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Effect of phosphorus, Zinc and Mycorrhizae application on growth and nutrient uptake in capsicum under protected conditions

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Abstract

The present pot culture experiment was conducted to see the effect of Phosphorus, Zinc and Mycorrhizae on capsicum in net house, Department of Soil Science and Water Management, Dr YSPUHF Nauni-Solan during the year 2016. The experiment consists of twenty four treatment combinations with four levels of phosphorus (Pa, PB, Pc and Pd) equivalent to 0, 237.5, 355.5 and 475 kg ha⁻¹ Single Super Phosphate, three levels of Zinc (Zna, ZnB and Znc) equivalent to 5, 7.5 and 10 kg ha⁻¹ Zinc Sulphate and two levels of Arbuscular Mycorrhizal Fungi Inoculation (Ia and IB i.e. 0 and 15 g per pot). The results revealed that the plant height, root length, above ground nutrient total uptake as well as below ground nutrient total uptake by plant increases with increase in dose of Phosphorus (from 0 to 475 kg ha⁻¹), Zinc (from 5 to 10 kg ha⁻¹) and Mycorrhizal Inoculation (from 0 to 15 g per pot). Along with this total Nitrogen content in aboveground portion and below ground portion also increases with increase in application of P, Zn and Mycorrhizae. The distribution of total Phosphorus content however remained variable in above ground portion but in below ground portion, it increases with increase in application of P, Zn and AM inoculation. The potassium content remains variable in both above ground and below ground portion of the plant.

Keywords: Phosphorus, zinc, arbuscular mycorrhizae, antagonistic interaction, capsicum

Introduction

Capsicum annuum L. commonly known as capsicum or bell pepper is a domesticated species of the plant genus Capsicum of family Solanaceae. Capsicum is an important crop in mid hills of India. It is an economically beneficial crop to the farmers of the state which is cultivated over area of about 2,260 ha with an annual production of 3,900 tonnes (Anonymous, 2013) [3]. Macronutrients as well as micronutrients plays a significant role in whole development process of the plant, the root development, formation of ATP and various functions are carried out by Phosphorus (Shaheen et al. 2007) [22]. Whereas micronutrients like Zinc is involved in carbohydrates protein metabolism and sexual fertilization (Cakmak et al. 1989) [7] Phosphorus and Zinc are the two most essential nutrients which are responsible for the growth and development of plant. Both of these nutrients are antagonistic mutually in some cases, further leads to reduction in yield as well as nutrient uptake in many crops mainly due to Phosphorus or Zinc deficiencies (Bhardwaj et al. 2019) [4]. In plants, the interactions which are widely investigated is Phosphorus-Zinc interactions (Marschner, 1995) [18]. These interactions may be of two types whether increasing P application decreases/does not decrease Zinc concentration in the plant (Floneragan and Webb, 1993) [9]. Increase in application of P is likely to reduce Zn concentration in grains and inplant (Buekert et al. 1998) [5]. The higher P application and lower Zinc application increases P concentration in plant, which leads to P toxicity contributing to the symptoms resembling Zn deficiency (Loneragan et al. 1982) [15]. Translocation of Zinc from roots to the upper parts of plant can be impaired by enhanced level of P supply. In the roots of plant, high P concentration may causes Zinc to be bind within the root cell and thus remains unavailable for the plant. Whereas in case of high concentration of zinc in the roots results in formation of soluble Zn phosphates which is further responsible for unequal distribution of zinc between roots and upper parts of the plant (Cakmak and Marschner, 1987)

In order to alleviating Zn deficiencies or any other nutrient deficiency, mycorrhizae can be used which not only improves the nutritional status of the plant but also promotes P or Zn nutrition of the plant independently (Bhardwaj *et al.* 2019) ^[4]. Similarly it has also been reported that benefits of mycorrhizal association were more when plants grow under P/Zn deficiency. Also soils in mid hills conditions of the state are high in phosphorus which can affect the uptake of zinc in the plant. AM colonization in such conditions is expected to improve the availability of micronutrients. Keeping this in view the present study was carried out to see the effect of phosphorus and zinc in plant nutrition and mycorrhizae on growth and nutrient uptake in capsicum.

