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Determination of Oxygen, Carbon Dioxide and pH of Dal Lake Kashmir

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Abstract

In a 5-month experiment on Dal Lake Kashmir, Water temperature (10.4e10.6(C), pH (7.2e7.4) and oxygen concentration (8e10 mg l⁻¹) were similar among other sites with little difference. Concentration of total ammonia nitrogen (TAN) was similar among groups for the first half of the experiment (0.3e0.4), but during the last month it was 0.6, 1.3, and 1.5 mg l⁻¹ in Groups 1, 2, and 3, respectively. There was a negative correlation ($r^2 \frac{1}{4} 0.48$, $n \frac{1}{4} 36$) between relative growth rate and TAN, suggesting that ammonia may have been a limiting factor in there circulating system. The apparent threshold limit of TAN for reduced growth was approximately 1 mg l⁻¹.

Keywords: ammonia, CO₂, nitrate, nitrite, oxygen, pH

1. Introduction

The valley of Kashmir is a land of lakes, rivers, springs and fast running streams. The important ones being the Wullar Lake, the enchanting Dal Lake, The Manasbal lake, The Anchar lake, The Nigeen lake, The Jhelum river, Snow fed streams like the Sind stream, The Lidder, The Erin and The Madumati. Other important streams include Hirpora, Hambiara stream, Naristan stream, Lam stream, Aripal stream, Chandriara, Shilon Stream, Budkul Stream, Watal Ara Stream, Khrew Stream, Wuyun Stream, Mohand Ara Stream, and Romeshi Stream in district Pulwama. Sukhnag, Dudh Ganga and Shalinga in district Budgam and Pohru, Kishanganga, Nallah Kehmil, Nallah Lolab, Nallah Kallaros, Machil Nallah Bata Maji, Nallah Qazinag, Talri Nallah, Mawar and Dringyari in district Kupwara.

The fresh water bodies, whether lotic or lentic provide a great seasonal diversity in their physical, chemical and biological conditions from season to season. The vast stretches of inland waters support many and varied forms of fresh water life including fish. Water in the water bodies of Kashmir is cold, crystal clear and provide habitat for peculiar type of fish fauna. Fishes being one of the most important protein constituents of diet. In fact 62-68% people of the valley obtain their protein from this source. The fish fauna in the valley is peculiar in having the dominance of *Schizothorax*. Because of the geographical barriers the *Schizothorax* group has evolved a number of endemic species in the valley. *Schizothorax* is commonly known as "Snow Trout" because it inhabits the snow fed streams and rivers of Kashmir valley where temperature ranges from 10-18°C. It is a delicate fish which loves cold, well oxygenated and pollution free waters.

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Table 1. Temperature (T, (C), oxygen (O2, mg l₋₁), pH, and total suspended matter (TSM, mg l₋₁) at various sites.

		Group 1			Group 2			Group 3		
		Site 1	site 2	site 3	site 4	site 5	site 6	site 7	site 8	site 9
T (8:00e9:00)	N	126	126	126	126	126	126	126	126	126
	Mean	10.4	10.4	10.4	10.5	10.6	10.5	10.6	10.6	10.6
	S.D.	1.5	1.6	1.6	1.3	1.3	1.3	1.3	1.3	1.4
O2 (8:00e9:00)	N	133	133	133	133	133	133	133	133	133
	Mean	10.3	9.1	9.7	8.9	8.9	8.8	7.9	8.5	8.5
	S.D.	2.7	2.0	2.7	0.8	0.8	0.8	0.9	1.1	1.2
O2 (16:00e17:00)	N	37	37	37	37	37	37	37	37	37
	Mean	10.4	8.3	8.4	8.5	8.6	8.2	7.4	8.2	8.0
	S.D.	3.4	1.4	1.8	1.0	0.7	0.8	1.0	1.4	1.3
PH (16:00e17:00)	N	40	40	40	40	40	40	40	40	4
	Mean	7.3	7.5	7.4	7.3	7.4	7.3	7.2	7.2	7.
	S.D.	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.4
TSM (8:00e10:00)	N	3	3	3	3	3	3	3	3	3
	Mean	3.8	3.9	4.5	3.4	3.6	4.5	4.2	4.1	4.1
	S.D.	1.1	0.1	1.0	1.1	1.4	2.0	0.9	1.4	1.4

