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In vitro efficacy of biocontrol agents and fungicides against anthracnose of french bean caused by *Colletotrichum lindemuthianum* (Sacc. and Magn.)

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Abstract

Bean anthracnose caused by *Colletotrichum lindemuthianum* (Sacc. and Magn.) is an important seed borne disease throughout the world. This pathogen overwinters on bean seed hence the losses can be 100%. Three fungicides alone namely Hexaconazole, Captan, Carbendazim and three in combinations i.e. Hexaconazole + *Bacillus cereus*, Captan + *Trichoderma harzianum* and Carbendazim + *Pseudomonas fluorescens* were used at different concentrations (100, 250, 500 and 750 ppm) by poisoned food technique. Alone Carbendazim and in combination, Carbendazim + *P. fluorescens* found complete inhibition of *C. lindemuthianum* at 100 ppm, while Captan at all concentration (52.48, 62.27, 68.05 and 69.75%) and in combination Hexaconazole + *B. cereus* (53.30, 63.98, 80.89 and 81.77%) were found minimum inhibition. Three bio control agents viz. *B. cereus*, *T. harzianum* and *P. fluorescens* were evaluated by dual culture technique, to check their antagonistic effect on the growth of *C. lindemuthianum*. Among bio agents *T. harzianum* observed (73.87%) maximum mycelium growth inhibition while *B. cereus* minimum (31.33%).

Keywords: French bean, anthracnose, *Colletotrichum lindemuthianum*, fungicides and bioagents

Introduction

French bean (*Phaseolus vulgaris* L.) is a leguminous vegetable belongs to the family Fabaceae. It is commonly known as Rajmash, haricot bean or common bean. French bean is considered to have originated in Mexico Maibam *et al.* (2015) [17]. Common bean (*Phaseolus vulgaris* L.) is a key grain legume crop and a vital source of nutrition worldwide. The FAO reports that half of the world's common bean production occurs in low income, food deficit countries where this staple crop contributes to food security. The other half is produced in countries like the U.S., where common bean is an important economic crop with 769 thousand ha of dry and snap beans planted in 2012, and with a farm gate value of \$1.5 billion (Anonymous, 2013) [3]. Yesuf (2005) [25] emphasized that diseases are known to be the major factors which threaten the productivity of common beans in general and common bean in particular. Anthracnose (*Colletotrichum lindemuthianum*), rust (*Uromyces appendiculatus*), angular leaf spot (*Phaeoisariopsis griseola*) and common bacterial blight (*Xanthomonas campestris* pv. *phaseoli*) are common diseases of bean in Ethiopia Habtu *et al.* (1996) [12]. The fungus *Colletotrichum lindemuthianum* is often present in or on the seed of the infected pods. Infected pods may show yellowish to brown sunken lesion (Agrios, 2005) [2]. Morphology of *C. lindemuthianum* causing anthracnose of bean and reported that, conidia were hyaline, single celled, dumbbell shaped born in acervuli bearing setae observed by Junaid *et al.* (2014) [13]. Plants at all stage are susceptible and susceptibility increase with age of the plant. Infection of a susceptible cultivar under favourable condition may results in 100% yield loss (Fernandez *et al.*, 2000) [9]. Heavy and frequent rains with moderate temperature (19-25°C) and high humidity (<70%) favours the progress of anthracnose disease (Aggarwal *et al.*, 2017) [1]. In fungicides Carbendazim and *Trichoderma* sp. in biocontrol agents were reported effective to reduce disease (Rajeshia *et al.*, 2010) [21]. Ten fungicides at 0.05%, 0.1% and 0.15% against *C. gloeosporioides* causes anthracnose of papaya and reported that Carbendazim inhibited (100%) per cent mycelium growth followed by Hexaconazole (93.50%) and Captan (84.09%) observed by Tasiwal *et al.* (2008) [22].

Keeping the above facts, the present investigation was carried out, some fungicides and bio agents against anthracnose of french bean caused by *Colletotrichum lindemuthianum*.

Materials and Methods

The present investigation was conducted in the laboratory of the Department of Plant Pathology, College of Horticulture, VCSG UHF Bharsar, Pauri Garhwal, Uttarakhand during 2017-18.

Collection and isolation of the pathogen: The infected pods of bean showing typical symptoms of anthracnose were collected from Vegetable Research and Demonstration Block, Bharsar. Cut into small pieces of (1-2 mm) size along with some healthy portion. These are surface sterilized with 70% ethanol and 0.1% mercuric chloride (HgCl_2), for about 30 seconds followed by three washings in sterilized distilled water. Then the segments were blotted on the blotter papers and transferred into Petri plates containing 20 ml of molten and cooled potato dextrose agar (PDA) and incubated at 28 ± 1 °C for 7 days. The incubated plates were observed periodically for fungal growth (Baudoin, 2011) [15]. Isolated pathogen transferred into sterilized Petri plates containing PDA medium and incubated at 24 ± 1 °C for 7 days Junaid *et al.* (2014) [13].