Materials and Methods Site description

The study was carried out in mid hills of Himachal Pradesh and the experimental site was situated in net house of Department of Soil Science and Water Management, Dr Y S Parmar University of Horticulture and Forestry Nauni, Solan, Himachal Pradesh. It is located at 30°52′ North latitude and 77°11′ East longitude and elevation of 1175 m above mean sea level.

Climate and Soil

The climate in which experiment was carried out lies in the sub-tropical zone but is slightly biased towards temperate climate. The soil of the experimental area falls in order Inceptisols (USDA Soil taxonomy), near neutral in reaction, medium in available N, K and high in available P, Zn, Cu, Mn, but deficient in Fe.

Plant material and potting mixture

One month old capsicum seedlings were selected for conducting the present studies. The seedlings were grown in pots filled with soil taken from field.

Treatment details

There were 24 treatment combination and details of treatments are given as follow:

1. P- LEVELS (Four levels)

- P_A: No application of P
- P_B: 237.5 kg ha⁻¹ SSP (50% of P)
- P_C: 355.5kg ha⁻¹ SSP (75% of P)
- P_D: 475 kg ha⁻¹ SSP (100% of P)

2. Zn - LEVELS (Three levels)

- Zn_A : 5 kg $ZnSO_4 ha^{-1}(50\% \text{ of } Zn)$
- \mathbf{Z} n_B: 7.5 kg ZnSO₄ ha⁻¹(75% of Zn)
- $Zn_{C:}$ 10 kg $ZnSO_4$ ha⁻¹(100% of Zn)

3. Inoculation with AM- Fungal Consortia: I_A : Inoculated and I_B : Uninoculated

AM fungal consortia were applied @15 grams pot⁻¹ by layering method (Jackson *et al.* 1972) $^{[12]}$. N and K application along with FYM to be applied uniformly at the recommended rates to all the treatments were applied by calculating the dozes per 10 kg of soil in the pots.

Table 1:	Treatment	Combinations:	$4 \times 3 \times 2 = 24$
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Treatment combinations without mycorrhiza	Treatment combinations with mycorrhiza
Treatment _{1:} P _A Zn _A I _A	Treatment _{13:} P _A Zn _A I _B
Treatment _{2:} P _A Zn _B I _A	Treatment _{14:} P _A Zn _B I _B
Treatment _{3:} P _A Zn _C I _A	Treatment ₁₅ : P _A Zn _C I _B
Treatment ₄ : P _B Zn _A I _A	Treatment ₁₆ : P _B Zn _A I _B
Treatment _{5:} P _B Zn _B I _A	Treatment _{17:} P _B Zn _B I _B
Treatment ₆ : P _B Zn _C I _A	Treatment _{18:} P _B Zn _C I _B
Treatment7: PcZnAIA	Treatment ₁₉ : P _C Zn _A I _B
Treatment ₈ : P _C Zn _B I _A	Treatment _{20:} P _C Zn _B I _B
Treatment9: PcZncIA	Treatment ₂₁ : P _C Zn _C I _B
Treatment ₁₀ : P _D Zn _A I _A	Treatment _{22:} P _D Zn _A I _B
Treatment ₁₁ : P _D Zn _B I _A	Treatment _{23:} P _D Zn _B I _B
Treatment ₁₂ : P _D Zn _C I _A	Treatment ₂₄ : P _D Zn _C I _B

Plant growth analysis

The plant growth parameters like plant height, root length was measured at the end of crop season and measured in centimeters. Total Nitrogen of plant samples (above ground portion and below ground portion) was determined by microkjeldahl method as described in A.O.A.C (1980) [1], total Phosphorus content was determined by vanado-molybdo yellow colour method (Jackson, 1973) [11]. Total Potassium content was determined by flame photometric method. Micronutrients like Fe, Mn, Zn and Cu were determined in the extract using Atomic Absorption Spectrophotometer.

