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Comparative therapeutic efficacy of various teat dip solutions in caprine mastitis

Ankur Tomar, PC Shukla, Brejesh Singh and Amir Amin Sheikh

Abstract

Mastitis is an inflammatory condition of mammary glands characterized by changes in physical characteristics of udder or milk. Its diagnosis is based on clinical signs and increased SCC. The higher SCC indicated the higher possibility of pathogen contamination. In the present study a total of 284 lactating does belonging to the government farms and private goat keepers in and around Jabalpur were screened using MCMT as a diagnostic test for clinical mastitis. The mean pH value recorded as 6.82 ± 0.07 in T₅ group (Inj. Enrofloxacin + H.F.D. + Aloe vera) was nearer to the normal mean value (6.54 ± 0.00) of Somatic Cell Count obtained (18.51 ± 3.77) indicated significant reduction in somatic cell count (SCC). Further the lactose content ($4.36^{B}\pm0.06$) and fat content ($4.23^{B}\pm0.06$) were reported to be increase significantly in T₅ group, whereas mean value of protein ($3.41^{B}\pm0.01$) and SNF ($11.25^{B}\pm0.03$) revealed gradual increase in all the groups under therapeutic trial, but a significant increase was observed in T₅ group. So, on the basis of decreased pH and SCC values and a significant increase in milk parameters viz, lactose %, fat%, protein% and SNF% reported in T₅. Further, in treatment the maximum cure rate was also reported in T₅ group showed overall superiority in terms of the efficacy of the teat dip used under T₅ group as compared to the other teat dips used in T₁ (Inj-Enrofloxacin), T₂ (Inj-Enrofloxacin + Aloe vera), T₃ (Inj-Enrofloxacin + HFT) and T₄ (Inj-Enrofloxacin + 0.4% Chlorhexidine Gluconate).

Keywords: Mastitis, MCMT, somatic cell count, teat dip

Introduction

Mastitis is well known pathological condition of mammary tissues. It refers to the inflammation of mammary glands and is often encountered in dairy goats after parturition. Clinical mastitis is a major cause of loss in milk production to goat husbandry, eventually incurring economic losses to the poor farmers. However, in small ruminants since the milk production is always presumed to be secondary. Hence, mastitis never received prompt attention and remained neglected. The present study was undertaken to assess the efficacy of different teat dips in treating clinical mastitis. For this purpose lactating goats belonging to different private goat keepers and livestock farm Adhartal and Amanala in and around Jabalpur were screened for mastitis. The results of the curative therapies were judged by retesting the milk samples of treated animals at various days by SCC, milk pH, Lactose%, Fat%, Protein% and SNF% and also involved the curative rate under various groups post-treatment.

Material and Methods

Collection of milk samples

The udder of each goat was thoroughly washed with potassium permangnate solution (1:1000), wiped with clean cloth and allowed to dry before collection of the milk samples. The mid stream milk samples was collected from each half of the lactating goats.

Testing of milk samples

Modified California Mastitis Test (MCMT)

The MCMT was performed as per the method described by Shukla (1980) ^[1]. The reagent was prepared by adding 2ml stock solution B (Bromocresol purple reagent) to make volume 100ml by adding remaining volume of stock solution A (Sodium lauryl sulfate reagent).

MCMT Grading

MCMT Grading Equal quantity of milk and MCMT reagent was added in a mastitis paddle,

giving gentle swirling motion in a horizontal plane within a minimum agitation did mixing of the contents. In negative cases, the mixture remains liquefied. Grading of the test of positive samples was done according to the intensity of viscous and gel formation, reflecting the degree of inflammation and leukocyte count. It was scored as trace, 1+, 2+ and 3+ reaction if depending on the amount of gel formation.

Somatic Cell Count

The procedure for SCC was adopted as per method by Shukla (1980)^[1]. The Ieukocyte count in the mastitic milk was made to assess the degree of infection in the repective halves. The CMT positive milk (showing flakes or change in consistency) samples were collected in sterile small glass vials. The name or number of goats from which the sample was collected was labeled on vials. The milk sample collected in vials was transported on ice to the laboratory at College of Veterinary Science and Animal Husbandry, Jabalpur for further examination.

