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#### Nayee Pranav J

Department of Agricultural Chemistry and Soil Sciences, Anand Agricultural University, Anand, Gujarat, India

#### Mistry Girish J

Assistant Research Scientist, Micro Nutrient Research Project, Anand Agricultural University, Anand, Gujarat, India

#### Kharadi Rahul S

Department of Agricultural Chemistry and Soil Sciences, Anand Agricultural University, Anand, Gujarat, India

#### Patel Dipak H

Department of Agricultural Chemistry and Soil Sciences, Anand Agricultural University, Anand, Gujarat, India

**Correspondence** Nayee Pranav J Department of Agricultural Chemistry and Soil Sciences, Anand Agricultural University, Anand, Gujarat, India

# Effect of EDTA on phytoextraction of Pb by sunflower (*Helianthus Annuus* L.)

# Nayee Pranav J, Mistry Girish J, Kharadi Rahul S and Patel Dipak H

#### Abstract

The study entitled, "Effect of EDTA on phyto extraction of Pb by sunflower (*Helianthus annuus* L.)" was undertaken by conducting a micro-plot experiment in the net house of Micronutrient Research Project (ICAR), Anand Agricultural University, Anand (Gujarat) during *kharif* season of the year 2015. The soil used for the experiment was alkaline in reaction with low in available  $P_{205}$  and medium in K<sub>2</sub>0. The Pb status was 0.00025 mg kg<sup>-1</sup> soil, which was spiked with 300 mg Pb kg<sup>-1</sup> soil before 30 days of sowing for the purpose of artificial contamination for the study. Three levels of EDTA (3, 6 and 9 mmol kg<sup>-1</sup> soil) and three periods of EDTA application (P<sub>1</sub>: 6<sup>th</sup>, 7<sup>th</sup> and 8<sup>th</sup> weeks after sowing, P<sub>2</sub>: 9<sup>th</sup>, 10<sup>th</sup> and 11<sup>th</sup> weeks after sowing and P<sub>3</sub>: 12<sup>th</sup>, 13<sup>th</sup> and 14<sup>th</sup> weeks after sowing) and one absolute control were kept for the study. The lead content of seed, shoot and root was significantly more under E<sub>9</sub>P<sub>1</sub>, combination than the rest of the combination except E<sub>6</sub>P<sub>2</sub> for seed and shoot and E<sub>9</sub>P<sub>2</sub> for root. The uptake of Pb by shoot and total uptake were significantly more under E<sub>9</sub>P<sub>1</sub> as compared to the rest except E<sub>6</sub>P<sub>2</sub>, while uptake by seed was maximum under E<sub>6</sub>P<sub>2</sub>. Remediation of Pb polluted soil is possible by using EDTA.

Keywords: lead, sunflower, root, shoot

#### Introduction

Soil contamination by heavy metals is a global environmental issue due to the rapid development of intensive agriculture and industry in many parts of the world. Elevated concentrations of heavy metal not only lead to reductions in the microbial activity and fertility of the soil, and in crop production (McGrath et al., 1995) <sup>[18]</sup>, but also threaten human health through the food chain. The remediation of soil and water contaminated with heavy metals has become a challenging task facing regulators and scientific communities. Recently, phytoextraction, the use of plants to extract heavy metals from contaminated soils, has been receiving an increasing amount of attention. Lead can be accumulated in plant organs and agricultural products (Mahmoud and El-Beltagy, 1998) <sup>[17]</sup> and human body consequently enter human food chain (Wagner, 1993) [27]. As a result of consumption of food, lead accumulates in human body and it may cause renal failure, brain and liver damage and it can attack the nervous system and cause failing of sickness (Lucky and Kenugopal, 1997)<sup>[15]</sup>. Lead is one of the most difficult pollutants to control (Salt et al., 1998)<sup>[23]</sup>. Phytoextraction is a soil cleanup technology that uses the ability of metal accumulator plant to extract metals from contaminated soil with their roots and to concentrate these metals in above ground plant parts (DE Salt et al., 1995; Chaney et al., 1997; Lai et. al., 2007 and Lai et. al., 2008) [3, 12, 13, <sup>22]</sup> and the metal-accumulating plant material can be safely harvested and removed from the site. Lead (Pb) is potential pollutant that readily accumulated in soil and sediments. Apart from natural weathering processes, lead contamination of the environment has resulted from industrial activities viz. mining and smelting processes, agricultural activities such as application of insecticide and municipal sewage and urban activities viz. use of lead in gasoline, paints and other materials (Sharma et al., 2005) [24]. The Pb-contaminated soils are difficult to remediate with natural phytoextraction that utilizes hyper accumulators. In order to enhanced the availability of Pb in soil solution and its trance location from root to shoot, application of some chelating agents such as ethylene diamine tetraacetic acid (EDTA), diethylene trinitrilo pentaacetic acid (DTPA), nitrilotriacetic acid (NTA), ethylenediamine disuccinate (EDDA) have been proposed by various workers (Huang et al., 1997; Meers et al., 2005; Kumar et al., 2011) <sup>[9, 19, 11]</sup>. EDTA is probably the most efficient chelate to increasing concentration of various metals especially lead in above ground plant tissues.

