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Recent advances in preservation of boar semen in liquid state: An overview

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Abstract

Current century of artificial insemination (AI) of female pigs, knowledge about the preservation of boar semen is still very limited. In pig liquid semen preservation was mainly done at 15-20 °C for 5 days. The short-term and long-term extenders are used according to the time required for AI. In short-term extender both viability and fertility of semen was restored whether in long-term storage although the viability of semen was maintain but the fertility was extremely reduced with the time of preservation. The present study would try to give an overview about the preservation technique of boar semen in terms of extender, preservation time, preservation temperature, viability and fertility.

Keywords: Boar, semen preservation, method, extender, fertility

Introduction

Pigs are considered as scavenging animal and that the underprivileged are involved in pig production. Pig keeping is important in north-eastern states of India and particularly for the tribal communities [1]. Pig occupies a unique role among the meat-producing animals of the NE region. It is an animal of choice for meat especially to tribal population in northeast India as pig rearing is considered to be the most encouraging and appropriate livestock enterprise to narrow down the gap between the availability and requirement of animal meat in this part of the country and it also plays an important role in improving the socio-economic status of the weaker rural community and creates employment for the unemployed youth of the region [2]. According to the 19th livestock census, pig population in India stands at 10.29 million compared to world population of 977.02 million [3] which comprises 1.05% of world's pig population. Pig consisting of 2.01% of total livestock (512.05 million) in India and contribute only 0.95% of the global share in pig production [4].

The superior quality elite boar is exploited for artificial breeding of female pigs to maximum possible extent by artificially inseminating large number of female animals merely from a single ejaculate [5]. To achieve the said goal, extender of good quality is required for extending the boar semen which plays an crucial role in improving the post thaw semen quality in terms of spermatozoa viability, motility, plasma membrane and acrosomal integrity, mitochondrial membrane potential (MMP) etc. [5].

The temperature of semen storage is an important factor in retaining the fertilizing capacity of boar spermatozoa [6]. In modern piggery industry, the artificial insemination (AI) of sows is mostly based on liquid cooled semen [7, 8]. For preservation of liquid boar semen, the temperature 16-20 °C is still common practice as sperm cryopreservation remains suboptimal in this species [9]. To preserve spermatozoa for prolonged periods, their metabolic activity needs to be reduced by diluting it in an appropriate medium and lowering the temperature [8]. As boar spermatozoa very sensitive to cold shock, the boar sperm alters the viability [10]. Storage time lower than 72 h, it is preferable to use a short-term extender and more than 4 days is recommended to use a long-term extender on a higher sperm concentration to compensate for reduced sperm viability due to ageing [11]. Therefore, an assessment of integrity of the sperm membrane of diluted semen is important for determining which extender to use and how long the semen can be stored [12].

Preserving of Boar Semen in Liquid State

Boar spermatozoa are particularly sensitive to cold shock, which induces changes in the plasma membranes of spermatozoa, resulting in a reduction in their biological properties [13]. The effect of cold shock on spermatozoa is dependent on the rate of their cooling, the time and temperature range of storage, as well as the type of applied extender [14]. Extended boar semen is cooled to between 15° and 18°C and is maintained at this temperature for several days until use for AI [8, 15, 16]. Semen was preserved in 1.5 ml micro-centrifuge tubes at 15°C in BOD incubator up to 72 h [17].

Extender Used for Preservation of Boar Semen in Liquid State

The fertilizing capacity of the stored boar semen depends mainly on fresh semen quality, spermatozoa dose used for insemination, and composition of extender [18]. The progressive motility is the most commonly used indicator of both viability and fertility of boar semen, though a high level of motility does not guarantee better reproduction results [7, 19]. Spermatozoa maintain their metabolism and motility, principally through glycolytic pathways [20]. Any extender must contain an energy source, which is often glucose, as other sources such as galactose, ribose and trehalose have been reported to give worse results [11]. Different combinations of sugars (glucose) and non-sugars energy substrates (citrate and lactate) during a 7-day period of storage at 17 °C give suitable results [21]. A practical level and for current production purposes, diluents can be divided into two major groups: Diluents for short-term preservation (1-3 days) and diluents for long-term preservation (over 4 days). For example of short-term diluents are- Beltsville Liquid (BL-1), Beltsville Thawing Solution (BTS), Illinois Variable Temperature (IVT), Kiev, Vital etc. The long-term diluents are- Acromax, Androhep, Modena, MR-A, Mullberry III, Reading, X-cell, Zorlesco, ZORPVA etc. [8]. The first of the so-called long-term diluents was the Zorlesco medium (Gottardiet *et al.*, 1980).KOtOWO-3 extender was mostly used extender in Poland [22].