Preparation of milk smear

The smear of milk for SCC was prepared within one hour of it's collection to minimize disintegration of leukocyte. Each milk sample was uniformly mixed by gentle shaking of the vials and the milk (0.01 ml) was spread with sterilized bacteriological loop, over one cm rectangular area on a clean microslide. The milk smear from the test sample was stained by modified Newman's stain. A total of 30 fields were counted under oil immersion lens and average number of cells per field was worked out. The average number of cells was multiplied by the multiplication factor of the microscope i.e. 497512 to obtain the number of cells per ml of the milk.

Microscopic Factor Determination

The diameter of the field of 10 x eye piece was predetermined by using stage micrometer. The lowest division of micrometer scale was 0.01 mm. Accordingly, the diameter was measured and obtained as 0.016 cm. The area of the microscopic field was determined by the formula. The calculation was made as under:

Diameter	= 0.016 cm		
Radius	= 0.008 cm		
Area	= 3.14 x (0.008)2		
	= 0.000201		
Microscopic factor	= 100 x (1/Area)		
	= 497512 x (1/0.000201)		
	= 497512.43 or 497512		
The same calibrated	research microscope	Was	

The same calibrated research microscope was used throughout the course of study (Shukla, 1980)^[1].

Milk pH

Milk pH was estimated by digital pH meter standardized by non buffer solution thereafter, the pH reading of the normal and mastitic milk sample was recorded on day 0 (pretreatment) and on day 5 and 10 (post-treatment).

Analysis of milk samples

Analysis of milk sample was done by milk analyser to estimate Protein, Lactose, Fat and Solid not fat (SNF) and reading of the normal and mastitic milk sample was recorded on day 0 (pre-treatment) and on day 5 and 10 (posttreatment). Under the present study the score +3 was taken for the therapeutic regimen. The +3 score is resultant as a result of infiltration of leukocytes in milk leading to gel formation and viscosity depending upon the intensity of infection as reported by Shukla, (1980)^[1]. The infected does were divided into 5 groups, each comprising of 6 animals for this study while 6 healthy does were kept as control Table 1.

Table 1: Specific therapies for various groups of does under study

Group	No. of does	Drugs
Tc	6	Healthy Control
T_1	6	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD for 5 days.
T ₂	6	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera for 5 days.
T 3	6	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + HFT Dip 20% for 5 days.
T_4	6	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip 0.4%chlorhexidine gluconate for 5 days.
T 5	6	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera + HFT Dip

The results of the curative therapy were judged by retesting the milk samples after completion of the therapy on (day 5^{th} and day 10^{th}).

Modified California Mastitis Test (MCMT)

In present study, a total of 284 does were screened out of which 30 (10.56%) does were found to be positive as formation of gel or viscous mass on CMT paddle with MCMT score +3. Under the present study the clinical symptoms like pyrexia, reduction in milk yield, udder became tender, swollen, enlarged, painful inflamed, redness sudden onset anorexia are the visible signs recorded in does affected with clinical mastitis. However, there is reduction in milk yield, discoloration of milk, presence of clots, flakes with yellowish fluid or blood watery in consistency streaks of blood as also reported by Smith and Rogunisky (1977) ^[2], Devendra and Mcleroy (1982) ^[3], Thomas *et al.*, (1988) ^[4], Muhammad *et al.*, (1995) ^[5], and (Radostits,2000) ^[6]. On the post treatment the physical properties as: colour, consistency and odour of the milk was found to be apparently normal.

Result and Discussion Somatic Cell Count (SCC)

Shangguan (2008)^[7] defined SCC in milk as the combinations of the WBCs known as leukocytes which includes macrophages (66 to 68%) lymphocytes and neutrophils in milk and relatively small amount of epithelial cells accumulated to a greater amount producing an abnormal SCC in milk. Hence, SCC plays an important role in innate mammary gland immune defense system. The SCC increases as per the stage of lactation due to more number of handling for milking thereby more chances of infection as Scott et al., [8] (2002)advocated SCC as better predictor of bacteriological status than the CMT score in goats and sheep. During the present study results obtained showed a drastic decrease in Somatic Cell Count (18.51±3.77) was recorded in T₅ group followed by T₃ (20.17±3.70), T₂ (21.28±3.77), T₄ (24.04 ± 3.71) and T₁ (25.43 ± 3.46) groups Table 2.