(Huang et al., 1997; Vassil et al., 1998; Meers et al., 2005; Kumar et al., 2011; Pal et al., 2012) [9, 19, 11, 21]. In phytoextraction research, two main strategies can be identified. The first is the use of metal hyperaccumulating species (Baker et al., 1994; Brown et al., 1994; Kumar et al., 1995) <sup>[11, 1, 2]</sup>. A second phytoextraction approach involves the use of high biomass producing species, such as sunflower (Helianthus annuus), and chemically enhancing their shoot levels to increase the removal efficiency. A number of soil amendments have been reported in literature which could render soil trace metals more phytoavailable, among which ethylene diamine tetraacetate (EDTA) has taken a predominant place (Cooper *et al.*, 1999; Epstein *et al.*, 1999; Shen et al., 2002) <sup>[4, 6, 25]</sup>. The plant species used for the enhanced phytoextraction experiment in this study is Helianthus annuus. In order to generate location specific information on "Effect of EDTA and on phytoextraction of Pb by Sunflower (Helianthus annuus L.)" the present study will be conducted during kharif, 2015 at Pot house of Micronutrient Research Project (ICAR), AAU, Anand.

# **Materials and Method**

Rapid industrialization and urbanization have created enormous problems of environmental pollution due to disposal of large quantity of effluents. Reports indicate that the untreated and contaminated industrial effluents pollute the soils with heavy metals which need due to attention for remediation. Therefore, to meet the objectives as mentioned earlier, the present study was undertaken to study the effect of chelateor on phytoextraction for heavy metals uptake. The present investigation on "Effect of EDTA on phytoextraction of Pb by Sunflower (Helianthus annuus)" was undertaken by conducting a micro-plot study in 2015. Three levels of EDTA (3, 6 and 9 mmol kg<sup>-1</sup> soil) and three periods of EDTA application (P1:  $6^{th}$ ,  $7^{th}$  and  $8^{th}$  weeks after sowing, P2:  $9^{th}$ ,  $10^{th}$ and 11th weeks after sowing and P3: 12th, 13th and 14th weeks after sowing) and one absolute control were kept for the study. After harvest of the crop, soil samples were collected and air dried in laboratory. The air dried soil samples were prepared by using wooden mortar and pestle and passed through 2 mm sieve. The soil samples were preserved in polythene bags for their chemical analysis later. The samples were washed with 0.2% detergent, 0.03 N HCl, single and double deionized water in a sequence and air dried. Then samples were dried in paper bags at 70° C till constant weight in a hot air oven and preserved for further analysis. The dried plant samples were cut and ground in a stainless steel blade mixer and were preserved in polythene bags for further analysis. The concentration of micronutrient and heavy metal in plant were expressed in terms of mg kg<sup>-1</sup>. Nutrient uptake was calculated by using yield and nutrient content data. The following formula was used to compute the nutrient uptake.