Purification, preservation and identification of pathogen: Pathogenic isolates from host plant species were isolated from the moist chambers as well as on PDA Petri plates. All the isolates were cultured under aseptic conditions in a laminar air flow and incubated at 28 ± 1 °C for 7 days. Culture purified from single colony appearing on PDA after observing under microscope and then maintained on PDA slants at 4 °C in a refrigerator for further use, sub culturing of the stored cultures was done periodically. The study was undertaken to confirm the identity of the isolated pathogen. Fungal growth, examined through the compound microscope, after making slides in lactophenol cotton blue stain. The shape and size of acervuli, conidia/spore; conidia hyaline, dumbbell, with obtuse ends and acervuli black in colour and saucer shaped. Microscopic observation was taken at (40 x) Junaid *et al.* (2014) [13].

In vitro study

The evaluation of fungicides and bio-agents alone and in combination against *C. lindemuthianum* were tested by, poisoned food technique (Nene and Thapliyal, 1993) [18]. Three fungicides alone namely Hexaconazole, Captan, Carbendazim and three in combinations i.e. Hexaconazole + *Bacillus cereus*, Captan + *Trichoderma harzianum* and Carbendazim + *Pseudomonas fluorescens* including control were used at different concentrations (100, 250, 500 and 750 ppm) with three replication and Petri plates were incubated at 28 ± 1 °C for 7 days.

Bio-agents by dual culture technique (Faheem *et al.*, 2010) [18] with five replications were maintained and incubated at 28 ± 1 °C, observed after 7 days. Three bio agents including control, namely *Bacillus cereus*, *Trichoderma harzianum* and *Pseudomonas fluorescens* were evaluated to check their antagonistic effect against *C. lindemuthianum*.

Per cent mycelium inhibition in each treatment over control was calculated with the following formula given by Vincent (1947) [23].

Where, I= Per cent inhibition, C=Radial growth in control, T= Radial growth in treatment,

The data had obtained and analyzed by using simple complete randomized design (CRD) with the help of OPSTAT and STPR3.

Results and Discussion

Three fungicides and three bio-control agents alone and in combination with control were evaluated (Table -1). In alone Carbendazim (100%) inhibition was found highly effective at all concentrations (100, 250, 500 and 750 ppm) followed by Hexaconazole showed the disease inhibition of (75.02, 77.29, 82.55 and 84.98%) respectively at all concentration. Captan at all concentration showed mycelia inhibition of (52.48, 62.27, 68.05 and 69.75%). Deshmukh *et al.* (2010) [7] also found that Carbendazim was found highly effective with maximum inhibition (100%) against anthracnose of bean. Patil *et al.* (2009) [20] tested among chemicals, Mancozeb + Carbendazim (0.2%) was found most effective in inhibiting 96.26 per cent growth of *C. gloeosporioides* followed by Carbendazim (0.1%) 68.34 per cent, Mancozeb and Copper oxychloride. Rajesha *et al.* (2010) [21] found among different systemic fungicides tested, Carbendazim inhibited cent per cent (100%) mycelial growth of *C. lindemuthianum* followed by propiconazole (100%) and difenoconazole (84.87%) at a concentration of 400 ppm. Gawade and Suryawanshi (2009) [11] evaluated five fungicides viz., Carbendazim, Chlorothalonil, Difenconazole, Hexaconazole and Propiconazole (each @ 100, 150 and 250 ppm.) against *C. truncatum* causing soybean anthracnose and reported that Carbendazim as a most effective with maximum mycelium inhibition (90.59%), followed by Propiconazole, Hexaconazole, Difenconazole and Chlorothalonil. Madhusudan (2002) [15] observed ten fungicides against *C. truncatum*, causing anthracnose of soybean. They reported that, Benomyl and Carbendazim inhibited per cent mycelium growth of the fungus at all the three concentrations tested (0.025, 0.05 and 0.1%), followed by the Combi fungicides Carbendazim 12% + Mancozeb 63% (SAAF 75 WP) with mycelial growth inhibition of (99.22%) and (85.92%) at 0.25 and 0.2% concentration, respectively.

In combinations Carbendazim + *Pseudomonas fluorescens* was found most effective with complete inhibition at 100 ppm. Followed by Captan + *Trichoderma harzianum* and Hexaconazole + *Bacillus cereus*. In other way control was observed least effective with (0.00%) inhibition. Vivekanand *et al.* (2018) [24] also observed that combi, *Pseudomonas fluorescens* + Carbendazim were tested against chilli anthracnose caused by *C. capsici* at 50, 100, 200, 400 and 600 ppm, they have found mycelia inhibition i.e.(60.86, 66.20, 69.59, 71.17 and 76.57%). Basamma and Kulkarni (2017) [4]; Khan and Shahzad (2007) [14] also found the compatibility between these treatments combination.