 Table 2: Mean and standard errors of Somatic Cell Count (10⁵ cells/ml) in various treatment groups at different intervals

Treatment	I	Overall		
Treatment	0	5	10	Overall
Control	9.95 ^a ±1.28	9.95 ^a ±1.28	9.95 ^a ±1.28	9.95 ^b ±0.70
T1	$43.94^{a}\pm2.99$	17.41 ^b ±2.13	14.93 ^b ±2.22	25.43 ^a ±3.46
T2	$41.45^{a}\pm3.56$	$11.61^{b}\pm 2.10$	10.78 ^b ±2.37	21.28 ^a ±3.77
T3	40.63 ^a ±2.00	10.78 ^b ±2.37	$9.12^{b}\pm 2.00$	20.17 ^a ±3.70
T4	$43.94^{a}\pm3.94$	14.92 ^b ±1.82	13.27 ^b ±1.66	24.04 ^a ±3.71
T5	$39.80^{a} \pm 1.82$	$9.12^{b} \pm 1.53$	$6.63^{b} \pm 1.66$	18.51 ^{ab} ±3.77
NT / N.C.		• . •.	1	11.00

Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

These values were little bit higher than the normal values as also reported by Okada (1960)^[9], Deutz et al., (1990)^[10] and Khodke et al., (2009) [11] as 7.5, 8.80 and 6.84 ×1 lakh cells/ml respectively. On post treatment the decrease number of SCC was obtained as a result of treatment effect leads to decrease inflammation due to decrease in number of PMN cells. On the other hand, the increased values of SCC reported pre treatment in all the groups can be attributed to increase PMN/neutrophils (Kitchen, 1981) ^[12], as a result of various enzymes into the body fluid was from damaged tissues or inflammed cells and liberation of parenchyma cells of udder resulted in increased SCC of milk (Kitchen et al., 1970) [13]. As compared to control group 9.95 lakh cells /ml of milk; Similar observations have been reported by Vihan and Rubino (1996) ^[14] as 70 lakh cells/ml of milk in infected does as compared to uninfected goats with 3.2 lakh cells / ml of milk.

Milk pH

Milk pH could serve as the best indicator to assess the condition of udder health in dairy animals, and mentioned that highly significant difference in milk pH was due to severity in mastitis. The findings of the present study revealed that mean value of pH was increased in all the groups on day 1st with control group comparatively but on post treatment these values were decreased in all the groups significantly. The reduction in the pH was maximum in T5 group as 7.22±0.04, 6.69±0.05 and 6.56±0.04 on day 0, 5 and 10 post treatments respectively Table 3. The observations of Khodke et al., (2009)^[11], are nearby our findings as 6.40, as the normal ph values reported by them respectively. The probable reason for increased pH may be due to severity of mastitis that may attribute to lowered acidity as found in mastitic milk (Horvarth et al., 1980)^[15]. This lowered acidity is due to the reduction in lactose content, as the lactic acid formation is minimum in mastitc milk. The decreased pH value is because of reduction in the alkalinity due to inflammation thereby increased Na⁺ and Cl⁻ ions in the milk following damage to mammary epithelium and also as a result of treatment as also observed by Ali and Hasan, (1998) [16], as 6.73 and 6.77 respectively. The study on viability of pH in milk was conducted by Agnihotri and Rajkumar (2007) [17], and observations were b/w 6.53 to 6.34 and concluded the reason being effect of the breed, stage of lactation, health of animal and bacterial invasion in raw milk. Whereas, Imran et al., (2008) ^[18], reported the pH 6.93. Further, Juarez and Ramos (1986) ^[19] have reported milk pH in clinical mastitis in does between 6.50 to 6.80 in their studies.

 Table 3: Mean and standard errors of pH in various treatment groups at different intervals