Many extenders are formulating to enhance preservation capabilities by providing compounds to protect the spermatozoa against cold shock [6]. Three different commercial extenders devised for short-term (BTS+) or long-term preservation (MR-A and X-Cell), were used to test the storage of semen from four mature, fertile boars at 17 °C for 96 hours [9]. The Safe cellplus extender is better than BTS to preserve boar semen in the liquid state at 16-18 °C in Large White Yorkshire boars [23].

Additives Used for Preservation of Boar Semen in Liquid State

Additives which impart anti-oxidant, sperm membrane stabilization properties and sperm motility enhancer are deliberately used with semen extender for better preserving quality. A number of additives have been studied over the past few years and their effect on improving the quality of boar semen. The additives like antioxidants, sugars, cholesterol and proteins, are effective for improving the fertilizing ability even in post thaw stage [5].

Antioxidants are the agents, which break the oxidative chain reaction-eliminating, taking up, or reducing the formation of ROS [24] and thereby reduce the oxidative stress [25, 26]. Based on their chemical property, antioxidants may also be categorized as enzymatic (e.g. glutathione reductase or, GSH,

SOD, and catalase), and non-enzymatic (e.g. vitamins C, E, and B, carotenoids, carnitine, cysteines, pentoxifylline, metals, taurine, hypotaurine, and albumin) [24]. BSA was included in the extenders as they maintain sperm motility, membrane integrity and reproductive performances [27].

Funahashia and Sano [28] reported that addition of glutathione and cysteine can improve the viability and functional status of boar spermatozoa during liquid preservation and boar spermatozoa penetrated *in vitro* even after preservation in the presence of cysteine at 10 °C for 29 days.

Polyunsaturated fatty acids and antioxidants supplementation of boars had a beneficial effect on the biological characteristics of the spermatozoa, which could be useful for semen preservation at different temperatures [29]. Heparin induces *in vitro* capacitation changes in the boar spermatozoa as evident by highest hyper activation of spermatozoa at 4 hours of incubation in crossbred Hampshire boar as reported by Haque *et al.* [30] and Talukdar *et al.* [31]. Addition of lipoprotein fractions in semen extender enhanced cold shock resistance of boar spermatozoa, particularly during storage at 5 °C [7].

Temperature and Storage of Preserved Liquid Boar Semen

Buhr *et al.* [32] stored boar semen at 25 °C for 160 minutes. Strzezek *et al.* [29] had preserved the Hampshire boar semen diluted with Kortowo-3 extender at 5 °C and 16°C for 5-6 days. Althouse *et al.* [6] preserved the boar semen after diluting with Androhep extender at 5 different storage temperatures of 8, 10, 12, 14 and 17 °C for 48 hours. Perez *et al.* [33] incubated boar semen in a water bath at 37 °C for 2 hours. Kumaresan *et al.* [34] have used BTS for preservation of Hampshire boar semen at 18°C up to 96 hours. Kanshi *et al.* [35, 36] have used Lactose egg yolk (LEY) dilutor for preservation of boar spermatozoa at 15°C in a BOD incubator upto 96 hours. Khan and Kumar [37] have used BTS solution for boar semen preservation at 18 °C up to 120 hours in 24 mature fertile boars of four different genetic groups (HS, CB-50%, CB-75% and CB-87.5%). The BTS, Kiev and Modena extender was used by Gobindasamy *et al.* [38] for preservation of Hampshire boar semen at 17°C up to 5 days. Patra *et al.* [39] preserved the Large White Yorkshire, Tamworth and T & D (cross of Tamworth and Desi) semen up to 72 hours by using BOD incubator. By using Modena extender, Chutia *et al.* [17] have used GEPS extender for preservation of boar semen at 15 °C for 72 hours. Reddy *et al.* [40] have used Farmer friendly Boar Semen Extender (FBSE) stored at refrigerator temperature (5-8 °C) and Beltsville Thawing Solution (BTS) stored at BOD incubator (15-18 °C) for 24 hours of Large White Yorkshire boar semen. Karunakaran *et al.* [23] preserved the semen of Large White Yorkshire boar by using BTS and SAFE cell plus extender at 16-18°C over a period of 7 days. Haque *et al.* [41] preserved the Hampshire boar semen at 15 °C up to 120 hours.