Nutrient uptake for Micronutrient (g plot<sup>-1</sup>) =  $\frac{\text{Nutrient content } (\text{mg kg}^{-1}) \text{ x Yield } (\text{g plot}^{-1})}{1000}$ 

# **Result and Discussion**

# Pb content in plant and its uptake

The perusal of data given in Table 4.3 reveal that the Pb content and in plant was significantly influenced by both levels of EDTA and period of its application, even comparison of the rest of the treatments with control showed significant variations in Pb content in different parts of the plant, which is graphically presented in Fig.4.3 and Fig.4.4.

Among the different levels of EDTA,  $E_9$  achieved significantly the highest Pb content in seed (150.85 mg kg<sup>-1</sup>) and root (539.70 mg kg<sup>-1</sup>), whereas in shoot Pb content was noted significantly higher under EDTA level  $E_6$  (626.82 mg kg<sup>-1</sup>) than  $E_3$ , but it was at par with  $E_9$  (605.32 mg kg<sup>-1</sup>).

The effect of different periods of application of EDTA was found significant for Pb content in seed, shoot and root. Among the different periods, P<sub>2</sub> recorded significantly the highest Pb content in seed (175.69 mg kg<sup>-1</sup>) and root (479.36 mg kg<sup>-1</sup>). The Pb content in shoot was found significantly higher under period P<sub>1</sub> (667.38 mg kg<sup>-1</sup>) than P<sub>3</sub> (241.60 mg kg<sup>-1</sup>), but it was on par with P<sub>2</sub> (646.26 mg kg<sup>-1</sup>). Comparison of control with rest of the treatments also showed significant differences. The significantly higher Pb content was observed with the rest of the treatments than control.

The interaction effect of E×P was significant for Pb content in seed, shoot and root (Table 4.3a, 4.3b and 4.3c). The combination of E×P indicated that the combination of E<sub>9</sub>P<sub>1</sub> recorded significantly higher values for Pb content in seed (205.26 mg kg<sup>-1</sup>), shoot (964.53 mg kg<sup>-1</sup>) and root (619.77 mg kg<sup>-1</sup>), which was at par with Pb content in seed (192.52 mg kg<sup>-1</sup>) and shoot (956.41 mg kg<sup>-1</sup>) under combination of E<sub>6</sub>P<sub>2</sub> and Pb content of root (598.87 mg kg<sup>-1</sup>) under combination of E<sub>9</sub>P<sub>2</sub>.

# Pb uptake

The perusal of data given in Table 4.3 reveal that the Pb uptake by seed, shoot, root and total uptake was significantly influenced by both levels of EDTA and periods of its application, even comparison of rest of the treatments with control showed significant variations in Pb uptake by various parts of the plant.

Among the different levels of the EDTA,  $E_6$  noted significantly higher Pb uptake in seed (21.97 mg plot<sup>-1</sup>) than  $E_3$  (14.54 mg plot<sup>-1</sup>), but it was at par with  $E_9$  (21.74 mg plot<sup>-1</sup>). The Pb uptake by shoot was significantly more under  $E_9$  (298.20 mg plot<sup>-1</sup>) than  $E_3$  (148.56 mg plot<sup>-1</sup>), but it was on par with  $E_6$  (277.32 mg plot<sup>-1</sup>). Among the different levels of the EDTA,  $E_9$  achieved significantly the highest Pb uptake by root (16.15 mg plot<sup>-1</sup>) and total uptake (336.09 mg plot<sup>-1</sup>). The application periods of EDTA indicated that  $P_2$  reported significantly the highest Pb uptake by seed (27.82 mg plot<sup>-1</sup>), shoot (316.69 mg plot<sup>-1</sup>), root (14.99 mg plot<sup>-1</sup>) and total uptake (359.50 mg plot<sup>-1</sup>).

The comparison of control with rest of the treatments showed higher Pb uptake by seed, shoot and root as well as total uptake as compared to control.

The interaction effect of  $E \times P$  was significant for Pb uptake by seed, shoot and total uptake by sunflower plant (Table 4.3d, 4.3e and 4.f). The  $E_6P_2$  combination noted significantly higher Pb uptake by seed (30.44 mg plot<sup>-1</sup>) than the rest of the combinations, but it was on par with  $E_9P_2$  (29.21 mg plot<sup>-1</sup>) and  $E_9P_1$  (27.24 mg plot<sup>-1</sup>). The Pb uptake by shoot (455.91 mg plot<sup>-1</sup>) was significantly more than rest of the treatment combinations except  $E_6P_2$  (440.27 mg plot<sup>-1</sup>) combination. The total uptake of Pb also followed the similar trend, wherein  $E_9P_1$  (499.17 mg plot<sup>-1</sup>) registered significantly higher Pb uptake than rest of the combinations barring  $E_6P_2$ (440.27 mg plot<sup>-1</sup>) combination.