I	0		
0 5 10		10	Overall
$6.54^{a}\pm0.01$	$6.54^{a}\pm0.01$	$6.54^{a}\pm0.01$	6.54°±0.00
$7.56^{a}\pm0.04$	6.99 ^b ±0.06	6.84 ^b ±0.11	7.13 ^a ±0.08
7.41ª±0.02	6.92 ^b ±0.03	6.67°±0.11	$7.00^{ab}\pm0.08$
$7.40^{a}\pm0.12$	6.91 ^b ±0.03	6.64°±0.06	6.98 ^{ab} ±0.09
7.57 ^a ±0.02	$6.98^{b}\pm0.06$	$6.78^{b}\pm0.11$	7.11 ^a ±0.07
$7.22^{a}\pm0.04$	$6.69^{b} \pm 0.05$	$6.56^{b} \pm 0.04$	$6.82^{b} \pm 0.07$
	0 6.54 ^a ±0.01 7.56 ^a ±0.04 7.41 ^a ±0.02 7.40 ^a ±0.12 7.57 ^a ±0.02 7.22 ^a ±0.04	Interval (days 0 5 6.54 ^a ±0.01 6.54 ^a ±0.01 7.56 ^a ±0.04 6.99 ^b ±0.06 7.41 ^a ±0.02 6.92 ^b ±0.03 7.40 ^a ±0.12 6.91 ^b ±0.03 7.57 ^a ±0.02 6.98 ^b ±0.06 7.22 ^a ±0.04 6.69 ^b ±0.05	Interval (days)0510 $6.54^{a}\pm0.01$ $6.54^{a}\pm0.01$ $6.54^{a}\pm0.01$ $7.56^{a}\pm0.04$ $6.99^{b}\pm0.06$ $6.84^{b}\pm0.11$ $7.41^{a}\pm0.02$ $6.92^{b}\pm0.03$ $6.67^{c}\pm0.11$ $7.40^{a}\pm0.12$ $6.91^{b}\pm0.03$ $6.64^{c}\pm0.06$ $7.57^{a}\pm0.02$ $6.98^{b}\pm0.06$ $6.78^{b}\pm0.11$ $7.22^{a}\pm0.04$ $6.69^{b}\pm0.05$ $6.56^{b}\pm0.04$

Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

Schalm *et al.*, (1971) ^[20], mentioned the explanation of elevated pH due to increased permeability of glands to blood components due to increased movements of bicarbonate ions into the milk.

Lactose

On the analysis of milk samples maximum increase in overall lactose % (4.36 \pm 0.06) was obtained in T₅ group followed by T₃ (4.24 \pm 0.08), T₂ (4.23 \pm 0.06), T₄ (4.11 \pm 0.06) and T₁ (4.09 \pm 0.08) groups Table 4.

 Table 4: Mean and standard errors of Lactose (%) in various treatment groups at different intervals

Treatment	I	Omenall			
1 reatment	0 5		10	Overall	
Control	4.65 ^a ±0.01	4.65 ^a ±0.01	4.65 ^a ±0.01	4.65 ^a ±0.00	
T1	3.80 ^b ±0.12	4.18 ^{ab} ±0.13	4.29 ^a ±0.13	4.09°±0.08	
T2	3.99 ^b ±0.08	4.30 ^a ±0.09	$4.40^{a}\pm0.11$	4.23 ^{bc} ±0.06	
T3	3.90 ^b ±0.11	4.35 ^a ±0.10	4.47 ^a ±0.12	$4.24^{bc} \pm 0.08$	
T4	3.88 ^b ±0.05	$4.19^{a}\pm0.11$	$4.26^{a}\pm0.11$	4.11°±0.06	
T5	$4.10^{b} \pm 0.08$	4.43 ^a ±0.06	$4.56^{a}\pm0.07$	4.36 ^b ±0.06	

Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

Lactose content was more indicative of SCC than fat, protein, SNF and total solids although lactose content decreased with higher SCC. The different level of high SCC representing severity of infection in goats did not affect the other milk components. The stage of lactation affects the composition and lactose content. In early lactation there is high production of milk (Shangguan 2010)^[7]. Similar observations have been reported by Arora (2013) ^[21] as 4.45 % lactose content as normal. The decreased lactose content recorded in mastitic milk under the study is well explained by decreased lactate production by udder tissues. So, the alkalinity of mastitic milk gets reduced keeping the slight acidity of milk, as a result of acidic group of casein, citrate, phospohrous and dissolved CO₂ in milk (Kitchen et al., 1970)^[13] and (Schalm et al., 1971)^[20]. There was a significant difference in fat and lactose percentage in milk of goat with SCM as compared to milk of uninfected animal, while the percentage of lactose in milk of uninfected is also compared to the milk of infected does. Lactose is synthesized in the gland cells of the udder from glucose and galactose during infection reduce secretary action of mammary cells due to destruction of epithelial cells by the

leukocytes. These changes were linked with many factor such as breed, feed, environmental condition and age (Hassan, 2013)^[22].