Soil analysis

Composite soil samples were collected from pots and analyzed by standard methods. The organic carbon was estimated by rapid titration method (Wakley and Black, 1934) ^[26]. Available N, P and K in soil were determined by Alkaline Potassium permanganate method (Subbiah and Asija, 1956) ^[24]; Ammonium molybdate method using olsen's extractant (Olsen *et al.* 1972) ^[20] and Ammonium Acetate method (Merwin and Peech, 1951) ^[19] respectively. The DTPA extractable Fe, Mn, Zn and Cu were estimated on Atomic

Absorption Spectrophotometer (Lindsay and Norwell, 1978)

Results and Discussions Above ground parameters

As depicted in Table 2, the two-way interaction shows that P×Zn and I×Zn interaction was non-significant however the I_BP_D interaction recorded significantly higher plant height (72.5cm). These results are in accordance with the results of Cwala et al. (2010) [8] who observed that growth and development of capsicum plant was affected by commercial mycorrhizal inoculants. Among macronutrient content in above ground portion, in case of total nitrogen content, the P×Zn interaction was non-significant. The interaction I×Zn and I×P was significant with maximum value of total nitrogen content was observed in I_BZn_C and I_BP_D. These results are in line with the results obtained by Abbott and Robson (1984) who concluded that uptake and translocation of nitrogen increases with the application of AM fungi. The effect of P, Zn and Mycorrhizae on total phosphorus content was observed significant only for I×Zn and I×P interactions with maximum value recorded in I_BZn_A and I_BP_D treatment. These results are similar with those obtained by Tanwar and Aggarwal (2013) $^{[25]}$, who demonstrated that the dual inoculation and multi inoculation of bell pepper plants with *Glomus mosseae* had increased the phosphorus content. Also the in case of total potassium content, two way interaction P×Zn and I×P was significant with maximum value in $P_{\rm A}Zn_{\rm C}$ and $I_{\rm B}P_{\rm A}$ treatment and the interaction I×Zn was nonsignificant. These results are similar with the results found by Maksoud *et al.* (1994) $^{[6]}$ who observed that AM mycelium increases the potassium content in plants.

Among micronutrient content, all three interactions ($P \times Zn$, $I \times P$ and $I \times Zn$) were significantly affected with maximum value of total K content in $P_A Zn_A$, $I_B P_A$ and $I_B Zn_A$ treatments.

These observations are similar with the findings of Halder and Mandal (1981) [10], who reported that application of phosphorus causes a decrease in concentration of iron in the shoots. The two factor interaction was also significant with P_AZn_C recorded higher total Mn content. On the other hand, I_BZn_C and I_BP_A were maximum and significantly higher. The results are in line with those obtained by Halder and Mandal (1981) [10], who reported that application of phosphorus caused a decrease in the concentration of manganese in shoots also the application of zinc, increases the concentration of manganese in shoots (Cakmak and Marshner, 1987) [6] in a synergistic manner with Fe. In case of total Zn content in above ground parts of the plant, all three interactions were

Table 2: Effect of P, Zn and Mycorrhizae application on above ground parameters.