Fertility of Preserved Liquid Boar Semen

We should consider that the relationship between semen quality (which the diluent preserves) and resultant fertility of semen [11]. While comparing between the diluents used for short-term and long-term, there is no significant differences in fertility or litter size in the first 3-4 days of storage [42]. The Kiev extender provides better fertility than BL-1 after 1 day (74.5% vs 64.7%) or 3 days (65.9 vs. 60.5%) of storage [43]. The fertilization rate with Androhep and control was 82% and 81% respectively [16]. Frangez *et al.* [44] reported that there is

no significant difference in fertility rate between BTS and Merck extender. The diluent MR-A was able to maintain the same fertility rate and number of live births using semen stored for up to 5 days as reported by Martinez *et al.* [45]. Subsequently, similar fertility rate was found when compared the diluents BTS, MR-A and Modena [18]. No difference was recorded in terms of fertility when compared the diluent Androhep, BW25, or Kiev stored for 3 or 5 days [46]. The diluents MR-A, BTS, Modena and Androhep in comparison were found that there are no significant differences in fertility neither among the diluents nor according to the time of preservation (1-2 days vs. 3-4 days) as reported by Laforest and Allard [47].

Conclusion

After almost a century of artificial insemination in swine, knowledge about the preservation of boar semen is still very limited. The demands for future studies should be taken place with the following directions. The design of new diluents has been based on an empirical model, and the new models are needed to be evaluated and optimized with its components. There is a possibility of preservation of boar semen at temperatures below than 15°C would help to reduce the metabolic activity and protect against the detrimental effects of microbial contamination. Deep intrauterine insemination technique would change the scenario of preservation such that very less spermatozoa per dose and reduced insemination volume could require new preservation conditions and better diluent need to be assessed. The choice of diluent should depend on its proposed use. If the time of collection and insemination is less than 3 days then short-term diluent preferred or if more than 4 days then long-term diluent is mostly the choice. But the precaution will be taking care that any loss of sperm viability, fertility and litter size should not affect much every possible effort should be made to ensure the best semen quality. So the choice of dilution should be always aimed to optimize the subsequent effects in a particular condition of each farm.

