDL-17-1, IGCOL- PWND-17-2, IGCOL-KS-10-1, IGCOL-KS-13-2, IGCOL-CHHTD-18-1 and IGCOL-MUNG-17-1) were exhibited moderately resistant, eleven genotypes (IGCOL-BNR-17-1, IGCOL-MNDG-17-1, IGCOL-GB-17-1, IGCOL-DK-17-1, IGCOL-BL-12-1, IGCOL-JS-12-1, IGCOL-KDKN-17-1, IGCOL-PWD-17-1, IGCOL-CHMD-18-1, IGCOL-LHDD-18-1 and White Gauriya (S.C.)) were recorded moderately susceptible and two genotypes *viz.*, IGCOL-KOTA-17-1 and IGCOL-SPL-17-1 were observed to be susceptible while, two genotypes (IGCOL-NGR-17-1 and IGCOL-PWND-17-1) were found highly susceptible against *Phytophthora* leaf blight disease of colocasia under *in-vivo* condition.

Keywords: Colocasia genotype, Phytophthora, Screening and PDI

### Introduction

Colocasia is a tuber crop, grown throughout India because of its wide adoptability, large scale acceptability and high return unit area<sup>-1</sup> (Gurung, 2001) <sup>[1]</sup>. Indian states: Andhra Pradesh, Bihar, Gujarat, Karnataka, Kerala, Madhya Pradesh Maharashtra, Tamil Nadu and West Bengal where it is grown well in lowland and upland area. The corms and cormels are consumed as staple or subsistence food in developing countries. This is member of the Araceae family and is believed to be one of the earliest cultivated tuber crops in the world. Colocasia is now the fifth most-consumed tuber vegetable worldwide (Lokesh *et al.*, 2014) <sup>[2]</sup>. Cormels are rich source of calcium, phosphorus, protein, starch and vitamin C (Fageria *et al.* 2006) <sup>[3]</sup>. Nutritive value of colocasia is given as per 100gm fresh tuber *i.e.* moisture(g) 73.1, Protein(g) 3.0, Fat(g) 0.1, Minerals(g) 1.7, fibre(g) 1.0, Carbohydrate(g) 21.10, Energy Kcal 97.0, Calcium(Mg) 40, Phosphorus(mg) 140, Iron(mg) 1.7, Vitamin A (IU) 24, Thiamine(mg) 0.09, Niacin(mg) 0.4 and Riboflavin (mg) 0.03 (Misra *et. al.*, 2008) <sup>[4]</sup>.

The crop is affected by several pathogens among them *Phytophthora colocasiae* Raciborski a oomycetes fungus cause the most destructive disease and is responsible for causing leaf blight and lead to heavy yield losses (25 to 50 per cent) of taro in India (Gadre and Joshi, 2003) <sup>[5]</sup>. Leaf blight of colocasia was reported for the first time in India, by Butler and Kulkarni, (1913) <sup>[6]</sup>. Occurrence of Phytophthora blight and subsequent yield loss may have a negative impact in cultivation of this crop by the farming communities. In this situation, it was necessary to find out a resistant variety if any among the available genotypes against *Phytophthora colocasiae* under field conditions. Hence keeping these facts in mind present experiment was laid out to screening collected genotypes against *Phytophthora colocasiae* which cause severe losses in colocasia cultivation.

### **Materials and Methods**

Colocasia genotypes were collected from overall Chhattisgarh and were grown at Research cum Instructional Farm of Saheed Gundadhur College of Agriculture and Research Station, Jagdalpur. Experiments were conducted during Kharif season of 2018 and 2019 respectively, using Randomized Block Design (RBD) which comprised twenty four treatments with three replications. Visual observations on initial and final disease symptoms were recorded in the field to know the development of the disease in a plant population under natural condition. For disease incidence, 5 plants were randomly selected and recorded. The selected plants were tagged and observations on disease incidence were recorded on same plants, twice at 15 days interval during both of year. The disease rating scale proposed by Little and Hills (1978) [7], was followed to record the observations and per cent disease intensity (PDI) was calculated by the formula given by Mckinney (1923) [8]. A scale from 0 to 5 was used for assessing the disease for estimation of leaf blight where in, 0 grade— No disease, 1 grade -1-10% disease, 2 grade -11-25% disease, 3 grade -26-50% disease, 4 grade - 51-75%, 5 grade >75% disease. Recorded data were analysed online in OPSTAT statistical software package for agricultural research workers developed by Sheoran *et al.* (1998) [9] at CCS HAU, Hisar. Critical difference was calculated at the level of 0.05% degree of freedom.