Treatments	Plant	NT (0/)	P	K	Fe	Mn	Zn	Cu	Total uptake
combinations	Height (cm)	N (%)	(%)	(%)	(mg kg-1)	(mg kg ⁻¹)	(mg kg ⁻¹)	(mg kg ⁻¹)	(g plant ⁻¹)
P_AZn_A	51.3	7.60	0.85	10.49	390.96	240.80	72.63	47.15	3.21
P_AZn_B	53.0	7.72	0.81	10.64	376.00	249.64	79.36	45.17	3.36
P_AZn_C	53.2	7.88	0.76	10.71	351.50	256.12	81.95	37.10	3.43
P_BZn_A	52.8	7.69	0.91	10.35	349.22	225.85	66.12	46.10	3.29
P_BZn_B	55.3	7.85	0.85	10.45	331.00	238.80	71.38	39.72	3.45
P_BZn_C	57.3	8.08	0.82	10.49	310.31	249.35	77.09	36.55	3.57
P_CZn_A	57.5	8.74	0.99	9.98	328.50	219.77	61.05	43.20	3.66
P_CZn_B	58.5	8.96	0.93	10.35	313.50	224.74	66.47	37.00	4.06
P_CZn_C	60.5	9.18	0.88	10.41	281.25	237.53	74.08	32.18	4.16
P_DZn_A	60.0	8.93	1.11	9.66	286.95	214.47	57.28	35.45	3.73
P_DZn_B	60.5	9.04	1.07	10.18	282.76	218.85	61.73	34.29	4.16
P_DZn_C	63.3	9.25	1.03	10.31	267.75	231.92	67.67	32.08	4.28
Mean	56.9	8.41	0.92	10.33	322.47	233.98	69.73	38.83	3.70
CD 0.05	NS	NS	NS	0.13	8.21	1.52	1.46	0.50	0.09
I_AZn_A	44.4	7.50	0.86	9.59	298.42	221.98	59.58	41.98	3.13
I_AZn_B	45.8	7.65	0.82	9.85	281.63	227.87	64.74	38.08	3.28
I_AZn_C	47.2	7.90	0.80	9.90	250.40	238.31	72.30	33.95	3.40
I_BZn_A	66.4	8.98	1.07	10.64	379.39	228.46	68.96	43.98	3.81
I_BZn_B	67.9	9.13	1.01	10.96	370.00	238.14	74.72	40.01	4.24
I_BZn_C	69.9	9.29	0.94	11.06	355.00	249.15	78.09	35.00	4.33
Mean	56.9	8.41	0.92	10.33	322.47	233.98	69.73	38.83	3.70
CD 0.05	NS	0.06	0.02	NS	5.81	1.07	1.04	0.36	0.06
I_AP_A	42.5	7.20	0.72	10.03	319.79	243.10	73.81	42.01	3.00
I_AP_B	43.3	7.30		9.87	284.68	234.49	66.73	39.47	3.06
IaPc	47.3	8.04	0.81	9.71	268.33	222.53	63.02	37.47	3.48
I_AP_D	50.0	8.19	1.01	9.51	234.47	217.42	58.59	33.07	3.54
I_BP_A	62.5	8.26	0.89	11.19	425.86	254.60	82.14	44.27	3.67
I_BP_B	66.8	8.45	0.95	10.99	375.67	241.51	76.33	42.11	3.81
I _B P _C	70.3	9.87		10.78	347.17	232.16	71.37	37.45	4.44
I_BP_D	72.5	9.95	1.13	10.59	323.83	226.07	65.85	34.81	4.58
Mean	56.9	8.41		10.33	322.47	233.98	69.73	38.83	3.70
CD 0.05	1.5	0.07	0.02	0.11	6.71	1.24	NS	0.41	0.07

Significantly affected with maximum value of total Zn content observed in PAZnC, IBZnC and IBPA. These results are in line with the results of Ortas et al. (2011) who showed that plants inoculated with Glomusmosseae, G. intraradices,,G. etunicatum, G. clarum, G. caledonium and the mixture of these fungi had increased Zn content compared to control plants. The total Cu content in plants also recorded significant higher values in P_AZn_A, I_BP_A and I_BZn_A. Similar results were also recorded by Halder and Mandal (1981) [10] who found that P application decrease concentration of Cu in the shoots. The total nutrient uptake by plant in above ground portion also affected significantly with maximum value of total nutrient uptake recorded in PDZnC, IBZnC and IBPD. These results are in conformity with those of Srinivasan et al. (2012) who concluded that in crop plants, with inoculation of mycorrhizae there is increase in uptake of macro and micronutrients.

Below ground parameters

As given in Table 3, among below ground parameters, the maximum root length was observed in P₃Zn₂, I₁Zn₂ and I₁P₃. These results are same with those of Kim et al. (2010) [13] who that red pepper plants inoculated with Methylobacterium oryzae strains resulted in a significant increase in root length and root fresh weight compared to untreated control plants. Among macronutrient content of below ground portion (root), the total Nitrogen content was significantly affected with maximum value of total Nitrogen content in treatments PDZnC, IAZnC and IBPD. These are similar results with findings obtained by Kim et al. (2010) [13] who observed that mycorrhizal inoculation addition resulted in significantly higher nitrogen accumulation in the roots of red pepper plants as compare to control. In case of total phosphorus content, I×P and I×Zn interaction had nonsignificant effect on total P content. However P×Zn

interaction had significantly affect with maximum value total P content in P_DZn_C treatment. These results are in accordance with studies carried out by Kim *et al.* (2010) ^[13] who showed that the addition of AM fungi increased the phosphorus (P) content by 23.3% compared to untreated control. The total K content in below ground portion was significantly affected by application of P, Zn and mycorrhizae with maximum value of

total K observed in P_AZn_C , I_BZn_C and I_BP_A . These results are in accordance with results of Maksoud *et al.* (1994) [16] who found that arbuscular mycorrhizal mycelium influences the potassium content in plants. Among micronutrient content in roots, P_AZn_A , I_BZn_A and I_BP_A interaction shows significantly higher Fe content in roots. Similar results were also