The Pb content in different plant parts was increased with the increased levels of EDTA. The rise in Pb content in seed and root was 79.58 per cent and 118.91 per cent, respectively under  $E_9$  over  $E_3$ . In case of shoot it was 94.00 per cent under  $E_6$  over  $E_3$ . The Pb uptake was also increased as the levels of EDTA increased. The rise in the Pb content and uptake in

plant parts due to effect of EDTA on Pb, which increased the bioavailability of Pb in soil and thus lead accumulated in the plant. This results are supported by Omar *et al.* (2015) <sup>[20]</sup>, Hovseyan and Greipsson (2007) and Liphadzi and Khirkham (2009) <sup>[14]</sup> in sunflower, Kumar *et al.* (2011) <sup>[10]</sup> and Pal *et al.* (2012) <sup>[21]</sup> in mustard, Hovseyan and Greipsoon (2007) Ghasemi-Fasaei (2012) <sup>[7]</sup> and Ebrahimi (2013) <sup>[5]</sup> in maize.

The influence of periods of EDTA application was significant for Pb content and uptake by different plant parts of sunflower. The data indicated that maximum content and uptake of Pb by all the plant components were under  $P_2$ (application of EDTA at 9, 10 and 11 weeks after sowing) period of application. The phytoextraction of lead was improved under mid period of application because application of EDTA in early growth period of plant reduced plant growth and dry weight of plant because of Pb toxicity. The results reveled that application of EDTA at P2 (9th, 10th and 11th week after sowing) is the right period for bioaccumulation of lead and its removal by plant. The similar finding was also noted by Sinegani and Khalilikhah (2008) in sunflower, who also reported that the application of EDTA before seeding was inferior to its application after sowing. Between the periods (10 and 30 days after sowing), application at 30<sup>th</sup> day was better in extraction of Pb than 10 days after sowing.

The increase in Pb content and uptake in plant in rest of the treatments over control was due to the application of EDTA. The EDTA increased the Pb concentration in soil solution which was absorbed by the plant root and translocated in shoot. Similar results were also reported by Madrid and Khirkham (2002) <sup>[16]</sup>.

**Table 4.3:** Effect of EDTA levels and its application periods on Pb

 content and uptake in plant

Treatmonta	Pb content (mg kg <sup>-1</sup> )			Pb uptake (mg plot <sup>-1</sup> )			
Treatments	Seed	Shoot	Root	Seed	Shoot	Root	Total
EDTA Levels (mmol kg <sup>-1</sup> soil)							
E <sub>3</sub>	84.00	323.10	246.61	14.54	148.56	9.25	172.35
E <sub>6</sub>	138.58	626.82	414.46	21.97	277.32	13.12	312.41
E9	150.85	605.32	539.70	21.74	298.20	16.15	336.09
S. Em. ±	2.88	11.10	11.86	0.79	7.76	0.65	7.38
C.D. at 5%	8.49	32.74	34.99	2.32	22.89	1.91	21.76
Application periods							
P1	145.60	667.38	395.76	20.81	290.04	10.88	321.73
P2	175.69	646.26	479.36	27.82	316.69	14.99	359.50
P3	52.14	241.60	325.65	9.63	117.34	12.66	139.63
S. Em. ±	2.88	11.10	11.862	0.79	7.76	0.65	7.38
C.D. at 5%	8.49	32.74	34.99	2.32	22.89	1.91	21.76
Control Vs. Rest							
Control	10.71	172.51	127.19	2.13	109.12	7.23	118.47
Rest Treatment	124.48	518.41	400.25	19.42	241.36	12.84	273.62
S. Em. ±	3.70	14.26	15.24	1.01	9.97	0.83	9.48
C.D. at 5%	10.91	42.07	44.97	2.981	29.42	2.45	27.96
Interaction E x P	Sig.	Sig.	Sig.	Sig.	Sig.	NS	Sig.
C.V. (%)	7.6	6.9	9.5	13.3	10.2	15.8	8.6