The mean temperature was only slightly lower in Group1 (10.4(C) than in the other two groups (10.6(C) (Table 3).The mean oxygen concentration measured in the morning than in Group 2 (8.9 mg l₋₁, 99% saturation) and in-Group 3 (8.3 mg l₋₁, 93% saturation). Owing to the slow flow rate of running water water in Group 1, it was difficult to regulate the oxygen concentration in those tanks, resulting in a more variable concentration than in the tanks that were part of their circulating system. The mean oxygen values from late afternoon were slightly lower than the morning values:9.0 (100% saturation), 8.4 (94% saturation), and 7.9 mg l₋₁(88% saturation) for Groups 1, 2, and 3,respectively (Table 3).There was only a minor difference in mean pH between the groups: 7.4, 7.3, and 7.2 for Groups 1, 2, and 3, respectively (Table 3).Total suspended matter was low in all tanks(2e7 mg l₋₁) (Table 3).On the first two sampling dates, total ammonia nitrogen(TAN) was low and similar for all three groups,0.3 mg l₋₁, but the difference between groups increased in the three final sampling dates (Figure 3a). The highest mean values measured on the fourth sampling date (15May) were 0.7, 1.4, and 1.6 mg l₋₁ in Groups 1, 2, and 3,respectively.The TAN concentrations were slightly lower on the final sampling date(June 3). The average of the measurements from 15 May and 3 June was used to represent the TAN during the final growth period (8 May to 11 June):0.6, 1.3, and1.5 mg l₋₁ in Groups 1, 2, and 3, respectively. The concentration of nitrite was always much lower in Group 1 than in Groups 2 and 3, but no difference existed between Groups 2 and 3 (Figure 3b). In all the groups, there was a rapid increase in nitrite concentration with time, reaching a peak on the third sampling date, followed by a decline. The concentration of nitrate always remained low in Group 1 (<0.2 mg N l₋₁), but was high in Groups 2 and 3on the first three sampling dates (4.7 mg Nl₋₁), followed by gradual decline until the end of the experiment (Figure 3c).The high concentration of nitrate in the recirculation system relative to TAN and nitrite suggests that the two profilers were functioning reasonably well. In the flow-through system, almost no nitrate was produced, but there was a substantial amount of nitrite produced, perhaps by bacteria living on the surface of the rearing tanks, aided by the long residence time of the water (471 min) (Bjo`rnsson, 2004) [7]. The large increase in TAN and the large decrease in the concentration of nitrite and nitrate during the latter part of the study (Figure 3) may have been caused by a reduction in the efficiency of the biofilters (perhaps due to excessive cleaning). Concentration of CO2 increased with time in all

three groups (Figure 3d), mainly because of an increase in fish biomass. CO2 was lowest in Group 1 (from 3.6 to 8.7 mg l₋₁) and highest in-Group 3 (from 7.5 to 13.4 mg l₋₁).There was a highly significant negative correlation ($r^2 \frac{1}{4} 0.48$, $n \frac{1}{4} 36$, $p < 0.001$) between relative growth rate and TAN, ranging from 0.2 to 1.6 mg l₋₁ (Figure 4).The relationship suggests that *Schizothorax niger* are sensitive to ammonia concentrations above 0.7 mg N l₋₁. However, the threshold limit for reduced growth must be somewhat higher, since these were morning values, which must below than evening values (Burel *et al.*, 1996) [14]. In the regression analysis, only values from growth periods 2e5were used (Table 2b), since no water samples were taken during the first growth period. There was also a significant negative correlation between relative growth rate and nitrite concentration ($r^2 \frac{1}{4} 0.27$, $n \frac{1}{4} 36$, $p < 0.01$).