Table 3: Effect of P, Zn and Mycorrhizae application on below ground parameters

Treatments	Root length (cm)	N	P	K	Fe	Mn	Zn	Cu	Total uptake
combinations	combinations Koot length (cm)	(%)	(%)	(%)	(mg kg-1)	(mg kg ⁻¹)	(mg kg-1)	(mg kg-1)	(g plant ⁻¹)
P_AZn_A	9.5	3.49	0.26		129.50	124.90	30.65	24.63	0.62
P_AZn_B	10.4	3.64	0.30	2.82	124.00	125.48	30.79	18.83	0.72
P _A Zn _C	11.2	3.75	0.34	2.89	119.70	127.73	35.20	14.95	0.78
P_BZn_A	10.0	3.49	0.29	2.49	121.48	124.10	34.60	14.64	0.72
P_BZn_B	11.1	3.73	0.32	2.74	118.95	124.10	35.79	14.05	0.78
P_BZn_C	11.7	3.85	0.38	2.80	115.13	127.45	37.46	14.20	0.90
P _C Zn _A	10.3	3.53	0.35	2.36	113.65	121.30	35.84	12.28	0.76
P_CZn_B	11.6	3.76	0.38	2.61	112.28	124.50	39.70	10.66	0.84
PcZnc	12.1	3.87	0.40	2.77	110.70	126.18	41.14	10.10	0.95
P_DZn_A	10.9	3.72	0.36	2.28	110.18	119.93	40.32	10.83	0.80
P_DZn_B	12.4	3.78	0.40	2.46	107.63	123.22	41.34	10.35	0.89
P_DZn_C	12.8	3.91	0.46	2.68	106.20	125.83	41.94	10.13	1.09
Mean	11.2	3.71	0.35	2.62	115.78	124.56	37.06	13.80	0.82
CD 0.05	NS	0.06	0.02	0.03	1.79	0.80	1.07	0.24	0.06
I_AZn_A	9.1	3.30	0.30	2.27	115.79	119.03	33.12	13.24	0.69
I_AZn_B	9.7	3.63	0.32	2.30	111.78	119.85	34.91	12.73	0.77
I _A Zn _C	10.2	3.86	0.37	2.36	108.95	121.66	36.17	11.59	0.83
I_BZn_A	11.2	3.81	0.33	2.56	121.61	126.09	37.58	17.94	0.76
I_BZn_B	13.1	3.82	0.37	3.02	119.65	128.80	38.90	14.21	0.85
I_BZn_C	13.7	3.83	0.42	3.20	116.91	131.93	41.70	13.10	1.03
Mean	11.2	3.71	0.35	2.62	115.78	124.56	37.06	13.80	0.82
CD 0.05	NS	0.04	NS	0.02	1.27	0.56	0.76	0.19	0.04
I_AP_A	8.8	3.48	0.27	2.36	117.97	121.27	29.70	17.73	0.61
I_AP_B	9.4	3.57	0.31	2.34	114.30	120.52	32.32	12.31	0.74
I _A P _C	9.9	3.60	0.36	2.30	110.30	120.15	36.59	10.07	0.82
I_AP_D	10.5	3.73	0.38	2.24	106.12	118.78	40.32	9.97	0.88
I_BP_A	11.9	3.77		3.14	130.83	130.80	34.72	21.20	0.81
I_BP_B	12.4	3.81	0.35	3.01	122.73	129.92	39.58	16.28	0.86
I_BP_C	12.8	3.84	0.38	2.85	114.12	127.83	41.19	11.95	0.88
I_BP_D	13.6	3.87	0.43		109.88	127.20	42.08	10.90	0.97
Mean	11.2	3.71	0.35	2.62	115.78	124.56	37.06	13.80	0.82
CD 0.05	NS	0.05	NS	0.02	1.46	0.65	0.88	0.22	0.05