Reference

1. Rangnekar DV. In: Livestock in the livelihoods of the underprivileged communities in India: A review. Published by International Livestock Research Institute, Nairobi, Kenya, 2006, 3-6.
2. Rahman S, Barthakur S, Kalita G. Pig production and management system in Aizawl district of Mizoram, India. Health care. 2008; 95:5.
3. Statistical Year Book. Food and Agriculture Organization of the United Nations. Regional Office for Asia and the Pacific, Bangkok, 2014.
4. Talukdar P, Talukdar D, Sarma K, Saikia K. Prospects and Potentiality of Improving Pig Farming in North Eastern Hill Region of India: An Overview. International Journal of Livestock Research. 2019; 9(1):1-14.
5. Hussain M, Begum SS, Kalita MK, Ahmed K, Nath R. Additives used in semen preservation in animals: A short review. International J Chemical Studies. 2018; 6(5):354-361.
6. Althouse GC, Wilson ME, Kuster C, Parsley M. Characterization of lower temperature storage limitations of fresh-extended porcine semen. Theriogenology. 1998; 50(4):535-543.
7. Fraser L, Strzezek J. The use of comet assay to assess DNA integrity of boar spermatozoa following liquid preservation at 5 degrees C and 16 degrees C. Folia Histochemica ET Cytobiologica. 2004; 42(1):49-55.
8. Pipan MZ, Mrkun J, Kosec M, Nemec Svete A, Zrimsek P. Superoxide dismutase: A predicting factor for boar semen characteristics for short-term preservation. Bio Med Research International, 2014, <https://doi.org/10.1155/2014/105280>
9. Ambrogi M, Ballester J, Saravia F, Caballero I, Johannisson A, Wallgren M, Rodriguez Martinez H. Effect of storage in short and long term commercial semen extenders on the motility, plasma membrane and chromatin integrity of boar spermatozoa. International Journal of Andrology. 2006; 29(5):543-552.
10. Pursel VG, Johnson LA, Schulman LL. Fertilizing capacity of boar semen stored at 15 °C. Journal of Animal Science. 1973; 37(2):532-535.
11. Gadea J. Semen extenders used in the artificial insemination of swine. Spanish Journal of Agricultural Research. 2003; 1(2):17-27.
12. Frydrychova S, Cerovsky J, Lustykova A, Rozkot M. Effects of long-term liquid commercial semen extender and storage time on the membrane quality of boar semen. Czech Journal of Animal Science. 2010; 55(4):160-166.
13. Johnson LA, Weitze KF, Fiser P, Maxwell WMC. Storage of boar semen. Animal Reproduction Science. 2000; 62(1-3):143-172.
14. Paulenz H, Adnøy T, Fossen OH, Soderquist L, Berg KA. Effect of deposition site and sperm number on the fertility of sheep inseminated with liquid semen. Veterinary Record. 2002; 150(10):299-302.
15. Perez-Llano B, Lorenzo JL, Yenes P, Trejo A, Garcia-Casado P. A short hypoosmotic swelling test for the prediction of boar sperm fertility. Theriogenology. 2001; 56(3):387-398.
16. Ardon F, Dohring A, Le Thi X, Weitze KF, Waberski D. Assessing *in vivo* Fertilizing Capacity of Liquid-Preserved Boar Semen According to the 'Hanover Gilt Model'. Reproduction in Domestic Animals. 2003; 38(2):161-165.
17. Chutia T, Biswas RK, Tamuli MK, Deka BC, Sinha S, Goswami J, Kayastha RB. Effect of holding of semen and washing of seminal plasma on quality and fertility of Hampshire boar semen preserved at liquid state. Animal Reproduction Science. 2014; 145(3-4):141-149.
18. Johnson LA, Aalbers JG, Grooten HG. Artificial Insemination of Swine: Fecundity of Boar Semen Stored in Beltsville TS (BTS), Modified Modena (MM), or MRA and Inseminated on One, Three and Four Days after Collection 1 2. Reproduction in Domestic Animals. 1988; 23(2):49-55.
19. Kuster CE, Althouse GC. The fecundity of porcine semen stored for 2 to 6 days in Androhep® and X-CELL™ extenders. Theriogenology. 1999; 52(3):365-376.
20. Rodriguez-Gil JE. Biological aspects of the mature boar spermatozoon. In: Boar reproduction, Springer, Berlin, Heidelberg, 2013, 49-64.
21. Medrano A, Pena A, Rigau T and Rodriguez Gil JE. Variations in the proportion of glycolytic/non glycolytic energy substrates modulate sperm membrane integrity and function in diluted boar samples stored at 15-17 °C. Reproduction in Domestic Animals. 2005; 40(5):448-453.
22. Strzezek J, Fraser L, Holody D, Wysocki P. Biochemical properties and usefulness of boar semen for liquid