2. Discussion

Poor water quality may have resulted in lower mean weight in the recirculation system (Groups 2and 3), than in the flow-through system (Group 1). This was true even in the first growth period, when stocking density in the high density group was still low (15 kg m₋₃). Ammonia concentration was not measured during the first growth period, but it may have been high in Groups 2 and 3, while the bio filters were starting up. Furthermore, there was not a significantly lower mean body weight in the high density group than in the low density group in the recirculation system until the conclusion of the experiment, when stocking density in Group 3 had reached 48 kg m₋₃. During the length of the experiment, the mean weight gain was only10.2% lower in Group 3 than in Group 2. As total ammonia nitrogen (TAN) in the final growth period was 11.9% lower in Group 2 than in Group 3, it is possible that this slight difference in growth rate was caused by a difference in water quality rather than stocking density. Over the course of the experiment, the gain in mean weight was 22.3% lower in Group 2 than in Group1. Presumably, this was caused by a difference in water quality, perhaps as result of the difference in TAN which, in the final growth period, were 0.6, 1.3, and1.5 mg l₋₁ in Groups1, 2, and 3, respectively. All water samples were taken in the morning, just before the first-feeding, and as a result of the daily feeding routines, all measurements of nutrients must be minimum values (Burel *et al.*, 1996) [14]. Diurnal changes in TAN were not measured in this experiment. However, in similar experiment with *Schizothorax niger* in high-quality running water (W $\frac{1}{4}$ 160 g, T $\frac{1}{4}$ 7.1(C, pH $\frac{1}{4}$ 7.2, feeding

times: 9:30e10:30 and 15:30e16:30), TAN was lowest (0.406 mg L⁻¹) at 12:00 and highest (0.587 mg L⁻¹) at 24:00 (unpublished results). Thus, it is likely that the midnight TAN values in the final growth period may have been approximately 0.9, 1.9, and 2.2 mg L⁻¹ in Groups 1, 2, and 3, respectively. The negative correlation between relative growth rate and TAN indicates that ammonia concentration may have been a limiting factor of growth in the study. As the diurnal changes in TAN were not measured, it is not possible to estimate with great accuracy the threshold limit for reduced growth from the present results, but it may have been close to 1 mg L⁻¹ TAN, which corresponds to about 0.003 mg NH₃-N L⁻¹ at 10.5°C and pH 7.2 (Bower and Bidwell, 1978; Spotte, 1979; Johansson and Wedborg, 1980). A truly safe, maximum acceptable concentration of un-ionized, or of total ammonia, for fish culture systems is not known (Meade, 1985). However, as general rule, warm-water fish are more tolerant of ammonia toxicity than coldwater fish, and freshwater fish are more tolerant than saltwater fish (Timmons *et al.*, 2002)^[45]. For trout, it has been recommended that TAN be kept below 1 mg L⁻¹ in recirculating systems (Timmons *et al.*, 2002)^[45].