reported by Halder and Mandal (1981) [10] who reported decrease in Fe concentration with increase in P application. Similarly the two factor interaction shows statistically higher values of Mn in roots of P_AZn_C, I_BZn_C and I_BP_A plants. These findings are in consonance with the findings of Halder and Mandal (1981) [10] who found a decrease in the concentration of manganese in roots with increasing P and Zn application. The two factor interactions were also found significant for P_DZn_C , I_BZn_C and I_BP_D interactions. These results are similar to those obtained by Olsen (1972) [20] who reported that high level of soil available P or with high Phosphorus application, Zinc in roots may accumulate and metabolic disorder at cellular level may occur due to Phosphorus and Zinc imbalance. The two factor interactions were also found significant for P_DZn_C, I_BZn_C and I_BP_D interactions. These results are similar to those obtained by Olsen et al. (1972) [20] who reported that high level of soil available P or with high Phosphorus application, Zinc in roots may accumulate and metabolic disorder at cellular level. In case of total Cu content in roots, two way interaction show PAZnA, IBZnA and IBPA interaction had significantly higher Cu content in the roots. The total Nutrient uptake by root in two factor interaction reveals that total nutrient uptake by root was recorded maximum in P_DZn_C, I_BZn_C and I_BP_D. These results are in

accordance with the results carried out by Marscher and Dell (1994) [17] who reported that the mycorrhizal infection enhances plant growth by increasing nutrient uptake.

Soil parameters

As represented in Table 4, I×P interaction and P×Zn interaction shows significant effect on OC whereas I×Zn interaction was non-significant. The organic carbon contents of soil increased significantly in with maximum values under mycorrhizae treated soils as compared to mycorrhizal uninoculated soils. Overall the results show that the Phosphorus and Zinc levels in association with inoculation with AM fungi show higher levels of organic carbon in the soils. Among macronutrient content in soil, two way interactions were significant with maximum soil N in P_CZn_C, I_BZn_C and I_BP_C. The results are in line with the values obtained for organic carbon status of these soils indicating better soil health at medium levels of application in combination with inoculation. In case of available P in soil, two factor interaction also exhibit high soil P in PDZnB, IBZnC and I_BP_D interactions. The results are in line with the findings of Yusnizar and Rahmawati (2014) $^{[27]}$ who also reported that a combination of mycorrhizae and phosphate can