Fat, Protein and SNF

The results found for Fat, Protein and SNF are presented in Table 5, 6 & 7. Under the study maximum increase in fat % (4.23±0.06) was obtained in T₅ group followed by T₃ (4.10±0.06), T₂ (4.08±0.05), T₄ (4.00±0.06) and T₁ (3.98±0.05) groups along with maximum increase in protein % (3.41±0.01) was obtained in T₅ group followed by T₃ (3.31±0.03), T₂ (3.29±0.03), T₄ (3.31±0.03) and T₁ (3.20±0.01) groups and also maximum increase in SNF % (11.25±0.03) was obtained in T₅ group followed by T₃, T₂, T₄ and T₁ groups followed by T₃ (11.15±0.03), T₂ (11.12±0.02), T₄(11.01±0.03) and T₁(10.98±0.04) groups.

 Table 5: Mean and standard errors of Fat (%) in various treatment groups at different intervals

Treatment]	Overall		
Treatment	0 5 10		Overall	
Control	$4.52^{a}\pm0.02$	$4.52^{a}\pm0.02$	$4.52^{a}\pm0.02$	4.52 ^a ±0.00
T1	$3.76^{b}\pm0.05$	$4.00^{a}\pm0.06$	$4.18^{a}\pm0.09$	3.98°±0.05
T2	$3.88^{b}\pm0.06$	4.11 ^{ab} ±0.06	4.25 ^a ±0.09	$4.08^{bc} \pm 0.05$
T3	$3.82^{b}\pm0.06$	4.18 ^a ±0.09	$4.30^{a}\pm0.10$	$4.10^{bc} \pm 0.06$
T4	3.71 ^b ±0.05	4.08 ^a ±0.03	4.21ª±0.07	4.00°±0.06
T5	$3.96^{b} \pm 0.05$	$4.32^{a}\pm0.09$	4.42 ^a ±0.09	4.23 ^b ±0.06

Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

These values of fat % were nearer to the values obtained by Hassan (2013)^[22] and Getaneh *et al.*, (2016)^[23] as 4.20 and 4.8 respectively. Whereas, the observations of Arora (2013)^[21] were differed from these results as 3.80. Significant difference was lower in fat % of milk from infected animal with SCM had very high increase in the activity of enzyme called lipase, that cause milk fat breakdown and release free fatty acids that produce off flavors in milk and cause great loss to dairy industry. In goats the data indicated a decrease in total protein from uninfected to infected animals (Hassan, 2013)^[22]. Milk protein% is increased as lactation progressed a significant low fat content of milk during mid lactation then that from early or late lactation with altered level of SCC and lactose percentage was observed by Shangguan (2010)^[7].

 Table 6: Mean and standard errors of Protein (%) in various treatment groups at different intervals

Treatment	I	0		
Ireatment	0 5 10		Overall	
Control	$3.52^{a}\pm0.01$	$3.52^{a}\pm0.01$	$3.52^{a}\pm0.01$	$3.52^{a}\pm0.00$
T1	3.13°±0.01	$3.20^{b}\pm0.01$	$3.26^{a}\pm0.01$	$3.20^{d} \pm 0.01$
T2	3.17 ^b ±0.02	$3.32^{a}\pm0.04$	3.38 ^a ±0.05	3.29°±0.03
T3	3.19 ^b ±0.02	3.33 ^a ±0.04	3.41 ^a ±0.05	3.31°±0.03
T4	3.14°±0.01	$3.20^{b}\pm0.01$	$3.26^{a}\pm0.01$	$3.20^{d} \pm 0.01$
T5	3.32°±0.01	3.43 ^b ±0.01	$3.49^{a}\pm0.02$	3.41 ^b ±0.01
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Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

The results also showed with significant higher values in SNF % there was increase in protein percent obtained in uninfected milk. The study indicated decrease in SNF in infected ewe's

milk depend on the destruction that occur by invasion of pathogens to the mammary tissues cause decrease in synthetic activity of Mammary gland (Hassan, 2013) ^[22]. The nutritional component of goat milk include higher chloride content and fat %. However, they are affected by age, breed, stage of lactation does of animal processing and preparation of produced milk and managmental facilities. Hence, the goat milk with its unique composition could be a valuable alternate. The milk composition can be affected by a wide array of factors viz: breed, age, stage of lactation and diet of animal Marchel *et al.*, (2011) ^[24].