Table 1: Effect of fungicides and bioagents at different concentration on per cent mycelia growth inhibition of *C. lindemuthianum*

Treatments	Concentration							
	100 ppm		250ppm		500ppm		750ppm	
	G	I	G	I	G	I	G	I
Control	38.10	0.00	38.10	0.00	38.10	0.00	38.10	0.00
Hexaconazole	9.52	75.02	8.63	77.29	6.66	82.55	5.70	84.98
Captan	18.09	52.48	14.33	62.27	12.18	68.05	11.50	69.75
Carbendazim	0.00	100	0.00	100	0.00	100	0.00	100
Hexaconazole + <i>Bacillus cereus</i>	17.77	53.30	13.66	63.98	7.26	80.89	6.92	81.77
Captan + <i>Trichoderma harzianum</i>	11.01	70.58	8.70	77.22	7.83	79.39	6.48	82.92
Carbendazim + <i>Pseudomonas fluorescens</i>	0.00	100	0.00	100	0.00	100	0.00	100
CD (p=0.05)	1.52	-	2.38	-	1.63	-	1.47	-

G=Average mycelia growth in (mm); I= Average mycelia growth inhibition in (%).

Antagonistic effect of three bio-agents with control were observed for mycelia growth inhibition of *C. lindemuthianum* (Table-2). *Trichoderma harzianum* (73.87%) was found highly effective against test fungus, followed by *Pseudomonas fluorescens* (52.93%) and minimum in *Bacillus cereus* (31.33%) mycelia growth inhibition. In other way control was observed least effective with (0.00%) inhibition. Padder *et al.* (2010) [19] and Rajesha *et al.* (2010) [21] also reported the similar result against *C. lindemuthianum*. Fitsum *et al.* (2014) [10] have found the highest percentage of inhibition of the mycelia germination against *C. lindemuthianum*, *T. viride* was obtained (80.39%) followed by *T. harzianum* and *P. fluorescens*. Begum *et al.* (2015) [6] among bioagents *T. harzianum* -2 isolate found most effective (77.78%) inhibition of mycelial growth and minimum in *P. fluorescens* was least effective against *Colletotrichum capsici*. (Mahadev, 2018) [16] also found that *Trichoderma harzianum* was effective against bean anthracnose. Vivekanand *et al.* (2018) [24] also reported that among bioagents *T. harzianum* was found effective and minimum in *P. fluorescens* against chilli anthracnose, *C. capsici*.

Table 2: Effect of biocontrol agents against *C. lindemuthianum*

Treatments	Mycelia growth in (mm)	Mycelia growth inhibition in (%)
Control	38.10	0.00
<i>Bacillus cereus</i>	26.09	31.33
<i>Trichoderma harzianum</i>	9.92	73.87
<i>Pseudomonas fluorescens</i>	17.89	52.93
CD (p=0.05)	1.77	-

Conclusion

Three fungicides alone and three in combinations were used at different concentrations. It has found that alone Carbendazim and in combination (Carbendazim + *P. fluorescens*) complete inhibition of *C. lindemuthianum* at 100 ppm, while Captan and (Hexaconazole + *B. cereus*) had observed minimum growth inhibition. Three bio control agents among these *T. harzianum* found maximum mycelium growth inhibition while minimum in *B. cereus*.