N P K **Treatments Organic** Fe Zn Cu (mg kg-1) (mg kg-1) (mg kg-1) combinations Carbon (%) (mg kg-1) (mg kg-1) (mg kg-1) (mg kg-1) $P_A Z\underline{n_A}$ 1.13 152.00 38.96 106.48 2.29 5.11 1.62 0.60 39.83 5.13 2.54 0.74 P_AZn_B 1.18 155.18 110.00 1.64 5.25 $P_{A}Zn_{C} \\$ 1.18 162.58 40.53 112.00 1.68 2.67 0.83 2.53 1.19 163.23 39.99 109.50 5.13 1.70 0.68 P_BZn_A 5.16 1.22 167.07 42.20 111.75 1.72 2.86 P_BZn_B 0.81 1.28 175.50 44.34 111.75 5.19 1.78 3.06 0.84 P_BZn_C P_CZn_A 1.36 200.05 42.53 122.25 5.37 2.01 3.33 0.92 $P_{C}\underline{Z}\underline{n_{B}}$ 1.39 200.47 42.93 125.00 5.50 2.03 3.39 1.01 P_CZn_C 1.40 207.88 44.23 126.90 5.57 2.08 3.83 1.13 $P_D Z n_A \\$ 44.70 112.18 5.27 1.28 171.87 1.88 3.07 0.81 1.29 184.25 45.75 117.21 5.38 1.93 3.09 0.95 P_DZn_B 45.70 0.98 P_DZn_C 1.31 186.78 121.00 5.42 1.99 3.34 177.24 42.64 115.505.29 1.84 0.86 Mean 1.27 3.00 1.28 0.02 0.05 0.02 3.46 2.01 0.03 0.13 CD 0.05 165.86 40.45 105.14 5.07 1.56 0.77 I_AZn_A 1.16 2.85 $I_{A}Zn_{B} \\$ 1.18 169.03 41.48 110.73 5.09 1.61 2.91 0.85 $I_{A}Zn_{C}$ 1.20 172.42 43.31 113.38 5.15 1.68 3.10 0.88 I_BZn_A 1.32 177.72 42.64 120.06 5.37 2.04 2.76 0.74 I_BZn_B 1.36 184.45 43.88 121.25 5.50 2.05 3.03 0.90 5.56 122.45 2.08 1.39 193.96 44.08 3.34 1.00 I_BZn_C 1.27 177.24 42.64 115.50 5.29 3.00 0.86 1.84 Mean 2.45 0.90 1.42 0.04 CD 0.05 NS 0.02 0.02 0.09 1.11 148.42 39.26 103.73 4.93 1.34 2.37 0.72 $I_A P_A \\$ $I_{A}P_{B} \\$ 1.15 157.67 40.76 102.67 4.97 1.47 2.83 0.79 1.27 199.49 42.00 122.17 5.32 1.90 3.51 0.98 $I_{A}P_{C} \\$ 1.18 170.83 44.97 110.42 5.19 1.75 3.11 0.85 $I_{A}P_{D} \\$ $I_BP_A\\$ 1.22 164.76 40.29 115.25 5.39 1.95 2.63 0.73 $I_BP_B\\$ 1.31 179.53 43.59 119.33 5.35 1.99 2.81 0.76 1.50 206.11 44.45 127.27 5.64 2.18 3.52 1.06 I_BP_C 191.10 45.80 2.11 3.22 0.98 1.41 123.17 5.52 I_BP_D 5.29 Mean 1.27 177.24 42.64 115.50 1.84 3.00 0.86 CD 0.05 2.83 1.64 0.02 0.02 0.11 0.04

Table 4: Effect of P, Zn and Mycorrhizae application on soil parameters.

increase available P on growing media. Also I×P and P×Zn interaction had significant effect on available potassium. The trend was similar to that obtained in available N in soils. Similarly, among micronutrient content in soils, the two way interactions P×Zn, I×Zn and I×P were significantly affected by the application of P, Zn, mycorrhizae. The maximum value of available Fe, Mn, Zn and Cu were reported in $P_{\rm C}Zn_{\rm C}$, $I_{\rm B}Zn_{\rm C}$ and $I_{\rm B}P_{\rm D}$.

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References

- 1. AOAC. Official methods of analysis. Association of Official Analytical Chemists. Association of Analytical Chemists. Washington, DC, 1970, 101.
- Abbott LK, Robson AD. The effect of VA mycorrhizae on plant growth. In: Mycorrhizae, CL Powell and DJ Bagyaraj (eds.). CRC Press. Boca Raton, Florida, USA, 1970, 113-130.
- 3. Anonymous. Indian Horticulture Database, 2013, http://www.nhb.gov.in.
- Bhardwaj G, Sharma U, Brar PS. A Review on Interactive Effects of Phosphorous, Zinc and Mycorrhiza in Soil and Plant. International Journal of Current Microbiology and Applied Sciences 2019; 8(04):2525-2530.

- 5. Buekert A, Haake C, Ruckwied M, Marschner H. Phosphorus application affects the nutritional quality of millet grain in the Sahel. Field Crops Research. 1998; 57:223-235.
- 6. Cakmak I, Marschner H. Mechanism of phosphorusinduced zinc deficiency in cotton. III. Changes in physiological availability of zinc in plants. Plant Physiology. 1987; 70:13-20.
- 7. Cakmak I, Marschner H, Bangerth F. Effect of zinc nutrition status on growth, protein metabolism and levels of Indole-3-acetic acid and other phythohormones in Bean (*Phaseolus vulgaris* L.) Journals of Experimental Botany. 1989; 40(212):405-412.
- Cwala Y, Laubscher CP, Ndakidemi PA, Meyer AH. Mycorrhizal root colonization and the subsequent host plant response of soil less grown tomato plants in the presence and absence of the mycorrhizal stimulant. African Journal of Microbiology Research. 2010; 4:414-419.
- 9. Floneragan J, Webb MJ. Interaction between zinc and other nutrients affecting the growth of plants. In: Zinc in Soils and Plants. AD. Robson (ed.). Kluwer Academic Publishers, London, 1993, 121-125.
- 10. Halder M, Mandal LN. Effect of phosphorus and zinc on the growth and phosphorus, zinc, copper, iron and manganese nutrition of rice. Plant and Soil. 1981; 59:415-425.
- 11. Jackson ML. Soil chemical analysis. Prentice Hall of India Pvt. Ltd., New Delhi, 1973, 111-126.
- 12. Jackson NE, Franklin RE, Miller RH. Effects of vesicular-arbuscular mycorrhizal fungi on growth and