 Table 7: Mean and standard errors of SNF (%) in various treatment groups at different intervals

Treatment	Ι	Omercell		
Ireatment	0	5	10	Overall
Control	11.41 ^a ±0.09	11.41 ^a ±0.09	11.41 ^a ±0.09	11.41 ^a ±0.05
T1	$10.71^{b}\pm0.02$	11.09 ^a ±0.03	$11.14^{a}\pm0.02$	$10.98^{de} \pm 0.04$
T2	11.01 ^b ±0.04	$11.16^{a}\pm0.03$	11.19 ^a ±0.03	11.12 ^{cd} ±0.02
T3	10.97 ^b ±0.04	11.21ª±0.03	$11.28^{a}\pm0.03$	11.15 ^{bc} ±0.03
T4	10.82 ^b ±0.03	11.10 ^a ±0.03	11.13 ^a ±0.02	11.01 ^{de} ±0.03
T5	11.07°±0.02	11.31 ^b ±0.01	11.39 ^a ±0.02	11.25 ^b ±0.03

Note: Means with same superscript within a row for different days do not differ significantly from each other ($p \le 0.01$). Likewise means of groups within the overall column with same superscript do not differ from each other ($p \le 0.01$).

In view of the parameters studied it is concluded that the increased milk parameters i.e. lactose, fat protein and SNF % are directly correlated with each other. Simultaneously, there is reduction of inflammation and impact of treatment have support them for their enhanced values at post treatment.

Efficacy of Treatment

The efficacy of therapeutic agents in clinical mastitis are presented in Table 8.

Group T₁

The treatment followed in this group comprised of Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD for 5 days. The post treatment curative effect obtained on animals basis was 50%.

Group T₂

The animals belonging to this group were treated with Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera for 5 days as per the schedule. A total of 6 animals were treated for 5 days. The post treatment curative rate was found to be 66.66% effective.

Group T₃

In this group animals were treated with Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + HFT Dip 20% for 5 days The post treatment curative rate was found to be 66.66%.

Group T₄

The animal in this group were treated with Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip 0.4% chlorhexidine gluconate for 5 days combination was found to be 50% effective.

Group T5

The treatment followed in this group comprised of Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera + HFT Dip for 5 days the curative effect obtained was maximum in this group i.e. 83.33%.

Labic 0. Efficacy of unclabeling agents in chinear mastru	Table 8:	Efficacy	of thera	peutic a	agents in	clinical	mastitis
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Groups	Name of therapy	Animals treated	Animals cured	Animals cure percent
T1	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD for 5 days.	6	3	50
T ₂	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera for 5 days.	6	4	66.66
T3	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + HFT Dip 20% for 5 days.	6	4	66.66
T 4	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip 0.4%chlorhexidine gluconate for 5 days.	6	3	50
T ₅	Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera + HFT Dip	6	5	83.33

Conclusion

Significant decrease in pH and Somatic Cell Count, with maximum increase in lactose %, fat %, protein %, SNF % and maximum cure rate was noticed in T5 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera + HFT Dip) group. Hence, in view of the findings of present investigation, T5 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera + HFT Dip) group proved the efficacy and superiority of the teat dip used in this group, over all the other teat dips used in T1 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera for 5 days.), T2 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip Aloe Vera for 5 days), T3 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + HFT Dip 20% for 5 days) and T4 (Inj-Enrofloxacin @ 5mg/kg b.wt. I/M BD + Teat Dip 0.4% chlorhexidine gluconate for 5 days) groups.