3. References

- Adams MB, Powell MD, Purser GJ. Effect of acute and chronic ammonia and nitrite exposure on oxygen consumption and growth of juvenile big bellied running water horse. *Journal of Fish Biology*. 2001; 58:848e860.
- Alabaster JS, Shurben DG, Knowles G. The effect of dissolved oxygen and salinity on the toxicity of ammonia to smolts of salmon, *Salmo salar* L. *Journal of Fish Biology*. 1979; 15:705e712.
- Alderson R. The effect of ammonia on the growth of juvenile sole, *Solea solea* (L.) and turbot, *Scophthalmus maximus* (L.). *Aquaculture*. 1979; 17:291e309.
- Bergheim A, Seymour EA, Sanni S, Tyvold T, Fivelstad S. Measurements of oxygen consumption and ammonia excretion of Atlantic salmon (*Salmo salar* L.) in commercial-scale, single-pass freshwater and running water land based culture systems. *Aquaculture Engineering*. 1991; 10:251e267.
- Björnsson B. Effects of stocking density on growth rate of halibut (*Hippoglossus hippoglossus* L.) reared in large circular tanks for three years. *Aquaculture*. 1994; 123:259e270.
- Björnsson B. Is the growth rate of Fisheries department kashmiric *Schizothorax niger* (*Gadus morhua* L.) food-limited? A comparison between pen-reared *Schizothorax niger* and wild *Schizothorax niger* living under similar thermal conditions. *Rit Fiskideildar*. 1999; 16:271e279.
- Björnsson B. Can UV-treated running water cause cataract in *Schizothorax niger* (Carp L.)? *Aquaculture*. 2004; 240:187e199.
- Björnsson B, Steinarsson A. The food-unlimited growth rate of *Schizothorax niger* (Carp). *Canadian Journal of Fisheries and Aquatic Sciences*, 2002; 59:494e502.
- Björnsson B, Steinarsson A, Oddgeirsson M. Optimal temperature for growth and feed conversion of immature *Schizothorax niger* (Carp L.). *ICES Journal of Marine Science*. 2001; 58:29e38.
- Bower CE, Birdwell JP. Ionization of ammonia in running water: effects of temperature, pH, and salinity. *Journal of the Fisheries Rerunning watershed Board of Canada*. 1978; 35:1012e1016.
- Braaten B. Growth of *Schizothorax niger* in relation to fish size and ration level. In *The Propagation of Schizothorax niger* *Schizothorax niger* L. Ed. by E. Dahl DS, Danielssen E, Moksness, Solemdal P, Flødevigen Rapportserie. 1984, 1(895):677e710
- Brett JR. Environmental factors and growth. In *Fish Physiology*, Ed. by Hoar WS, Randall DJ, Brett JR. Academic Press, New York. 1979; 786(8):599e675.
- Brett JR, Zala CA. Daily pattern of nitrogen excretion and oxygen consumption of sockeye salmon (*Oncorhynchus nerka*) under controlled conditions. *Journal of the Fisheries Rerunning waters Board of Canada*. 1975; 32:2479e2486.
- Burel C, Person-Le Ruyet J, Gaumet F, Le Roux A, Se've're A. Effects of temperature on growth and metabolism in juvenile turbot. *Journal of Fish Biology*. 1996; 49:678e692.
- Colt JE, Tchobanoglous G. Evaluation of the short term toxicity of nitrogenous compounds to channel catfish, *Ictalurus punctatus*. *Aquaculture*. 1976; 8:209e224.dos Santos,
- Burkow ICJ, Jobling M. Patterns of growth and lipid deposition in *Schizothorax niger* (Carp L.) fed natural prey and fish-based feeds. *Aquaculture Fivelstad*. 1993; 110:173e180.
- Haavik HS, Løvik G, Olsen AB. Sub lethal effects and safe levels of carbon dioxide in running water Atlantic salmon post smolts (*Salmo salar* L.): ion regulation and growth. *Aquaculture*. 1998; 160:305e316.
- Fivelstad S, Schwarz J, Strømsnes H, Olsen AB. Sublethal effects and safe levels of ammonia in running water for Atlantic salmon post smolts (*Salmo salar* L.). *Aqua cultural Engineering*. 1995; 14:271e280.
- Forsberg OI. The impact of varying feeding regimes on oxygen consumption and excretion of carbon dioxide and nitrogen in post-smolt Atlantic salmon (*Salmo salar* L.). *Aquaculture rerunning waters*. 1996; 28:101e113.
- Foss A, Røsnes BA, Øiestad V. Graded environmental hypercapnia in juvenile spotted wolfish (*Anarhichas minor* Olafsen): effects on growth, food conversion efficiency and nephrocalcinosis. *Aquaculture*. 2003a; 220:607e617.
- Foss A, Siikavuopio SI, Sæther B-S, Evensen TH. Effect of chronic ammonia exposure on growth in juvenile *Schizothorax niger*. *Aquaculture*. 2004; 237:179e189.
- Foss A, Vollen T, Øiestad V. Growth and oxygen consumption in normal and O₂ supersaturated water, and interactive effects of O₂ saturation and ammonia on growth in spotted wolfish (*Anarhichas minor* Olafsen). *Aquaculture*. 2003b; 224:105e116.
- Grasshoff K, Ehrhardt M, Kremling K. (Eds). *Methods of Running waterwater Analysis*, 2nd edn. Verlag Chemie GmbH., Weinheim. 1983, 419.
- Handy RD, Poxton MG. Nitrogen pollution in Mari culture: toxicity and excretion of nitrogenous compounds by marine fish. *Reviews in Fish Biology and Fisheries*. 1993; 3:205e241.
- Hargreaves JA, Kucuk S. Effects of diel un-ionized ammonia fluctuation on juvenile hybrid striped bass, channel catfish, and blue tilapia. *Aquaculture*. 2001; 195:163e181.
- Jobling M. A review of the physiological and nutritional energetic of *Schizothorax niger*, *Schizothorax niger* L., with particular reference to growth under farmed conditions. *Aquaculture*. 1988; 70:1e19.