- preservation following atropine administration. *Journal of Veterinary Medicine Series A*. 1998; 45(1-10):459-470.
23. Karunakaran M, Chakurkar EB, Ratnakaran U, Naik PK, Mondal M, Mondal A, Singh NP. Characteristics of boar semen preserved at liquid state. *Journal of Applied Animal Research*. 2017; 45(1):217-220.
24. Bansal AK, Bilaspuri GS. Impacts of oxidative stress and antioxidants on semen functions. *Veterinary Medicine International*, 2011, doi:10.4061/2011/686137.
25. Miller JK, Brzezinska-Slebodzinska E, Madsen FC. Oxidative stress, antioxidants, and animal function. *Journal of Dairy Science*. 1993; 76(9):2812-2823.
26. Kumar H, Mahmood S. The use of fast acting antioxidants for the reduction of cow placental retention and subsequent endometritis. *The Indian Journal of Animal Sciences*, 2001, 71(7).
27. Zhang XG, Liu Q, Wang LQ, Yang GS, Hu JH. Effects of glutathione on sperm quality during liquid storage in boars. *Animal Science Journal*, 2016, Doi: 10.1111/asj.12545.
28. Funahashi H, Sano T. Select antioxidants improve the function of extended boar semen stored at 10° C. *Theriogenology*. 2005; 63(6):1605-1616.
29. Strzezek J, Fraser L, Kuklinska M, Dziekonska A, Lecewicz M. Effects of dietary supplementation with polyunsaturated fatty acids and antioxidants on biochemical characteristics of boar semen. *Reproductive Biology*. 2004; 4(3):271-287.
30. Haque A, Ahmed K, Kalita D, Tamuly S, Talukdar D. *In Vitro* Capacitation of Boar Spermatozoa: Role of Heparin. *International Journal of Livestock Research*, 2019; 9(3):112-118. Doi:10.5455/ijlr.20180720060946.
31. Talukdar DJ, Ahmed K. and Talukdar Papor. Cryo capacitation and fertility of cryopreserved semen. *International Journal of Livestock Research*. 2015; 5(6):11-18. Doi: 10.5455/IJLR.20150608041222.
32. Buhr MM, Canvin AT, Bailey JL. Effects of semen preservation on boar spermatozoa head membranes. *Gamete research*. 1989; 23(4):441-449.
33. Perez-Llano B, Yenes-Garcia P, Garcia-Casado P. Four subpopulations of boar spermatozoa defined according to their response to the short hypoosmotic swelling test and acrosome status during incubation at 37 °C. *Theriogenology*. 2003; 60(8):1401-1407.
34. Kumaresan A, Kadirvel G, Bujarbaruah KM, Bardoloi RK, Das A, Kumar S *et al.* Preservation of boar semen at 18 C induces lipid peroxidation and apoptosis like changes in spermatozoa. *Animal Reproduction Science*. 2009; 110(1-2):162-171.
35. Kanshi S, Kumar R, Singh MP, Sinha MP, Singh SK. Effect of diluters, breeds and storage period on boar semen characteristics. *Indian Journal of Animal Sciences*. 2012; 82(8):826.
36. Kanshi S, Kumar R. Effect of dilutors, breed and preservation time on extracellular enzymatic activity of preserved boar semen. *Journal of Experimental Biology and Agricultural Sciences*. 2016; 4:53-58.
37. Khan MH, Kumar S. Preservation of liquid boar semen: Effect of genotype, boar and sperm parameters on motility and acrosome integrity. *Veterinary Research*. 2015; 3(2):30-34.
38. Govindasamy K, Ponraj P, Thulasiraman S, Andonissamy J, Naskar S, Das A *et al.* Efficacy of different extenders on sperm characteristics and fertility in crossbred pigs of north-eastern India. *Veterinarskiarhiv*. 2016; 86(4):515-528.
39. Patra MK, Kent Y, Soya R, Ngullie L, Nakhro R, Deka BC. Performance appraisal of artificial insemination technique in pig under organized farm and field condition in Nagaland. *Indian Research Journal of Extension Education*. 2014; 14(4):55-60.
40. Reddy NV, Muralimohan K, Reddy KR, Latha C, Reddy KC, Sushma K. Fertility results of Artificial Insemination in swine performed with liquid boar semen stored at different temperatures: A comparative study. *The Pharma Innovation*. 2017; 6(7):377.
41. Haque A, Ahmed K, Kalita D, Talukdar D. Assessment of sperm viability in extended boar semen during long term storage at 15 °C. *Theriogenology Insight*. 2018; 8(1):15-19.
42. Ratto J, Jokinen L. Reports about number of swine inseminations and farrowing results in Finland 1989, comparison between two diluents EDTA and MR-A. *Reproduction in Domestic Animals*. 1990; 1:365-368.
43. Johnson LA, Aalbers JG, Willems CMT, Rademaker JHM, Rexroad CE. Use of boar spermatozoa for artificial insemination III. Fecundity of boar spermatozoa stored in Beltsville liquid and Kiev extenders for three days at 18° C. *Journal of Animal Science*. 1982; 54(1):132-136.
44. Frangez R, Gider T, Kosec M. Frequency of boar ejaculate collection and its influence on semen quality, pregnancy rate and litter size. *Acta Veterinaria Brno*. 2005; 74(2):265-273.
45. Martinez EA, Vazquez JM, Roca J, Lucas X, Gil MA, Parrilla I *et al.* Minimum number of spermatozoa required for normal fertility after deep intrauterine insemination in non-sedated sows. *Reproduction-Cambridge*. 2002; 123(1):163-170.
46. Weitze K. The use of long-term extenders in pig AI - a view of the international situation. *Pig News and Information*. 1990; 11:23-26, ISSN 0143-9014.
47. Laforest JP, Allard D. Comparison of four extenders for long term storage of fresh boar semen. *Reproduction in Domestic Animals*. 1995; 31(1):275-276.