- phosphorus content of three agronomic crops. Soil Science Society of America Proceedings. 1972; 86:64-67.
- 13. Kim K, Yim W, Triwedi P *et al.* Synergistic effects of inoculating arbuscular mycorrhizal fungi *Methylobacterium oryzae* strains on growth and nutrient uptake of red pepper (*Capsicum annuum* L.). Plant and Soil. 2010; 327:429-440.
- 14. Lindsay WH, Norvell WA. Development of DTPA soil test for Zn, Fe, Mn and Cu. Soil Science Society of American Journal. 1978; 42:420-428.
- 15. Loneragan JF, Grunes DL *et al.* Phosphorus accumulation and toxicity in leaves in relation to zinc supply. Soil Science Society of America Journal. 1982; 46:345-352.
- Maksoud MA, Haggag LF, Azzazy MA, Saad RN. Effect of VAM inoculation and phosphorus application on growth and nutrient content (P and K) of *Tamarindus* indica L. Seedlings. Ann. Afric. Sci. Caito. 1994; 39:355-363.
- 17. Marschner H, Dell B. Nutrient uptake in mycorrhizal symbiosis. Plant and soil. 1994; 159:89-102.
- 18. Marschner H. Mineral nutrition of higher plants, Academic Press. London, 1995.
- 19. Merwin HD, Peech M. Exchange ability of soil potassium in the sand, silt and clay fractions as influenced by the nature and complementary exchangeable cations. Soil Science American Proceedings. 1951; 15:125-128.
- 20. Olsen SR, Cole CV, Watanable FS, Olsen SR. Micronutrient Interactions. In: Micronutrients in Agriculture. JM Mortved, PM Goirdano and WL Lindsay (eds.). SSSA, Madison, WI, 1972, 243-264.
- 21. Ortas I, Sari N, Akpinar C, Yetisir H. Screening mycorrhizae species for plant growth, P and Zn uptake in pepper seedling grown under greenhouse conditions. Scientia Horticulturae. 2011; 128:92-98.
- 22. Shaheen AM, Abdel MM, Ali AH, Rizk FA. Natural and chemical phosphorus fertilizers as affected onion plant growth, bulbs yield and its some physical and chemical properties. Australian Journal of Basic and Applied Sciences. 2007; 1(4):519-524.
- 23. Sreenivasan M, Ashwin R, Bagyaraj DJ. Symbiotic response of Amla (*Emblica officinalis* Gaertn.) to different arbuscular mycorrhizal fungi. Journal of Soil Biology and Ecology. 2012; 32:37-44.
- 24. Subbiah BV, Asija GL. Rapid procedure for the estimation of the available nitrogen in soils. Current Science. 1956; 25:259-260.
- 25. Tanwar A, Aggarwal A. Multifaceted potential of bioinoculants on red bell pepper (F1 hybrid, Indam Mamatha) production. Journal of Plant Interactions, 2013. doi:10.1080/17429145.2013.7 65044.66
- 26. Walkley A, Black TA. An estimation of soil organic matter and proposed modification of the chromic acid titration method. Soil Science. 1934; 37:29-38.
- 27. Yusnizar J, Rahmawati M. Adoption and inovation of technology for farmers and women group at Blang Krueng village through mycorrhiza propagation techniques to increase production of chili pepper. Report of Lpkm Unsyiah, Banda Aceh, 2014.