References

- Shukla PC. A Study to evaluate the relative sensitivity of some of the indirect diagnostic tests available for detection of subclinical cases of mastitis. M.V.Sc and A.H. thesis (Veterinary Medicine). Jawahar Lal Nehru Krishi Vishvidhyalaya, Jabalpur, 1980.
- Smith MC, Roguinsky. Mastitis and other diseases of goats' udder. J Am Vet Med Assoc, 1977; 17(12):1241-1248.
- 3. Devendra C, McLeroy GB. Goat and sheep production in the tropics. Edn. 1st publ. ELBS. London. 1982; 4-5.
- 4. Thomas AD, Faulkner F, Norton, Trueman KF. Clinical and pathological observation on goats experimentally infected with *Psuedomonas pseudomallei*. Aust Vet J. 1998; 65(2):43-46.
- 5. Muhammad G, Athar M, Shakoor A, Khan MZ, Rehman F, Ahmad MT. Surf field mastitis test (SFMT): An inexpensive new tool for evaluation of wholesomeness of fresh milk. PJBS. 1995; 5:91-93.
- Radostits OM, Gay CC, Blood DC, Hinchcliff KW. Mastitis in veterinary medicine: Text book of the disease of cattle, sheep, pigs, goats and horses. Edn. 9th publ. Book power with Saunders, London. 2000; 611-613.
- Shangguan R. Effect of subclinical mastitis and stage of lactation on somatic cell count, milk composition and plasmin activity in goat milk. Master of Science Thesis (Life Science), China Agriculture University, Beijing, China, 2010.
- Scott MD, Woody P, Carol D, John B, Patrica AM, Dan S. Prevelance and incidence of subclinical mastitis in goats and dairy ewes in Vermount, USA. Small Rumen Res, 2002; 46(2-3):115-121.
- 9. Okada M. Histology of mammary glands. VII. Histological and histochemical studies of cells in the milk of domestic animals. Tohoku J Agric Res. 1960; 11:31-51.
- 10. Deutz A, Pernthaner A, Schlerka G, Barima W. Cell count of milk from sheep and goas and occurrence of bacterial mastitis in lower austraia. Wiener Tieraztliche monatsschrjit, 1990; 77(3):70-77.

- 11. Khodke MV, Bonde SW, Ambade RB. Alteration of enzyme aspartate transaminase in goat milk related to udder health status. Vet World, 2009; 2(1):24-26.
- 12. Kitchen BJ. Review of progress of dairy science: bovine mastitis: milk compositional changes and related diagnostic tests. J Dairy Sci, 1981; 48:167-188.
- 13. Kitchen BJ, Taylor GC, White IC. Milk enzyme and their distribution activity. J Dairy Sci. 1970; 37:279-288.
- 14. Vihan VS, Rubina R. Determination of lysosomal enzyme activity, somatic cell count, percent fat and protein in subclinical caprine mastitis. Somatic Cells and Milk of Small ruminants, Proceed Bella, Italy, Eaap publication, 1993; 1996:31-34.
- Horvath GA, Mohammed IH, Varga J, Szermeredi G, Quarini L. Effect of subclinical mastitis on milk composition. *Magyar Allatorvosok Lapja*, 1980; 35(9):615-619.
- Ali M, Hasan AK. Physical and chemical properties of goat milk. Mesopotammian J. Agricultur. 1988, 1998; 10(3):213-219.
- 17. Agnihotri MK, Rajkumar V. Effect on breed, parity and stage of lactation on milk composition of goats of goats in western region of India. Int J Dairy Sci. 2007; 2(2):172-177.
- Imran M, Khan H, Hassan SS, Khan R. Physic chemical characteristics of various milk samples available in Pakistan. J Zhejiang Univ Sci. 2008; 9(7):546-551.
- Juarez M, Ramos M. Physic-chemical characteristics of goat milk as distinct from those of cow milk. In International Dairy Federation (Ed.), Proceedings of the IDF Seminar Production and Utilization of ewe's and Goat's milk, Bulletin No. 202 Athens, Greece, 1986; 54-67.
- Schalm OW, Carroll ES, Jain NC. Bovine Mastitis 1st Edition, Lea and Febiger, Philadelphia, 1971.
- 21. Arora R, Bhojak N, Joshi R. Comparative aspects of goat and cow milk. Int J Eng. 2013; 2(1):07-10.
- 22. Hassan HJ. Variations in milk composition of some farm animals resulted by sub-clinical mastitis in aldiwania province. Onderstepoort J Vet Res. 2013; 12(2):17-24.
- 23. Getaneh G, Mebrat A, Wabie A, Kendie H. Review on goat milk composition and its nutritive value. J Nutr Health. 2016; 3(4):1-10.
- 24. Marechal L, Thiery Vautor, Lior L. Mastitis of milk and quality of milk products a review. Dairy Sci Technol, 2011; 91(3):247-282.