### Results and Discussion

All the 24 colocasia genotypes collected from different regions of entire Chhattisgarh state were screened for disease incidence against colocasia leaf blight (*Phytophthora colocasiae*) under *in-vivo* condition and results are presented in Table 1 and 2.

Table 1: Screening of colocasia genotypes for their reaction against leaf blight disease

Construes	Terminal per cent disease infection				
Genotypes	2018	2019	Mean	Score	
IGCOL-PITH-17-1	0.0 (0.0)*	0.0 (0.0)*	0.00	0	
IGCOL-KOTA-17-1	71.0 (57.4)	69.7(56.7)	70.35	4	
IGCOL-NGR-17-1	85.0 (67.3)	89.3 (71.7)	87.15	5	
IGCOL-SPL-17-1	70.7 (57.7)	74.3 (61.7)	72.53	4	
IGCOL-PWD-17-1	28.7 (31.7)	36.0 (35.4)	32.35	3	
IGCOL-PWND-17-1	95.3 (79.7)	98.7 (86.1)	97.00	6	
IGCOL-KS-10-1	19.3 (25.9)	18.3 (24.8)	18.80	2	
IGCOL-KS-13-2	21.3 (26.4)	16.0 (23.1)	18.65	2	
IGCOL-BL-12-1	29.0 (32.3)	27.0 (29.9)	28.00	3	
IGCOL-JS-12-1	28.7 (31.2)	28.7 (31.2)	28.70	3	
IGCOLS-BDL-17-1	10.3 (18.6)	12.7 (20.5)	11.50	2	
IGCOL-PKJ-17-1	0.0 (0.0)	0.0 (0.0)	0.00	0	
IGCOL-CHLN-17-1	0.0 (0.0)	0.0 (0.0)	0.00	0	
IGCOL-MNDG-17-1	13.7 (21.0)	12.7 (20.5)	26.40	3	
IGCOL-GB-17-1	26.0 (29.9)	27.7 (31.7)	26.85	3	
IGCOL- PWND-17-2	14.0 (18.1)	10.7 (15.7)	12.35	2	
IGCOL-BNR-17-1	26.3 (30.8)	26.3 (30.8)	26.30	3	
IGCOL-KDKN-17-1	34.7 (35. 1)	28.0 (31.2)	31.35	3	
IGCOL-MUNG-17-1	24.7 (27.9)	23.0 (30.1)	23.85	2	
White Gauriya (S.C.)	42.3 (40.5)	42.3 (40.5)	42.30	3	
IGCOL-DK-17-1	27.7 (31.5)	27.3 (30.9)	27.50	3	
IGCOL-CHHTD-18-1	23.7 (29.0)	21.3 (25.7)	21.50	2	
IGCOLS-CHMD-18-1	35.3 (36.3)	35.0 (36.1)	35.15	3	
IGCOLS-LHDD-18-1	35.7 (36.6)	35.3 (36.2)	35.50	3	
SEm±	4.65	4.30			
CD (0.05%)	13.29	12.28			

<sup>(\*</sup> figure in parentheses are arcsine values)

Among the twenty four genotype of colocasia, three genotypes (IGCOL-PITH-17-1, IGCOL-PKJ-17-1 and IGCOL-CHLN-17-1) showed least terminal per cent disease infection (0.00%) in both years (during 2018 & 2019). Likewise the average terminal per cent disease infection also 0.00% and were significantly superior over rest of the

genotypes. On an average it was followed by IGCOL-BDL-17-1 (11.50%), IGCOL-PWND-17-2 (12.35%) and IGCOL-MUNG-17-1 (23.85%) whereas, the highest terminal per cent disease was recorded in IGCOL-PWND-17-1 (97.00%) presented in Table 1.