 
 Table 4.3a: Interaction effect of EDTA levels and its application periods on Pb content (mg kg<sup>-1</sup>) in seed

EDTA levels Application Periods	E <sub>3</sub>	E <sub>6</sub>	E9
P1	63.02	168.52	205.26
P2	148.30	192.52	186.24
P3	40.67	54.70	61.06
S. Em. ±	4.99		
C.D. at 5%		14.71	

 
 Table 4.3b: Interaction effect of EDTA levels and its application periods on Pb content (mg kg<sup>-1</sup>) in shoot

EDTA levels \ Application Periods	E3	E6	E9
P1		689.51	
P2	455.79	956.41	526.58
P3	165.44	234.52	324.85
S. Em. ±	14.26		
C.D. at 5%	42.07		

 
 Table 4.3c:
 Interaction effect of EDTA levels and its application periods on Pb content (mg kg<sup>-1</sup>) in root

EDTA levels \ Application Periods	E3	E6	E9
P1		362.77	
P2	263.65	575.55	598.87
P3	271.44	305.05	400.46
S. Em. ±	11.86		
C.D. at 5%	34.99		

 
 Table 4.3d: Interaction effect of EDTA levels and its application periods on Pb uptake (mg plot<sup>-1</sup>) in seed

EDTA levels \ Application Periods	E <sub>3</sub>	E <sub>6</sub>	E9
P1	11.41	23.78	27.24
P2	23.81	30.44	29.21
P3	8.40	11.70	8.79
S. Em. ±	1.36		
C.D. at 5%	4.02		

 Table 4.3e:
 Interaction effect of EDTA levels and its application periods on Pb uptake (mg plot<sup>-1</sup>) in shoot

EDTA levels \ Application Periods	E3	E <sub>6</sub>	E9
P1	161.81	252.41	455.91
P2	208.09	440.27	301.72
P3	75.77	139.28	136.96
S. Em. ±	13.44		
C.D. at 5%		39.65	

 
 Table 4.3f: Interaction effect of EDTA levels and its application periods on Total Pb uptake (mg plot<sup>-1</sup>) in plant

EDTA levels \ Application Periods	E3	E <sub>6</sub>	E9
P1	180.53	285.48	499.17
P2	242.28	486.95	349.27
P3	94.24	164.82	159.83
S. Em. ±		12.77	
C.D. at 5%	37.69		

### References

- Baker AJM, Reeves RD, Hajar ASM. Heavy metal accumulation and tolerance in British populations of the metallophyte Thlaspi caerulescens J. & C. Presl. (Brassicaceae). New Phytol. 1994; 127:61-68.
- 2. Brown SL, Chaney RL, Angle JS, Baker AJM. Phytoremediation potential of Thlaspi caerulescens and bladder campion for Zinc- and Cadmium-contaminated soil. J Environ. Qual. 1994; 23:1151-1157.
- Chaney RL, Malik M, Li YM, Brown SI, Brewer EP, Angle JS, *et al.* Phytoremediation of soil metals. Environ Biotechnology. 1997; 8:279-284.
- Cooper EM, Sims JT, Cunningham SD, Huang JW, Berti WR. Chelate-assisted phytoextraction of lead from contaminated soils. J Environ. Qual. 1999; 28:1709-1719.
- Ebrahimi M. Effect of EDTA application on heavy metals uptake and germination of *Echinochloa crusgalii* (L.) Beave in contaminated soil. International Journal of Agriculture and Crop Science. 2013; 6(4):197-202.
- 6. Epstein AL, Gussman CD, Blaylock MJ, Yermiyahu U, Huang JW, Kapulnik Y, et al. EDTA and Pb-EDTA

accumulation in Brassica juncea grown in Pb- amended soil. Plant Soil. 1999; 208:87-94.