References

- Aggarwal SK, Mali BL, Rajput LS, Choudhary M. Epidemiology of anthracnose of black gram caused by *Colletotrichum lindemuthianum*. International Journal of Agriculture Sciences. 2017; 9(2):3656-3657.
- Agrios GN. Plant Pathology. 5th edition. Dept of Plant Pathology, University of Florida. Academic Press. An imprint of Elsevier, 2005, 487.
- Anonymous. National Agricultural Statistics Service, Crops and Plants USDA-NASS: Washington DC, USA, 2013.
- Basamma H, Kulkarni S. Studies on compatibility of *Bacillus subtilis* (Ehrenberg) Cohn. with chemical fungicides. Int. J Curr. Microbiol. App. Sci. 2017; 6(3):578-586.
- Baudoin ABAM. Laboratory exercises in Plant Pathology: An instructional kit, The American Phytopathological Society, 2011, 76.
- Begum S, Yumlembam RA, Marak TR, Nath PS. Integrated management of anthracnose of chilli caused by *Colletotrichum capsici* in West Bengal condition. The Bioscan. 2015; 10(1):1901-1904.
- Deshmukh AJ, Mehta BP, Patil VA. *In vitro* evaluation of some known bioagents to control *Colletotrichum gloeosporioides* Penz. and Sacc., causing anthracnose of Indian bean. Internat. J Pharma. Bio. Sci. 2010; 1(2):361-367.
- Faheem A, Razdan VK, Mohiddin FA, Bhat KA, Sheikh PA. Effect of volatile metabolites of *Trichoderma* species against seven fungal plant pathogen *in vitro*. Journal of Phytopathology. 2010; 2:34-37.
- Fernandez M, Casares A, Rodriguez R, Fueyo M. Bean germplasm evaluation for anthracnose resistance and characterization of agronomic traits: A new Physiological strain of *Colletotrichum lindemuthianum* infecting *Phaseolus vulgaris* L. in Spain. Euphytica. 2000; 114:143-149.
- Fitsum S, Amin M, Selvaraj T Alemayehu A. *In vitro* evaluation of some fungicides and bioagents against common bean anthracnose (*Colletotrichum lindemuthianum* Sacc. and Magnus) Briosi and Cavara. African Journal of Microbiology Research. 2014; 8(20):2000-2005.
- Gawade DB, Suryawanshi AP. *In vitro* evaluation of fungicides, botanicals and bioagents against *Colletotrichum truncatum* causing soybean anthracnose. Pl. Dis. Res. 2009; 24(2): 120-123.
- Habtu A, Ivan S, Zadoks JC. Survey of cropping practices and foliar disease of common beans in Ethiopia. Crop Protection. 1996; 15:179-186.
- Junaid JM, Shah TA, Bhat AH, Bhat NA, Dar NA, Ambardar VK. Morphology and status of occurrence of anthracnose of bean (*Phaseolus vulgaris* L.) caused by *Colletotrichum lindemuthianum* (Sacc and Magn.) Scrib in Kashmir Valley. The Bioscan. 2014; 9(1):235-241.
- Khan MO, Shahzad S. Screening of *Trichoderma* species for tolerance to fungicides. Pak. J Bot. 2007; 39(3):945-951.

15. Madhusudhan BS. Studies on soyabean anthracnose caused by *Colletotrichum truncatum* (Schw.) Andrus and Moore. M.Sc (Agri.) Thesis, UAS, Bangalore, Karnataka, (India), 2002.
16. Mahadev KK. Management on anthracnose of mungbean caused by *Colletotrichum lindemuthianum* (Sacc. and Magn.) Briosi and Cav. Thesis. M.Sc. College of Agriculture, Latur M.S. India, 2018.
17. Maibam N, Chandra S, Baiswar P, Majumder D, Saikia K. Host plant resistance and yield loss due to anthracnose caused by *Colletotrichum lindemuthianum* in french bean (*Phaseolus vulgaris*). Indian Journal of Hill Farming. 2015; 28(1):14-18.
18. Nene YL, Thapliyal PN. Fungicides in Plant Disease Control. 3rd Edn., Oxford & IBH Publishing Company Pvt. Ltd, New Delhi, last reprinted-2015, 1993, 531.
19. Padder BA, Sharma PN, Kapil R, Pathania A, Sharma OP. Evaluation of bioagents and biopesticides against *Colletotrichum lindemuthianum* and its integrated management in common bean. Not. Sci. Biol. 2010; 2(3):72-76.
20. Patil CU, Zape AS, Wathore SD. Efficacy of fungicides and bioagents against *Colletotrichum gloeosporioides* causing blight in *Piper longum*. International Journal of Plant Protection. 2009; 2 (1):63-66.
21. Rajesha G, Mantur SG, RaviShankar M, Boranayaka MB, Shadakshari TV. *In vitro* evaluation of fungicides and biocontrol agents against *Colletotrichum lindemuthianum* causing anthracnose of dolichos bean. International Journal of Plant Protection. 2010; 3 (1):114-116.
22. Tasiwal V, Benagi VI, Hegde YR, Kamanna BC, Naik KR. *In vitro* evaluation of botanicals, bioagents and fungicides against anthracnose of papaya caused by *Colletotrichum gloeosporioides* Penz. & Sacc. Karnataka J Agric. Sci. 2008; 22(4):803-806.
23. Vincent JM. Distortion of fungal hyphae in presence of certain inhibitors. Nature. 1947; 159. 239-241.
24. Vivekanand, Ravi S, Mishra RC, Nautiyal BP. Evaluation of fungicides, botanicals and biocontrol agents against chilli anthracnose caused by *Colletotrichum capsici*. Pl. Dis. Res. 2018; 33(1): 64-68.
25. Yesuf M. Seed borne nature of *Colletotrichum lindemuthianum* and its epidemic on common beans in the major bean growing areas of Ethiopia. A Ph.D. Thesis in Tropical Agriculture. Graduate School, Kasetsart University, 2005.