**Table 2:** Terminal disease severity and their reaction to different genotypes of colocasia

Disease rating scale	Terminal disease severity (%)	PDI (Mean)	Name of genotypes	Reaction
		0.00	IGCOL-PITH-17-1	
0	0	0.00	IGCOL-PKJ-17-1	I
		0.00	IGCOL-CHLN-17-1	
1	1-10	-	-	R
		11.50	IGCOL-BDL-17-1	
		12.35	IGCOL- PWND-17-2	
2	11-25	18.80	IGCOL-KS-10-1	MR
		18.65	IGCOL-KS-13-2	
		21.50	IGCOL-CHHTD-18-1	

		23.85	IGCOL-MUNG-17-1	
3	26-50	26.30	IGCOL-BNR-17-1	MS
		26.40	IGCOL-MNDG-17-1	
		26.85	IGCOL-GB-17-1	
		27.50	IGCOL-DK-17-1	
		28.00	IGCOL-BL-12-1	
		28.70	IGCOL-JS-12-1	
		31.35	IGCOL-KDKN-17-1	
		32.35	IGCOL-PWD-17-1	
		35.15	IGCOL-CHMD-18-1	
		35.50	IGCOL-LHDD-18-1	
		42.30	White Gauriya (S.C.)	
4	51-75	70.35	IGCOL-KOTA-17-1	S
		72.53	IGCOL-SPL-17-1	
5	Above 75	87.15	IGCOL-NGR-17-1	HS
		97.00	IGCOL-PWND-17-1	

Pooled data showed various levels of reactions among the collected genotypes. Three genotypes (IGCOL-PITH-17-1, IGCOL-PKJ-17-1 and IGCOL-CHLN-17-1) were found having no reactions i.e. resistant, six genotypes (IGCOL-BDL-17-1, IGCOL- PWND-17-2, IGCOL-KS-10-1, IGCOL-KS-13-2, IGCOL-CHHTD-18-1 and IGCOL-MUNG-17-1) were exhibited moderately resistant, eleven genotypes (IGCOL-BNR-17-1, IGCOL-MNDG-17-1, IGCOL-GB-17-1, IGCOL-BL-12-1, IGCOL-DK-17-1, IGCOL-JS-12-1, IGCOL-KDKN-17-1, IGCOL-PWD-17-1, IGCOL-CHMD-18-1, IGCOL-LHDD-18-1 and White Gauriya (S.C.)) were recorded moderately susceptible and two genotypes viz., IGCOL-KOTA-17-1 and IGCOL-SPL-17-1 were observed to be susceptible while, two genotypes (IGCOL-NGR-17-1 and IGCOL-PWND-17-1) were found highly susceptible against Phytophthora leaf blight disease of colocasia under in-vivo condition exhibited in Table 2. Similar results were also reported by Kumar and Dubey (1996) [10] screened 15 genotypes against colocasia leaf blight disease. The two local varieties were found to be immune to the disease. Lokesh et al. (2018) [2] screened eleven germplasms of C. esculenta and X. sagittifolium were evaluated for their response to leaf blight along with plant characters whereas, one germplasm was found Colocasia esculenta Acc 3 is locally known by Makkalakesa and it is a good germplasm for screening of other germplasm for the disease reaction. Saykar et al. (2019) [11] screened 9 varieties for disease reaction against Phytophthora colocasiae. Among 9 varieties none of the variety was showed neither immune nor resistant. TCbl 12-2, TCbl 12-5 and TCbl 12-7 were the moderately resistant and TCbl 12-1, TCbl 12-4, TCbl 12-6 and Mukteshwari were susceptible and TCbl 12-3 and Telia were susceptible genotype. These findings are in agreement with findings of Shankar et al. (2012) [12] found that the per cent infection of Phytophthora leaf blight ranged from 2-20% in 145 genotypes of which 53 were free from Phytophthora leaf blight. Padmaja et al. (2016) [13] found that Variety KCS-3 showed moderately resistant reaction to all the isolates of Phytophthora colocasiae.

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