- Ghasemi-Fasaei R. Effects of EDTA and Phosphorus levels on Lead Phytoremediation by Maize. International J of Agri. And Crop Sci. 2012; 4(23):1786-1790.
- Hovsepyan A, Greipsson S. EDTA-Enhanced Phytoremediation of Lead-Contaminated Soil by Corn. J of Plant Nutrition. 2005; 28:2037-2048.
- Huang JW, Chen J, Berti WR, Cunningham SD. Phytoremediation of lead contaminated soils: Role of synthetic chelates in lead phytoextraction. Environ. Sci. Technol. 1997; 31:800-805.
- 10. Kumar J, Srivastava A, Singh VP. EDTA Enhanced Phytoextraction of Pb By Indian Mustard (*Brassica juncea* L.). Plant Science Feed. 2011; 1(9):160.
- 11. Kumar PBAN, Dushenkov V, Motto H, Raskin I. Phytoextraction: the use of plants to remove heavy metals from soils. Environ. Sci. Technol. 1995; 29:1232-1238.
- 12. Lai HY, Chen SW, Chen ZS. Pot experiment to study the uptake of Cd and Pb by three Indian mustards (*Brassica juncea*) grown in artificially contaminated soils. International J Phytoremediation. 2008; 10:91-105.
- 13. Lai HY, Chen ZS. The effect of multi-dose EDTA application on the phytoextraction of Cd, Zn, and Pb by rainbow pink (*Dianthus chinensis*) in contaminated soil. Desalination. 2007; 210:236-247.
- Liphadzi MS, Kirkham MB. Partitioning and Accumulation of Heavy metals in Sunflower Grown at Biosolids Farm in EDTA-facilitated Phytoremediation. Bioremediation, Biodiversity and Bioavailability. 2009; 3(1):36-42.
- 15. Lucky TD, Kenugopal B. Metal toxicity in mammals. Plenum Press, New York and London, 1997.
- Madrid F, Kirkham MB. Heavy metal uptake by barley and sunflower grown in abandoned animal lagoon soil. 17<sup>th</sup> World Congress of Soil Science, Symposium no. 2002; 42:401.
- 17. Mahmoud WH, El-Beltagy A. Isolation, Identification and potential use of lead reduction from heavy metal polluted soil Menufiya. Journal Agriculture Research. 1998; 23:1461-1473.
- 18. McGrath SP, Chaudri AM, Giller KE. Long-term effects of metals in sewage sludge on soils, microorganisms and plants. Journal of Indian Microbiology. 1995; 14:94-104.
- 19. Meers E, Rutters A, Hopgood MJ, Samson D, Tack FMG. Comparison of EDTA and EDDS as potential soil amendments for enhanced phytoextraction of heavy metals. Chemosphere. 2005b; 58:1011-1022.
- Omar HRS, Mezori HAM. EDTA-assisted Phytoextraction of Lead from Artificially Polluted soil by Sunflower Plants. International conference on Chemical, Civil and Environmental Engineering (CCEE-2015), 2015, 5-6.
- Pal MK, Panwar BS, Singh D. Phytoextraction of Lead by Indian Mustard. Indian Journal of Agricultural Research. 2012; 46(1):23-29.
- 22. Salt DE, Blaylock MJ, Kumar NPVA, Dushenkov V, Ensley BD, Chet I, *et al.* Phytoremediation: A novel strategy for the removal of toxic metals from the environment using plant. Bio technol. 1995; 13:468-474.
- Salt DE, Blaylock M, Raskin I. Phytoremediation: Annu. Rev. Plant Physiology Plant Molecular Bio logy. 1998; 49:643-668.
- 24. Sharma P, Dubey RS. Lead toxicity in plants. Braz. J Plant Physiol. 2005; 17:35-52.

- 25. Shen ZG, Li XD, Wang CC, Chen HM, Chua H. Lead phytoextraction from contaminated soil with high-biomass plant species. J Environ. Qual. 2002; 31:1893-1900.
- 26. Vassil AD, Kapulnik Y, Raskin I, Salt DE. The role of EDTA in lead transport and accumulation in Indian mustard. Plant Physiology. 1998; 117:447-453.
- 27. Wagner GJ. Accumulation of heavy metals in crop plants and its consequences to human health. Advance in Agronomy. 1993; 51:173-177.