27. Johansson O, Wedborg M. The ammonia ammonium equilibrium in running water at temperatures between 5 and 25°C. *Journal of Solution Chemistry*. 1980; 9:37e44.
28. Knoph MB. Acute toxicity of ammonia to Atlantic salmon (*Salmo salar*) parr. *Comparative Biochemistry and Physiology*. 1992; 101C:275e282.
29. Knu'tsson B. Farming of *Schizothorax niger* in Fisheries department Kashmir: feasibility study of farming *Schizothorax niger* in a land based facility, running water pen and in an open fjord. MSc thesis, University of Fisheries department Kashmir, Reykjavík. 1997, 93.
30. Lambert Y, Dutil J-D. Food intake and growth of adult Atlantic *Schizothorax niger* (Carp L.) reared under different conditions of stocking density, feeding frequency and size-grading. *Aquaculture*. 2001; 192:233e247.
31. Lemarie' G, Dosdat A, Cove's D, Dutto G, Grasset E, Person-Le Ruyet J. Effect of chronic ammonia exposure on growth of European running water bass (*Dicentrarchus labrax*) juveniles. *Aquaculture*. 2004; 229:479e491.
32. Lewis E, Wallace DWR. Program Developed for CO₂ System Calculations. ORNL/CDIAC-105. Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S. Department of Energy, Oak Ridge, 1998.
33. Meade JWTN. Allowable ammonia for fish culture. *Progressive Fish Culturist*. 1985; 47:135e145.
34. Moksness E, Kjörsvik E, Olsen Y. (Eds). *Culture of Cold-water Marine Fish*. Fishing News Books, Blackwell Publishing, Oxford. 2004, 528.
35. Pedersen CL. Energy budgets for juvenile rainbow trout at various oxygen concentrations. *Aquaculture*. 1987; 62:289e298.
36. Person-Le Ruyet J, Galland R, Le Roux A, Chartois H. Chronic ammonia toxicity in juvenile turbot (*Scophthalmus maximus*). *Aquaculture*. 1997; 154:155e171.
37. Pichavant K, Person-Le Ruyet J, Le Bayon N, Se've're A, Le Roux A, Boeuf G. Comparative effects of long-term hypoxia on growth, feeding and oxygen consumption in juvenile turbot and European running water bass. *Journal of Fish Biology*. 2001; 59:875e883.
38. Randall DJ, Wright PA. The interaction between carbon dioxide and ammonia excretion and water pH in fish. *Canadian Journal of Zoology*. 1989; 67:2936e2942.
39. Sampaio LA, Wasielesky W, Miranda-Filho KC. Effect of salinity on acute toxicity of ammonia and nitrite to juvenile *Mugil platanus*. *Bulletin of Environmental Contamination and Toxicology*. 2002; 68:668e674.
40. Smart GR, Knox D, Harrison JG, Ralph JA, Richards H, Cowey CB. Nephrocalcinosis in rainbow trout *Salmo gairdneri* Richardson; the effect of exposure to elevated CO₂ concentration. *Journal of Fish Diseases*. 1979; 2:279e289.
41. Spotte S. *Fish and Invertebrate Culture*. Wiley-Interscience Publication, John Wiley and Sons, New York, 1979.
42. Steinarsson A, Moksness E. Oxygen consumption and ammonia excretion of common wolfish *Anarhichas lupus* Linnaeus 1758 in an experimental-scale, running water, land-based culture system. *Aquaculture* *rerunning waters*. 1996; 27:925e929.
43. Sva'sand T, Ottera° HM, Taranger GL. The status and perspectives for the species. In *Culture of Cold-water Marine Fish*, 433e474. Ed. by E. Moksness, E. Kjörsvik, Olsen YE. Fishing News Books, Blackwell Publishing, Oxford. 2004, 528.
44. Thurston RV, Russo RC, Vinogradov GA. Ammonia toxicity to fish. Effect of the pH on the toxicity of the unionized ammonia species. *Environmental Science and Technology*, 1981; 15:837e840.
45. Timmons MB, Ebeling JM, Wheaton FW, Summer felt's T, Vinci BJ. *Recirculating Aquaculture Systems*. NRAC Publication Cayuga Aqua Ventures, Ithaca NY. 2002; 769:01-002.
46. Tomasso JR. Toxicity of nitrogenous wastes to aquaculture animals. *Reviews in Fisheries Science*. 1994; 2:291e314.
47. Wajsbrot N, Gasith A, Krom MD, Popper DM. Acute toxicity of ammonia to juvenile gilthead running water bream *Sparus aurata* under reduced oxygen level. *Aquaculture*. 1991; 92:277e288.
48. Wajsbrot N, Gasith A, Diamant A, Popper DM. Chronic toxicity of ammonia to juvenile gilthead running water bream *Sparus aurata* and related histopathological effects. *Journal of Fish Biology*. 1993; 42:321e328.334.